

PHYTOTOXIC EFFECT OF EUGENOL TOWARDS TWO WEEDY SPECIES

S. VAID*, DAIZY R. BATISH, H. P. SINGH¹ AND R. K. KOHLI

Department of Botany, Panjab University, Chandigarh - 160 014

¹Department of Environment and Vocational Studies,

Panjab University, Chandigarh - 160 014

E-mail: dr.supriyavid@yahoo.com

KEY WORDS

Allelopathy
Cassia occidentalis
Bidens pilosa Volatile
monoterpene
Weed suppression

Received on :

30.04.2010

Accepted on :

21.08.2010

*Corresponding
author

ABSTRACT

A study was undertaken to assess the inhibitory / allelopathic potential of eugenol—a volatile monoterpene found in *Eugenia* spp. and several other aromatic plants, against two weedy species viz. *Cassia occidentalis* and *Bidens pilosa*. Eugenol significantly inhibited the germination of both the weeds even at very low concentration. However, the effect was more on *B. pilosa* compared to *C. occidentalis*. Likewise, the seedling growth measured in terms of seedling length and dry weight of both the test weeds was appreciably reduced in response to the test monoterpene. Not only the growth, even the content of the total chlorophyll and cellular respiration in both the test weeds were appreciably reduced, thereby indicating that eugenol negatively affects the photosynthetic efficiency and the energy metabolism of the weed species. Based on the study, it is concluded that eugenol possesses weed-suppressing ability and can be used for future weed management programmes either directly or by serving as a lead molecule.

INTRODUCTION

Weeds—the essential component of agroecosystems, interfere with crops and lead to enormous crop losses. Indiscriminate use of synthetic herbicides though has undoubtedly enhanced the much needed crop production, yet have led to a number of toxicological and environmental problems causing ill effects on human health (Macias *et al.*, 2001). Therefore, efforts are being made world over to search for safer and environmentally benign chemicals that have relatively shorter half-life. In this direction, biologically active natural products such as allelochemicals are fast being tested for weed management since they are environment friendly and have different modes of action (Dayan *et al.*, 2000). Besides, they also exhibit immense diversity and biological activity.

Among the natural plant products, volatile terpenes have been shown to be promising with potential weed suppressing ability (Kohli *et al.*, 1998; Singh *et al.*, 2002). These are commonly found as components of essential oils in a number of aromatic plants, e.g. *Salvia* spp, *Eucalyptus* spp., *Artemisia* spp. and *Pinus* spp. Besides, they also exhibit considerable phytotoxicity, e.g. volatile terpenes of *Salvia leucophylla* are most effective in inhibiting the growth of grasses (Muller *et al.*, 1964); *Eucalyptus* volatile terpenes reduce growth of a number of plants (Kohli and Singh, 1991); cineoles (1, 4- and 1, 8-) reduce the growth of weeds (Romagni *et al.*, 2000). Eugenol is one such monoterpene, which is a major component of essential oils from a number of aromatic species including *Eugenia* spp. and is biologically very active (Tworkoski, 2002).

But very little has been done to determine its allelopathic activity against weedy species. With a view to explore its herbicidal potential against a wide range of weeds, the present investigation was, therefore, undertaken to explore the phytotoxic effect of eugenol against two weedy species viz. *Cassia occidentalis* and *Bidens pilosa*. The objective of this study, therefore, is to explore the possible utility of eugenol for the management of obnoxious weeds and to determine its effect on physiological changes in plants.

MATERIALS AND METHODS

Collection of Material

Seeds of coffee weed (*Cassia occidentalis* L.) and hairy beggarticks (*Bidens pilosa* L.) were collected locally from wildy growing stands in the campus of Panjab University, Chandigarh. These were surface sterilized. Eugenol was purchased from Lancaster, UK.

Bioassay Studies

Seeds of both the weed species were divided into 9 groups of 50 each and dipped in distilled water for 16 hrs for imbibition prior to germination trial. These were then equidistantly placed in 6" diameter Petri dishes lined with two layers of moistened Whatman no. 1 filter paper. The filter paper was treated with 1, 2, 5, 10 and 20 μ L of eugenol per Petri dish. After the addition of the volatile terpenes, the Petri dishes were sealed. A similar set-up but without eugenol served as control. For each treatment 5 replicates were maintained. The entire set up was kept in an environmentally controlled seed

germinating chamber at 25 ± 2 °C and 75 ± 2 % relative humidity with a photoperiod of 16 / 8 hour day / night. After a week, seedling length and seedling dry weight were measured and the total chlorophyll content and respiratory activity were determined.

Estimations

Chlorophyll was extracted from 25 mg of tissue in 4 mL of dimethyl sulphoxide (DMSO) following Hiscox and Israelstam (1979). Its concentration was determined spectrophotometrically (Arnon, 1949) and the amount was expressed in terms of dry weight as suggested by Rani and Kohli (1991). Respiratory values were determined indirectly using 2, 3, 5 - triphenyl tetrazolium chloride as per the method of Steponkus and Lanphear (1967).

Statistical analysis

The data of percent germination, seedling growth, chlorophyll content and respiratory activity were analyzed by one-way ANOVA followed by Duncan's multiple range test.

RESULTS AND DISCUSSION

It is clear from the results that in response to different concentrations of eugenol, germination of both the weedy species was reduced (Fig. 1). Reduction in germination was more in case of *B. pilosa* compared to *C. occidentalis*. A complete inhibition of germination of *B. pilosa* was observed at $5\mu\text{L}$ concentration whereas in case of *C. occidentalis* it was seen at $20\mu\text{L}$ (Fig. 1).

Further, seedling length and seedling dry weight of both the weeds were significantly reduced compared to control. Growth of *B. pilosa* was more affected than *C. occidentalis* (Table 1). Inhibition of germination and growth of both the weedy species could be due to disruption of mitotic activity in the germinating seeds. Though, present study does not have supporting data in this direction, yet references available in literature can strengthen this fact (Romagni *et al.*, 2000). Several reasons have been put forward to find out the factors that disrupt mitosis

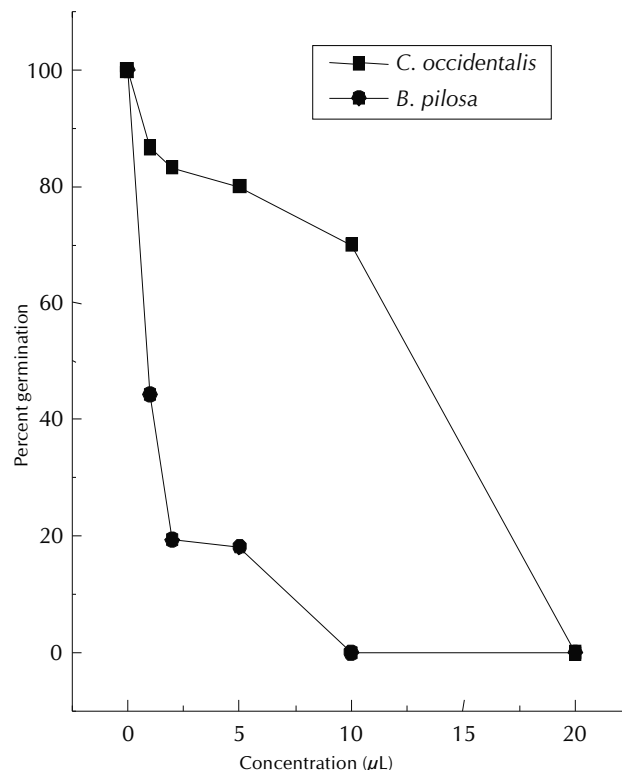


Figure 1: Effect of Eugenol on the germination of two test weed species

such as disruption of microtubule organization or alternation of cell wall biosynthesis (Lehnen and Vaughn, 1992).

The content of chlorophyll extracted in DMSO was significantly less in treated weed species compared to control. More reduction was observed in *B. pilosa* (Table 2). The decrease in chlorophyll suggests the diminishing photosynthesis efficiency in response to volatile terpenes. The mechanism as to how chlorophyll decreases in the target weed or whether it is due to decreased synthesis or enhanced

Table 1: Effect of Eugenol on the seedling length and seedling dry weight of *C. occidentalis* and *B. pilosa*

Concentration (µL)	Seedling length (cm)		Seedling Weight (mg)	
	<i>C. occidentalis</i>	<i>B. pilosa</i>	<i>C. occidentalis</i>	<i>B. pilosa</i>
0	9.33 ± 0.75 ^a	7.26 ± 1.09 ^a	10.4 ± 0.50 ^a	0.64 ± 0.05 ^a
1	6.38 ± 0.43 ^b	2.13 ± 0.48 ^b	7.70 ± 0.34 ^b	0.53 ± 0.05 ^b
2	5.83 ± 0.79 ^c	1.35 ± 0.30 ^c	7.13 ± 0.29 ^b ^c	0.49 ± 0.07 ^b ^c
5	3.94 ± 0.60 ^d	0.82 ± 0.25 ^d	6.73 ± 0.48 ^c	0.46 ± 0.03 ^c
10	2.06 ± 0.29 ^e	-	4.07 ± 0.37 ^d	-
20	-	-	-	-

Different superscripts in a column represent significant difference at $p < 0.05$.

Table 2: Effect of Eugenol on the total chlorophyll content and cellular respiration of *C. occidentalis* and *B. pilosa*

Concentration (µL)	Total Chlorophyll content (µg/mg)		Cellular Respiration (%)	
	<i>C. occidentalis</i>	<i>B. pilosa</i>	<i>C. occidentalis</i>	<i>B. pilosa</i>
0	8.75 ± 0.38 ^a	6.45 ± 0.07 ^a	100 ^a	100 ^a
1	4.40 ± 0.15 ^b	5.80 ± 0.42 ^b	47.3 ^b	36.1 ^b
2	3.96 ± 0.27 ^c	4.03 ± 0.05 ^c	45.3 ^b	29.0 ^c
5	3.40 ± 0.04 ^d	1.14 ± 0.8 ^d	40.4 ^c	13.2 ^d
10	0.16 ± 0.09 ^e	-	24.5 ^d	-
20	-	-	-	-

Different superscripts in a column represent significant difference at $p < 0.05$.

degradation could not be known. Nevertheless, there are available references indicating reduced levels of chlorophyll pigment in response to allelopathy / allelochemicals (Romagni *et al.*, 2000; Singh *et al.*, 2002).

The study on cellular respiration is an indirect method indicating rate at which O₂ molecule released through respiratory chain are trapped by 2, 3, 5 - triphenyl tetrazolium chloride (TTC) leading to the formation of water insoluble red formazan (Steponkus and Lanphear, 1967). The intensity of this red coloured formazan extracted in alcohol is determined. The present study reveals a considerable and appreciable reduction in cellular respiration in both the weed species when treated with eugenol (Table 2).

CONCLUSION

From the present study, it is clear that eugenol has a potential to reduce the growth and development of weed species and thus could be useful for future weed management programmes either directly or by serving as a the lead molecule.

REFERENCES

- Arnon, D. I. 1949.** Copper enzymes in isolated chloroplasts: Polyphenol oxidase in *Beta vulgaris*. *Pl. Physiol.* **24**: 1-15.
- Dayan, F. E., Romagni, J. G. and Duke, S. O. 2000.** Investigating the modes of action of natural phytotoxins. *J. Chem. Ecol.* **26**: 2079-2094.
- Hiscox, T. D. and Israelstam, G. F. 1979.** A method for extraction of chlorophyll from leaf tissue without maceration. *Can. J. Bot.* **57**: 1332-1334.
- Kohli, R. K. and Singh, D. 1991.** Allelopathic impact of volatile components from Eucalyptus on crop plants. *Biol. Plant.* **33**: 475-483.
- Kohli, R. K., Batish, D. R. and Singh, H. P. 1998.** Eucalypt oils for the control of parthenium (*Parthenium hysterophorus* L.). *Crop Prot.* **17**: 119-122.
- Lehnen, L. P., Jr. and Vaughn, K. C. 1992.** The herbicide sindone B disrupts spindle microtubule organising centres. *Pestic. Biochem. Physiol.* **44**: 50-59.
- Macias, F. A., Galindo, J. C. G., Varela, R. M., Simonet, A. M. and Castellano, D. 2001.** The use of allelopathic, studies in the search for natural herbicides. *J. Crop. Prod.* **4(2)**: 237-255.
- Muller, C. H., Muller, W. H. and Haines, B. L. 1964.** Volatile growth inhibitors produced by aromatic shrubs. *Science.* **143**: 471-473.
- Rani, D. and Kohli, R. K. 1991.** Fresh matter is not an appropriate unit for chlorophyll content: experiences from experiments on effects of herbicides and allelopathic substances. *Photosynthetica.* **25**: 655-658
- Romagni, J. G., Allen, S. N. and Dayan, F. E. 2000.** Allelopathic effects of volatile cineoles on two weedy plant species. *J. Chem. Ecol.* **26**: 303-313.
- Singh, H. P., Batish, D. R., Kaur, S., Ramezani, H. and Kohli, R. K. 2002.** Comparative phytotoxicity of four monoterpenes against *Cassia occidentalis*. *Ann. Appl. Biol.* **141**: 111-116.
- Steponkus, P. L. and Lanphear, F. R. 1967.** Refinement of triphenyl tetrazolium chloride method of determining cold injury. *Pl. Physiol.* **42**: 1423-1426.
- Tworokski, T. 2002.** Herbicide effects of essential oils. *Weed Sci.* **50**: 425-431.

INSTRUCTION TO AUTHORS

The Bioscan

An International Quarterly Journal of Life Science

THE JOURNAL

The Bioscan is an international quarterly journal of life sciences with international editorial board. The journal is online and details can be seen (downloaded from the site. www.thebioscan.in). For any query e-mail at m_psinha@yahoo.com & dr.mp.sinha@gmail.com can be used.

AIM & SCOPE

The journal aims to publish original peerly reviewed/ refereed research papers/reviews on all aspects of life sciences.

SUBMISSION OF MANUSCRIPT

Only original research papers are considered for publication. The authors may be asked to declare that the manuscript has not been submitted to any other journal for consideration at the same time. Two hard copies of manuscript and one soft copy, complete in all respects should be submitted. The soft copy can also be sent by e-mail as an attachment file for quick processing of the paper.

FORMAT OF MANUSCRIPT

All manuscripts must be written in English and should be typed double-spaced with wide margins on all sides of good quality A4 paper.

First page of the paper should be headed with the title page, (in capital, font size 16), the names of the authors (in capitals, font size 12) and full address of the institution where the work was carried out including e-mail address. A short running title should be given at the end of the title page and 3-5 key words or phrases for indexing.

The main portion of the paper should be divided into Abstract, Introduction, Materials and Methods, Results, Discussion (or result and discussion together), Acknowledgements (if any) References and legends.

Abstract should be limited to 200 words and convey the main points of the paper-outline, results and conclusion or the significance of the results.

Introduction should give the reasons for doing the work. Detailed review of the literature is not necessary. The introduction should preferably conclude with a final paragraph stating concisely and clearly the aims and objectives of your investigation.

Materials and Methods should include a brief technical description of the methodology adopted while a detailed description is required if the methods are new.

Results should contain observations on experiment done illustrated by tables and figures. Use well known statistical tests in preference to obscure ones.

Discussion must not recapitulate results but should relate the author's experiments to other work on the subject and give their conclusions.

All tables and figures must be cited sequentially in the text. Figures should be abbreviated to Fig., except in the beginning of a sentence when the word Figure should be written out in full.

The figures should be drawn on a good quality tracing/ white paper with black ink with the legends provided on a separate sheet. Photographs should be black and white on a glossy sheet with sufficient contrast.

References should be kept to a minimum and listed in alphabetical order. Personal communication and unpublished data should not be included in the reference list. Unpublished papers accepted for publication may be included in the list by designating the journal followed by "in press" in parentheses in the reference list. The list of reference at the end of the text should be in the following format.

1. **Witkamp, M. and Olson, J. S. 1963.** Breakdown of confined and non-confined Oak Litter. *Oikos*. **14**:138-147.
2. **Odum, E.P. 1971.** *Fundamentals of Ecology*. W. B. Sauder Co. Publ. Philadelphia.p.28.
3. **Macfadyen, A.1963.** The contribution of microfauna to total soil metabolism. In:*Soil organism*, J. Doeksen and J. Van Der Drift (Eds). North Holland Publ. Comp., pp 3-16.

References in the text should be quoted by the **author's name and year** in parenthesis and presented in year order. When there are more than two authors the reference should be quoted as: first author followed by *et al.*, throughout the text. Where more than one paper with the same senior author has appeared in on year the references should

Cont. P. 354