# STABILITY STUDY ON BASAL AND NON-BASAL BRANCHING GENOTYPES OF INDIAN MUSTARD (*Brassica juncea* L. Czern & Coss) UNDER DIFFERENT MOISTURE REGIMES

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# **ABSTRACT**

The present study was undertaken with 20 genotypes, under four environments conducted in Randomized Complete Block Design in 3 replications and evaluated 33 morpho-physio-biochemical traits suggested that significant G-E interaction including environment (linear), linear component of G-E interaction and pooled deviation (non-linear) were indicating considerable genetic variability for most of the studied traits. Role of the environmental variation has no influence on siliqua density (0.32), length of siliqua (1.50), chlorophyll content (0.49), proline (0.17), 1000 seed weight (1.51) and oil content (1.97). Out of eleven stable genotypes, seven (NRCDR-2, TM-151, Kranti, PKRS-28, TM-128, PM-28 and RAURD-78) in poor ( $\overline{\chi} > \mu$ , bi < 1, NS S2di), two

 $\overline{\chi} > \mu$ , bi > 1, NS S2di) environments. For none of the 11 stable genotypes days to first flower open reflected stability and needs attention for ideotype development except RH-8814 (= 41.42, bi = 1.69, S2di = 0.42) in rich environments. The major outcome of research is that NRCDR-2 is the most stable genotype in poor whereas popular variety Rajendra Suphlam stable in average environments can be suggested to farmers and also yield

(Rajendra Suphlam and KMR 10-2) in average ( $\overline{\chi} > \mu$ , bi = 1, NS S2di) and two (Rohini and RH8814) in rich (

# determinants RV, RL and HFPB are most important for residual moisture – rainfed environments.

# **INTRODUCTION**

The oilseeds scenario in the country had undergone a sea change in the last fifteen years while India changed from importer in the 1980s to a net exporter status during the early 1990s also termed as yellow revolution phase. Again, it has come back to net importer status importing more than 40 percent of its annual edible oil seeds which is mainly attributed to crop Brassicas because of low productivity of oilseed crops and year to year fluctuations in production in India which could be attributed to about 8.5 percent of the area under oilseeds is rainfed comprising mostly marginal and sub marginal lands with soils of poor fertility and also cultivated under resource constrained condition. Mustard seeds contain about 38-42 % oil, which is golden yellow, fragrant and considered among the healthiest and most nutritional cooking medium (Shekhawat et al. 2014).

Water stress is important abiotic factor (Campbell et al., 1992). The effect of drought stress is a function of genotype, intensity and duration of stress, weather conditions, growth, and developmental stages of rapeseed (Robertson and Holland, 2001). For getting higher yield of mustard, irrigation and fertilizermanagement are two important agronomic practices (Ray et al., 2014). The stable performance of genotypes for different plant characteristics, besides high yield, is very much desired for their commercial exploitation. However, the sensitive behavior of the existing varieties of mustard to different

growing environments (critical and normal) leads to fluctuations in its yield. The differential performance of the genotypes in varied agro-climatic conditions is due to a significant effect of genotype x environment (G x E) interactions.

For assessing the genetic worth and stability of basal and non-basal branching genotypes for yield and its components in Indian mustard, the present study with objective that outsized acreage of Indian mustard under conserved residual moisture – rainfed condition on farmers' fields, so keeping this in view it is proposed to study the stability of promising basal and non-basal branching genotypes for grain yield and morphological yield determinants and to select the stable genotypes (basal and non-basal branching type) for yield and yield attributing traits exhibiting good performance under different water regimes.

# **MATERIALS AND METHODS**

The experiment was laid out in Randomized Complete Block Design (RCBD) with three replications including check namely, Varuna (National Check) consisting of 20 genotypes for stability study, received from different All India Co-ordinated Research Project-Rapeseed and Mustard centres. During Rabi 2016-17, along with stability study, same set of entries (including check) under stability study were also subjected for evaluation of the physiological parameters by standardized

protocols (at 75DAS).

During Rabi 2016-17 at Crop Experiment Farm, Pusa the stability trial was sown under residual moisture only (E1) inside the Rain-out shelter which strictly avoids moisture condition makes difference from rainfed (E2) condition (which supposed to receive rainfall during crop phenological stages). whereas, irrigation at pre flowering stage (E3 at 45DAS) and in E4, one irrigation at pre flowering (45DAS) stage and second at pod formation stage (65DAS).

Basal branching genotypes are those having initiation of branch from base of plant up to 30 cm (Kumar et al., 1996), medium basal branching genotypes, branch arises from more than 30 cm up to 60 cm but in high basal branching genotypes, it ranges from more than 60 cm up to 93 cm.

Investigation was conducted in the field, laboratory and subsequently observations were recorded for stability study, namely days to first flower open (DFFO), days to physiological maturity (DPM), Days to cessation of flowering (DCF), Primary branches plant-1 (PBP-1), Secondary branches plant-1 (SBP-1), Length of siliqua (LS), Number of siliqua on primary mother axis (SPMA), Length of Primary Mother Axis(LPMA), Siliqua density (SD), Height of first primary branch (HFPB), Number of seeds siliqua-1 (SS-1), Root volume (RV), Root length (RL), Root Mass (RM), 1000 seed weight (TSW), Biological yield (BY), Harvest index (HI), Oil content (OC), Dry matter efficiency (DME), Relative Water Content (RWC), Leaf Membrane Stability Index (LMSI), Excised Leaf Water Loss (ELWL), Chlorophyll Content (CC), Catalase Activity (CA), Peroxidase Activity (PERO), Proline Accumulation (PRO), Relative Growth Rate (RGR), Leaf Area Index (LAI), Specific Leaf Weight (SLW), Drought Tolerance Index (DTI), Stress Intensity (SI), Oil Yield (kg/ha) (OY), and Grain Yield/Plot (kg/ha) (GY/P).

# **Relative Water Content (%)**

The relative water content was determined in fresh 4-5 leaf discs of 2 cm. diameter, excluding midrib. Discs were weighed quickly and immediately floated on double distilled water, in Petri dishes to saturate them with water for the next 24 hrs., in dark. The adhering water of the discs was blotted and turgor weight was taken. Dry mass of these discs was obtained, after dehydrating them at 70 °C for 48 hrs. Relative water content was calculated by placing the observed values in the following formula (Barr and Weatherley, 1962) in Ricinus communis  $RWC = \frac{Fresh \, mass - Dry \, mass}{Turgor \, mass - Dry \, mass} X100$ 

# Leaf Area Index (m2 / m2)

For determining LAI, SYSTRONIC LEAF AREA METER 211 was used during measurement of length and width of leaf. Leaf area of one leaf was calculated by multiplying by total number of leaves present in a unit area to obtain total leaf area and was

divided by unit ground area to get leaf area index. The leaf area index was calculated according to the formula given by Watson (1947) as mentioned below:

$$LAI = \frac{Leaf Area}{Ground Area}$$

# Relative Growth Rate (g g -1day-1)

Five Plants were uprooted at 35 and 65 days during their growth period. Uprooted plants were kept inside the oven and dried under 40°C for 72 hrs. Dried plants then weighed under electronic balance. The RGR was calculated by formula given by Williams (1946).

$$RGR = \frac{Loge\ W2 - Loge\ W1}{t2 - t1}$$

Where.

W1 and W2 are whole plant dry weight at t1 and t2 respectively; t1 and t2 are time interval in days

# Specific Leaf Weight (gcm-2)

Leaves from five plants were weighed under electronic balance and then area of the weighed leaves was taken by SYSTRONIC LEAF AREA METER 211. The SLW was calculated by formula given by Pearce *et al.* (1968) in alfalfa.

$$SLW = \frac{Leaf \ weight}{Leaf \ area}$$

# Leaf membrane stability index (%)

800g of leaves were put in 40 ml of distilled water inside test tube in two different sets. One set was kept at 40°C for half an hour and another set at 100°C for 15 minutes under water bath. The conductivity was measured for both the sets with the help of Conductivity Meter. The LMSI was calculated by formula given by Premchandra et al. (1990), as modified by Sairam (1994) in wheat.

$$LMSI = (1-C1/C2) \times 100$$

C1 = conductivity measured for sample at 40°C for half an hour under water bath.

C2 = conductivity measured for sample at °C for 15 minutes under water bath.

# Excised leaf water loss (%)

The fresh weight of five leaves from each plant were weighed and incubated inside incubator for 6 hrs at 28 °C and then weighed. The leaves were again kept for 24 hrs at 70°C and weighed. The ELWL was calculated by formula given by Malik, 1995 in wheat.

$$ELWL = \frac{Fresh \text{ weight - Weight after 6 hrs at 28}^{\circ}\text{C under incubator}}{Fresh \text{ weight - Dry weight after 24 hrs at 70}^{\circ} \text{ under oven}} X100$$

# **Drought tolerance index (%)**

In each replication, seed yield of five plants of all genotypes

were calculated. DTI can be calculated as mentioned below by Fischer and Maurer (1978) in wheat.

 $S = \frac{(1-Y/Yp)}{(1-X/Xp)}$ 

Y = Mean seed yield of a genotype in a stress environment

Yp= Mean seed yield of a genotype in a stress free environment

X = Mean seed yield of all genotype in a stress environment

Xp = Mean seed yield of all genotype in a stress free environment

### Stress intensity (%)

It is also known as Yield Stability Ratio (YS). SI can be calculated as mention below by Lewis (1954):

 $SI = (1 - Ys/Yn) \times 100$ 

Ys = Yield under stress

Yn = Yield under normal condition

# Harvest index (%)

It is the ratio of economic yield to the biological yield in per cent. Harvest index was calculated by following formula (Donald and Hamblin, 1976, Cereals).

$$H.I = \frac{Economic\ yield}{Biological\ yield}X100$$

Here.

Economic yield = Grain yield (g)

Biological yield = Total plant yield (g)

# Proline Accumulation in Leaves (µg/g Dry Wt.)

For the quantification of proline the method developed by

Bates et al. (1973) in soyabean and sorghum was used.

# Peroxidase Activity in Leaves (unit /g Fresh Wt.)

The activity of peroxidase was determined by the method of Palmiano and Juliano (1973) in rice.

# Catalase activity (unit/g Fresh Wt.)

The activity of Catalase was determined by the method given by Euler and Josephson (1927).

# RESULTS AND DISCUSSION

The principle objective of mustard breeding programme aims at improvement in yield, reliability in performance, stability and adaptation over a wide range of environments. Jinks and Jones, 1958, were of the view that degree of yield potential was the result from the combination of the range of gene effects. The knowledge of genotype – environment interaction gives a better appreciation of the measure of adaptability of the variety, a better means of studying individual plasticity and better insight of physiological and developmental process and gene action. The  $G \times E$  interaction has been shown to often reduce progress for selection (Comstock and Moll, 1963). Yield data and stability performance of genotypes across contrasting environments are essential to enable a breeder to select high yield and consistently performing genotypes.

Pooled analysis of variance (table 1) for different studied characters between the genotypes exhibited significant variation for most of the characters except for SD, LS, SPMA, SS-1, RM, LAI, CC, DTI, SI, PRO, TSW and OC which indicated genotypic variation in the studied genotype namely, Rajendra Suphlam, Pusa Mahak, KMR-10-1, Maya, Rohini, Kranti, TM151 developed at various AICRIP R and M centres located in mustard growing states across India(Schuster and Sra,1979). Environmental variation was observed for all the characters

Table 1: Analysis of variance for thirty - three morpho-physio-biochemical and quality traits used for stability studies in Indian mustard

Source	DF	Mean squares Days to first flower open	Days to physiological maturity	Days to cessation of flowering	Height of first primary branch	Primary branches per plant	Secondary branches per plant	Number of siliqua on Primary Mother Axis	Length of Primary Mother Axis	Siliqua density	Length of siliqua	Number of seeds siliqua-1
Genotype	19	47.50**	19.40**	15.51**	586.09**	2.26**	45.64**	72.94**	92.08**	0.13	0.47	1.49
Environment	3	1.47	24.45**	114.59**	670.39**	8.02**	123.79**	467.04**	155.51**	0.32	1.5	3.84*
G×E	57	6.27**	1.75**	5.18**	50.12**	0.45	6.95**	24.12**	25.15**	0.093	0.19	1.32
E(Linear)	1	4.43*	73.37**	343.78**	2011.19**	24.06**	371.39**	1401.12**	466.53**	0.97	4.51*	11.52**
G×E(Linear)	19	7.57**	0.79	6.60**	55.17**	0.37	12.34**	36.52**	50.36**	0.093	0.24	1.78*
Pooled deviation	40	5.34**	2.12**	4.25**	45.21**	0.47	4.05**	17.02**	11.92**	0.089	0.165	1.044
Pooled Error	152	0.43	1.38	2.38	1.41	0.045	0.11	0.37	2.2	0.001	0.012	0.065

Table 1 : Continued

Source	DF	Mean squ	ares								
		Tap root length	Root volume	Root mass	Relative growth rate	Leaf area index	Specific leaf weight	Chloro phyll content	Leaf membrane stability index	Relative water content	Excised leaf water loss
Genotype	19	14.70**	136.22**	1.84	67.95**	1.027	10.90**	0.031	93.94**	233.02**	88.96**
Environment	3	12.80**	18.36**	10.69**	390.67**	9.70**	56.26**	0.49	890.46**	9074.78**	380.16**
G×E	57	1.97**	1.24	0.44	6.48**	0.1	1.18	0.006	12.37**	28.89**	10.52**
E(Linear)	1	38.42**	55.08**	32.09**	1172.01**	*29.10**	168.80**	1.47	2671.39**	27224.36**	1140.50**
G×E (Linear)	19	1.77*	2.52**	0.82	9.15**	0.223	2.090 **	0.014	22.44**	55.39**	19.31**
Pooled deviation	40	1.97**	0.57	0.24	4.88**	0.0432	0.69	0.0018	6.97**	14.87**	5.82**
Pooled Error	152	0.058	0.104	0.0049	0.85	0.0301	0.048	0.0004	0.24	0.49	0.22

Table 1: Continued

Source	DF	Mean squares Catalase activity	Peroxidase activity	Proline accumu lation	1000 seed weight	Biological yield	Harvest index	Dry matter efficiency	Oil content	Grain yield ha <sup>-1</sup>	Oil yield ha- <sup>1</sup>
Genotype	19	2313.01**	1478.76**	0.026	1.098	201132.35**	18.43**	12.79**	0.52	121223.61**	18294.79**
Environment	3	4960.24**	346.21**	0.17	1.51	2022045.00**	42.6297**	26.4670**	1.9747	2904124.0000**	416008.9688**
G×E	57	266.31**	176.70**	0.0026	0.19	39335.77**	6.65**	4.59**	0.1001	12159.40**	1843.69**
E(Linear)	1	14880.72**	1038.63**	0.51	4.53*	6066135.00**	127.88**	79.40**	5.92*	8712372.00**	1248026.87**
G×E(Linear)	19	186.84**	126.68**	0.004	0.3	57685.87**	4.59**	3.44**	0.0782	26873.95**	4103.01**
Pooled deviation	40	290.73**	191.63**	0.0017	0.13	28652.69**	7.29**	4.90**	0.1055	4562.02**	678.33**
Pooled Error	152	1.8	5.35	0.0001	0.091	14327.93	1.98	1.4	0.0875	13039.2	1978.63

Table 1: Continued

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Source	DF	Mean square Stress intensity	es Drought tolerance index
Genotype	19	0.0104	0.1962
Environment	2	0.4355	15.9056**
G×E	38	0.0037	0.074
E(Linear)	19	0.8709	31.8112**
G×E(Linear)	20	0.0058	0.1148
Pooled deviation	114	0.0015	0.0316
Pooled Error	59	0.0045	0.0741

<sup>\*</sup>Significant at P = 0.05 \*\* Significant at P = 0.01

(significant mean squares due to environments) indicated environmental conditions were fluctuating and variable except for SD, LS, SS-1, LAI, CC, SI, PRO, TSW and OC reflected that these traits are unaffected and their expression in response to over or across the environment remains same (Gazal et al., 2013). Thomas et al. (1971); Baker (1969) and Byth et al. (1976) and Sagolsem et al. (2013) reported except for 1000 seed weight that significant  $G \times E$  interaction including Environment (linear) effects for most of the characters except for SD, CC, SI, PRO, TSW and OC revealed that average performance of genotype with respect to yield and other characters varied significantly. The linear components of G × E interaction was found highly significant for morphophysio-biochemical and quality traits, indicating that these characters were unstable and fluctuated in their expression with change in environment (Langer et al.,1979; Powel et al.,1986 and Ramagosa and Fox,1993). Moreover, predominance was observed from 12 traits out of 33 traits which might be responsible for adaptation in Indian mustard under moisture stress. Pooled deviation (non-linear) was significant for 19 traits out of 33 traits indicating considerable genetic diversity in the material. The highest pooled deviation (non-linear) was observed for traits namely, HFPB, SPMA, LPMA, RWC, PERO, BY, GY and OY which might be of practical value, to construct and test unity of multiple regression model to know more critically complex mechanism of adaptation (Langer et al., 1979; Chaudhary et al., 2004; Abou El-Nasr et al., 2006 and Sah et al., 2015).

The significance of genotype and G-E (linear) component emphasize that genotypes deviating slightly from the regression line of unit slope could be identified. Accordingly, three kinds of linear responses, namely bi = 1; bi = >1 and bi = <1 have been generally observed. However, in the present study negative bi (table 2) value were observed in characters namely, DFFO, DCF, HFPB, SBP-1, SPMA, LPMA, SD, LS, SS-1, RL, RV, RM, ELWL, PERO, TSW, HI and DME. Such type of linear

response could be attributed due to inadequacy of the scale use for the analysis and / or the inherent behaviour of the genotypes investigated (Knight, 1970).

DFFO, LPMA, SD, SS-1, RL, PERO, PRO, HI and DME showed variable range of bi value indicated different environmental responses in the studied genotypes and visualized the environmental ability to a large extent. It also suggested the possibility of selection for specific genotype patterns (Pfakler and Linksen, 1979 and Abou-El- Nasar et al., 2006).

Amongst 20 genotypes (table 4), 11 were stable in poor environment ( $\overline{X} > \mu$ , bi < 1,NS S2di), 2 in average environment ( $>\mu$  bi = 1,NS S2di) and rest 2 in rich environment ( $>\mu$ bi > 1,NS S2di) for grain yield ha<sup>-1</sup>. All the stable 11 genotypes, exhibited their stability for yield in poor environment (7 genotypes namely, NRCDR-2, TM-151, Kranti, PKRS-28, TM-128, PM-28, RAURD-78; in average environment (2 genotypes, Rajendra Suphlam and KMR-10-2); whereas in rich environment (2 genotypes, Rohini and RH8814).

It is noteworthy that the high yielding mustard genotype, Rajendra Suphlam was stable in average environment followed by two genotypes, Rohini and RH8814 (stable in rich environment), NRCDR-2, TM-151, Kranti and PKRS-28 (stable in poor environment) and KMR-10-1(stable in average environment) co-shared the yield position with TM-151( Biwi et al., 2016 and Tahira et al., 2016 a & b.

Under different moisture regimes (*i.e.* over environments), although Varuna (National Check) exhibited regression coefficient (bi) near unity, non-significant deviation from the regression line (S2di) but its mean performance () for grain yield ha<sup>-1</sup> was below population mean ( $\mu$ ). Hence, its average stability could not be confirmed due to less yield potential under moisture stress condition.

Out of four mustard genotypes (table 3) which have exhibited grain yield ha<sup>-1</sup> stability in poor environment. Three genotypes namely, NRCDR-2, TM-151 and Kranti are medium non-basal branching whereas PKRS-28 is basal-branching genotype.

NRCDR-2 was best stable genotype for grain yield ha<sup>-1</sup>in poor environment and subsequently exhibited stability in poor environment for RGR, LAI, SLW, HI, DME and OY; whereas stability in rich environment for DPM, HFPB, RV, CC, PRO and BY may be suggested to farmers for cultivation in residual moisture – Rainfed condition in Bihar.

TM-151, which showed stability for yield in poor environment has also exhibited similar stability for DCF, LAI, SLW, PRO, HI and DME; and for traits like DPM, SD, RV, RM, SLW, CC, BY

Table 2: Stability parameters for thirty - three morpho-	ters for	thirty	- three	morpho		physio-biochemical	mical a	and quality		traits in l	Indian mustard	nustard									
Characters	Days to	) first flow	Days to first flower open Days to p	Days to p	hysiological	ical	Days to	Days to cessation of	Jot	Height	of first prir	mary	Primar	'rimary branches per	es per	Second	econdary branches	ches	Numbe	Number of siliqua or	on
				maturity			flowerir	Jg.		branch			plant			per pla	nt		Primary	Mother Axis	kis (is
Varieties	×	<u>.</u>	S2di	×	<u>.</u>	S2di	×	<u>.</u>	S2di	×	<u>.</u>	S2di	×		S2di	×	<u>i</u>	S2di	×	<u>:</u>	S2di
DRMRLEJ902	40.75	-3.39	3.89	122.33	1.16	2.36**	95.33	95.33 0.06 11.0	11.01*	*45.04	1.7	90.52**	4.28	0.5	0.29	8.95	-0.56	8.46 **	49.52	1.61	25.82**
DRMR150-35	44.33	-0.25	0.89	121.33	0.86	4.81	96.33	1.	96:0-	43.75	0.95	17.80**	4.18		0.29	8.7	-0.24	1.5	50.97	2.09	15.92**
NRCDR2	41.58	0.16	8.16**	120.92	1.35	-1.45	96.58	1.22	2.04**	46.01	1.39	-1.39	4.42		_	6.65	0.99	1.65*	46.42	0.61	31.36**
RH8814	41.42	1.69**		120.83	1.21	-1.27	93.92	1.29	1.65	48.7	1.36	5.88**	4.35		0.13	6.4	1.03	0.09	46.75	98.0	50.73**
TM151	48.42	-11.3	3.66**	121.42	1.38	-2.06	92.92	29.0	0.54	54.18	1.73**	98.92**	3.93		0.01	6.55	-0.08	0.16	52.72	2.33 **	6.32**
TM128	50.17	-2.49	0.37	122.17	98.0	-0.59	93.5	0.42	-1.87	51.45	2.27**	160.84**	4.03		0.14	8.73	-0.14	1.75*	53.92	1.35	46.27**
KRANTI	4	7.11**	4.73**	120.08	0.43	0.08	94.75	0.46	-0.64	56.34	1.93**	238.27**	4.22		0.21	8.2	0.95	0.63	47.45	1.11	-0.03
KMR10-1	45.58	3.86**	1.17	121.08	1.18	-1.9	94.08	90.0	-1.72	45.14	96.0	13.89**	4.45		0.94	7.03	1.03	4.03 **	42.65	0.56	2.36**
MAYA	41.17	-0.99		122.17	_	5.49	95.42	0.85	3.40**	54.05	1.11	65.49**	4.1		0.13	7.58	1.47	1.36	51.15	1.69**	10.60**
ROHINI	47.58	16.91**!	* 51.00**	121.5	0.39	0.37	94.83	-0.03	-0.91	61.88	1.84**	13.68**	5.37		9.0	14.32	1.17	0.01	53.52	2.22**	14.21**
PKRS28	47.75	4.88** 0.41	0.41		0.02	-1.37	95.58	1.72**	96.0	25.62	0.38	-0.47	4.72		0.13	17.32	1.69**	**60.9	48.23	0.87	31.96**
PUSA MUSTARD 25	46.08	-5.01	11.15**	-	1.26	-2.13	96.33	2.18**	-1.83	47.87	1.63	**09.6	4.43		0.47	10.65	1.24	1.56*	49.88	0.57	3.67**
PUSA MUSTARD 28(NPJ-124)	48.08	3.40**	-0.01	121.08	1.04	-0.13	96.83	1.69**	-0.34	25.7	-0.06	10.72**	4.63		0.09	11.22	2.32 **	14.79**	51.07	1.6	0.52
RGN-13	50.83	3.92**	1.14	119.67	0.57	99.0-	97.75	1.52	8.48	24.4	0.76	-1.15	4.42		. 0.07	10.05	1.69 **	1.62*	48.87	0.68	37.23**
RAURD 212	45.5	1.16	0.02	121.75	0.23	-1.79	97.83	0.55	18.13*	*42.57	0.76	5.21**	5.3		0.74	9.62	0.04	0.31	49.82	0	19.26**
RAURD 78	49.75	2.55	-0.23	120.75	0.84	1.12	94.67	1.67**	-0.03	42.3	96.0	14.30**	4.57		-0.01	11.88	2.25 **	5.03	52.28	99.0	17.26**
VARUNA(CHECK)	46.42	3.78**		130.17	1.7	-1.	97.17	1.19	-2.27	27.16	0.28	-1.25	5.77		2.02*	11.53	1.49	3.03 **	46.18	0.64	4.18**
PUSA BOLD(CHECK)	44.58	2.94**	-0.13	121.17	1.44	96.0	95.5	1.28	0.24	51.78	1.21	6.42**	3.98		, 0.06	11.5	1.38	6.43 **	20.7	0.54	90.0-
PUSA MAHAK(JD-6) (CHECK)	42.25	-0.9	0.74	122.42	1.69	-0.26	96.17	1.35	2.93**	52.29	-0.22	127.14**	4.65		9.0	14.43	1.57	17.86**	51.43	-0.13	0.27
RAJENDRA SUFLAM(CHECK)	38.83	-7.83	11.71**	120.25	1.34	-1.41	89.17	92.0	-1.72	20.72	0.07	-0.34	7.12		92.0	17.78	0.75	2.11*	64.14	0.16	15.19**
SE(bi)	4.9101	_		0.8			0.4975			0.6705			0.6308	~		0.4672	~:		0.4929		
	45.254	_		121.533			95.233			43.348			4.645			10.457	4		50.383		
							1			1			1								

Characters	Length c	ength of Primary Mother	other	Siliquadensity	density		Length (	Length of siliqua		Number	Number of seeds siliqua-1	qua-1	Tap root length	length		Rootvolume	lume		Rootmass	iass	
	Axis																				
Varieties	×	þi	S2di	×	p.	S2di	×	p;	S2di	×	pi	S2di	×	pi	S2di		þi	S2di	×	io	S2di
DRMRLEJ902	61.49		26.30**	0.98	1.86**	0.025	5.02	0.72	-0.002	12.35	0.51	1.24	9.14	1.41	0.04		0.97	0.49	1.25	1.05	0.075
DRMR150-35	63.51		-0.2	0.94	0.15	0.003	4.8	-0.2	0.23	11.87	0.64	0.32	9.02	1.62**	2.41**		1.90**	1.23	1.78	1.03	0.254
NRCDR2	61.21		20.65**	6.0	0.2	0.021	4.76	1.67**	0.144	11.83	3.98**	0.27	10.19	0.52	0.02		2.56**	-0.1	1.82	1.74**	0.133
RH8814 63	63	Ŭ	7.02**	0.83	0.46	0.002	5.03	-0.38	0.044	11.73	-0.37	1.18	9.15	1.32	1.76*	7.28	0.5	0.27	2.22	2.62**	0.399
TM151	60.07		30.75**	1.25	2.60**	0.043	4.79	1.04	0.101	11.63	2.34**	3.04**	12.55	0.59	4.11*		1.16	0.03	1.99	2.07**	0.409
TM128	56.44		-0.86	1.15	0.82	900.0	4.66	1.89**	0.046	11.53	2.17**	2.96**	9.12	1.57	0.44		0.58	0.01	1.52	99.0	0.083
KRANTI	51.54		4.55**	1.26	4.43**	0.098	5.2	0.2	-0.011	12.97	2.02	6.13**	90.6	0.79	1.63*		0.03	-0.11	1.89	1.16	0.002
KMR10-1	58.39		1.98**	0.92	2.05**	0.099	5.13	0.48	0.211	13.2	0.71	92.0	9.34	<del>1</del> .	1.82*		1.16	-0.03	1.82	-0.18	0.038
MAYA	53.43		1.64*	1.55	0.04	1.134	5.61	0.4	0.03	12.45	5.07**	0.71	10.13	1.32	2.33**		0.48	0.32	2.43	0.85	0.778
ROHIN	56.78		17.12**	_	-0.09	0.017	5.13	-0.14	0.968	12.16	-5.6	0.01	10.2	1.86**	3.80**		1.23	0.14	4.05	-0.35	0.335
PKRS28	62.97		0.57	0.91	0.7	0.011	5.12	2.96**	0.003	11.3	-0.42	0.35	6	3.25	0.22		0.77	0.15	1.54	0.29	0.573
PUSA MUSTARD 2	5 60.74		-0.27	1.14	4.15**	0.107	5.29	1.68**	0.355	11.35	0.78	60.0	10.24	1.08	0.15		0.47	0.24	1.88	9.76	0.143
PUSA MUSTARD	58.64		7.76**	96.0	0.27	0.035	5.38	1.7	0.031	11.65	69:0-	0.42	9.51	0.32	1.28		1.38	0.75	1.79	1.03	990.0
28(NPJ-124)																					
RGN-13	58.63	1.36	31.80**	0.87	-0.46	0.007	5.31	2.05 **	0.092	11.97	0.67	0.29	10.01	-1.06	0.79		0.24	90.0-	1.57	1.35	0.002
RAURD 212	54.64	0.21	0.5	0.99	0.49	0.011	4.78	0.01	0.119	11.46	-0.62	0.3	10.64	0.11	0.54		-0.27	1.74*	1.85	1.63**	0.202
RAURD 78	54.93	-0.31	6.71**	0.98	-0.73	0.026	4.72	1.77**	0.386	12	1.90**	0.62	10.2	2.11**	0.17		0.08	0.61	1.42	6.0	0.025
VARUNA(CHECK)	61.86	0.53	4.97**	0.82	0.82	0.003	5.14	4.0	0.146	11.02	99:0-	0.04	10.54	-0.64	3.45		0.74	1.97*	96.0	0.31	0.001
PUSA BOLD(CHECK) 62.19	K) 62.19	4.01**	28.18**	0.89	0.56	0.059	5.37	1.49	0.042	12.12	2.31**	0.08	9.55	1.33	0.97		0.3	0.51	2.04	1.55	0.527
PUSA MAHAK(JD-6)	99 (9	6.0	3.00**	6.0	1.2	0.011	4.98	2.80**	0.153	11.69	1.85**	0.73	13.39	0.31	8.62**	19.77	3.71**	-0.04	2.3	96.0	0.741
(CHECK)																					
RAJENDRA SUFLAM 71.5	M 71.5	-0.26	-1.31	1.15	0.49	0.04	6.11	0.36	-0.01	13.23	0.44	0.03	16.93	0.77	3.73**	26.67	2.02**	1.22	3.2	0.61	-0.004
CE/hi)	0 71 51			1 2507			0.0567			1 2463			1 0120			0 7270			00.0		
JE(DI)	10170			/000.1			0.000			0.10403			1.0129			0.4000			0.39		
	59.998			1.021			5.116			11.974			10.395			8.624			1.965		

Table 2: Continued	per																				
Characters Varieties	Relative growth rate $(x * 10^{-3})$	growth rat	g).	Leafare	Leaf area index		Specific	Specificleafweight	ŧ	Chlorop	Chlorophyll content		Leafmem index	eaf membrane stability ndex	lity	Relativew	Relative water content	<b>.</b>	Excised	Excised leaf water loss	sso
	×	þi	S2di	×	þi	S2di	×	bi	S2di	_ ×	jq		<b>&gt;</b>	bi	S2di	×	i j	S2di	×	þi	S2di
DRMRLEJ902	22.42	1.46	-0.36	2.01	1.49	0.167		1.32			0.94	0.001		1.56	1.18	48.93	1.28	29.61**	34.18	0.25	2.10**
DRMR150-35	24.5	1.65**	3.34**	2.3	1.54	-0.028	6.56	1.53		0.99	1.34		37.66	1.68**	10.75**	53.09	1.35	37.55	34.18	0.23	13.31**
NRCDR2	31.58	96.0	-0.46	3.05	0.83	-0.024		0.83			1.25	•		0.62	1.68*	66.33	0.83	4.58**	25.59	1.24	1.89*
RH8814	28.75	1.87**	11.46**	2.61	1.67**	-0.029		2.57**			1.86	Ì		1.55	9.04	60.72	1.19	10.5	26.91	0.31	10.93**
TM151	30	1.23	2.93**	2.99	0.87	-0.025	8.53	1.01			1.33	·		1.05	3.93**	64.48	0.92	12.76**	27.83	1.12	2.18**
TM128	27.92	0.36	9.0	2.72	0.47	0.065		0.32			0.39			0.63	2.70**	62.78	0.71	-0.08	29.93	1.41	1.32
KRANTI	29.33	0.77	21.92**	2.91	0.4	0.052		0.45	*	1.04	1.07	Ì		9.0	31.21**	65.23	0.81	27.21**	27.3	1.54	27.68**
KMR10-1	28.92	1.33	0.05	2.89	0.84	-0.004		1.37		1.07	1.47	•		1.16	0.89	62.43	1.01	1.66**	29.62	0.73	2.31**
MAYA	26.5	0.92	2.34**	2.51	1.01	900.0		1.05		1.01	1.15	Ì		1.17	-0.11	59.12	96.0	0.37	29.83	1.24	3.16**
ROHINI	33.08	69.0	0.22	3.12	0.83	-0.025		0.92		1.13	1.42	Ì		0.5	4.02**	67.87	0.83	0.13	22.35	1.75**	6.0
PKRS28	28.17	1.21	0.07	2.88	0.93	-0.021		1.18		1.04	1.25	Ì		1.14	2.85	61.01	1.04	0.38	29.1	1.07	-0.08
PUSA MUSTARD 25 24.83	25 24.83	96.0	7.29**	2.26	1.21	0.035		26.0		0.97	0.44			1.2	13.98**	56.07	1.07	12.78**	31.29	1.13	0.08
PUSA MUSTARD 28 28.17	28 28.17	0.58	16.21**	2.77	0.47	0.037		0.63			0.91			0.32	23.06**	63.28	0.73	16.20**	27.5	1.93**	14.40**
(NPJ-124)																					
RGN-13	21.83	1.26	4.60**	1.97	:-	0.015	5.45	1.15	0.97	0.91	0.7	0.002	35.32	1.38	3.85**	51.04	1.15	16.62**	34.79	0.45	3.56**
RAURD 212	21.25	99.0	-0.01	1.98	0.93	-0.017		0.56	0.14		0.32		34.07	1.11	0.15	49.68	1.02	14.22**	34.25	1.05	3.32**
RAURD 78	27	99.0	8.47**	2.72	0.52	0.026		0.77	0.82		0.82	·	요	92.0	12.68**	59.73	0.75	48.56**	29.68	1.37	4.55**
VARUNA(CHECK)	21.08	1.12	-0.12	1.76	1.35	0.02		1.2	60.0		99.0		33.63	1.29	-0.02	45.58	1.29	9.48v	36.8	0.32	11.05**
PUSA BOLD(CHECK)	K) 19.25	1.03	-0.82	1.42	1.71**	-0.007		0.84	0.01		0.78		31.05	1.12	1.5	43.7	1.29	8.08	40.14	-0.13	**69.7
PUSA MAHAK(JD-6)	5) 23.75	0.48	2.76**	2.2	<del>-</del>	0.029		0.36	0.65		9.4		88.59	0.92	**06.9	90.99	0.91	15.23**	31.5	1.36	0.37
RAJENDRA SUFLAM 33.75 (CHECK)	M 33.75	0.8	0.82	3.29	0.79	-0.003	10.11	66.0	0.63	1.16	1.53	0.002	49.11	0.27	3.37**	70.95	0.87	21.69**	20.65	1.62**	1.27
SE(bi)	0.2888 26.604			0.1723 2.518			0.2873 7.263			0.158		0 1	0.2285 40.254			0.1045 58.403			0.3197		

-4035.16 64634.20\*\* 16917.89\*\* 8207.62\*\* 11802.91\*\* 3471.35\*\* -8833.57 87756.46\*\* 0449.61\*\* 45689.87\*\* 48877.13\*\* 41713.45\*\* -7478.34 -15379.6 -6007.9 -12360.31 -9742.27 -8214.94 7040.82 1.90 \*\* 0.74 0.95 0.84 0.19 0.8 1.32 1.25 1.06 1.23 1.23 1.28 0.56 1.04 0.7 1.45 0.63 0.81 0.54 0.83 Biological yield 1520.67 1598.33 1494.067 1819.17 1234.17 1423.33 1514.17 1666.67 1369.17 780.83 1869.17 1610.83 1229.58 1634.58 1543.33 1216.67 1368.33 1787.5 1197.5 1205 0.3 -0.09 -0.08 0.12 0.08 -0.07 -0.08 -0.09 0.02 0.64 0.05 0.27 0.07 0.04 -0.08 90.0 0.07 0.31 1000 seed weight 2.20 \*\* 2.49 \*\* 0.7627 2.16\*\* 2.05 \*\* 2.06 0.03 0.36 0.48 0.83 -1.91 0.73 1.02 5.25 4.9 5.02 4.69 5.25 5 4.21 5.12 4.27 4.92 6.34 5.03 5.41 0.002 0.005 0.01 0.001 0.004 0.001 0.001 0.001 0.002 Proline accumulation 1.63\*\* 1.62\*\* 1.85 1.34 1.04 0.71 0.85 1.31 0.77 0.81 0.95 0.2606 1.04 0.887 0.96 1.01 0.92 0.84 0.81 0.85 0.99 0.92 0.95 0.93 0.87 0.92 0.88 0.83 84.90\*\* 24.13\*\* 510.66\*\* 206.60\*\* 332.73\*\* 72.57\*\* 106.64\*\* 600.18\*\* 260.14\*\* 756.39\*\* 32.47\*\* 56.24\*\* 34.56\*\* \*\*06.62 16.40\*\* 75.89\*\* 61.69\*\* 45.02\*\* 14.13\*\* -1.23 0.55 2.81\*\* 3.22\*\* 0.47 0.88 3.19\*\* 2.23\*\* 2.55 \*\* 2.00\*\* -1.46 2.50\*\* Peroxidase activity 2.78\* -1.49 0.32 0.33 0.41 69.0 -0.68 227.074 216.52 248.08 240.38 234.29 238.33 254.42 234.7 206.36 227.51 183.15 260.76 222.34 237.3 223.4 236.3 202 1.9 93.35\*\* 583.07\*\* 1283.19\*\* 159.52\*\* 3.96\*\* 525.95 \*\* 701.24\*\* 29.46 05.53\*\* 261.72\*\* 361.08\*\* 771.07\*\* 254.84 \*\* 81.28 \*\* 49.31 \*\* 45.50\*\* 31.98\*\* 10.98\*\* 92.47\*\* 0.74 1.29 1.75\* 0.83 0.23 0.44 1.23 0.29 0.38 1.58 1.49 0.82 1.26 1.09 0.92 0.6 **Catalase** activity 0.95 139.508 175.05 149.82 111.58 108.94 126.53 152.41 162.77 148.14 157.94 134.06 128.24 105.93 136.91 99.62 180.05 148.4 125.9 170.3 9.0 -0.004 -0.004-0.004 -0.004 -0.003 -0.005 -0.004-0.005 -0.003 -0.004 -0.003-0.003 -0.003 0.005 Stress intensity 1.41 1.44 1.36 1.18 0.54 0.86 0.8 1.05 1.09 1.01 0.51 0.32 1.34 0.79 0.95 0.48 1.42 1.6 0.1846 0.41 0.38 0.37 0.36 0.42 0.33 0.36 0.45 0.38 Drought tolerance index -0.05 -0.03 -0.05 0.04 -0.050.12 1.41 1.22 1.05 0.66 0.94 0.84 1.07 1 0.58 1.27 0.83 0.98 0.65 1.28 1.49 bi 0.84 1.32 0.98 0.98 0.61 X
DRWRLEJ902 1.65
DRWR150-35 1.96
NRCDR2 1.37
RH8814 2.24
RM1814 1.57
RM128 1.37
KWANTI 1.37
KWANTI 1.31
KWANTI 1.34
KWANTI 1.34 RAURD 78 1.43 VARUNA(CHECK) 1.75 PUSA BOLD(CHECK) 1.6 PUSA MAHAK(ID-6) 1.62 0.141 USA MUSTARD 28 1.25 NPJ-124) 1.75 RAJENDRA SUFLAM 1.98 (CHECK) DRMR150-35 RGN-13 RAURD 212 Characters RAURD 78 TM151 TM128 KRANTI ROHINI RH8814 /arieties (CHECK) MAYA

Fable 2: Continued

Continued	Collinaea
Table 2.	I anne 7:

						_					_						*												
	S2di	-2017.16	-1671.01	-2017.03	-1748.61	-1053.39	-1669.87	-348.2	-1582.45	-2245.56	-2239.42	-959.89	-1723.05		-1307.5		201.22**	-2157.55	-1585.34	-2256.21		-1990.66		-2025.94		-1876.95			
Oil yield ha-1	þi	<u></u>	1.18	0.92	1.77**	0.97	0.65	0.78	1.16	0.97	1.39	<del>-</del>	0.98		0.71		1.04	6.0	0.73	1.04		0.84		0.81		1.17			
Oil yie	×	527.7	552.74	646.34	626.76	637.86	604.59	630.41	630.57	575.49	713.45	607.29	560.24		616.57		517.79	524.86	597.95	517.5		456.99		555.25		720.08		0.1	592.687
1	S2di	-13964.49	-10512.5	-13061.06	-11960.98	-4901.22	-11666.29	-1660.9	-10974.51	-14756.43	-14443.24	-4604.9	-11688.81		-8172.42		1648.03 * *	-14196.31	-10434.9	-14493.28		-12946.93		-12187.35		-12170.33			
Grain yield ha-1	.iq	1.1	1.16	6.0	1.75**	0.98	0.65	92.0	1.16	0.98	1.34	0.97	96.0		0.7		1.04	0.91	9.76	1.03		98.0		0.84		1.08			
Grai	×	1384.12	1421.15	1676.22	1698.9	1620.21	1555.4	1601.69	1620.21	1485.96	1816.95	1578.55	1430.41		1566.97		1337.83	1360.98	1532.25	1342.46		1171.18		1439.67		1897.96		0.1	1526.954
	S2di	0.28	-0.07	-0.09	-0.02	0.05	-0.04	-0.08	-0.1	-0.09	-0.07	0.14	-0.1		0.03		-0.06	-0.03	0.02	-0.1		0.11		0.07		0.03			
Dil content	bi	2.56**	0.94	0.62	1.29	1.88**	1.14	0.62	0.94	0.99	0.39	0.82	1.04		0.4		8.0	0.82	0.94	0.38		1.33		0.88		1.24			
0	×	38.32	38.88	38.56	38.81	39.3	38.81	39.38	38.96	38.77	39.27	38.56	39.15		39.26		38.73	38.58	39.09	38.5		39.12		38.58		38		0.5968	38.83
,	S2di	1.79*	-1.69	1.4	6.20**	-1.37	19.59**	1.6	7.95**	9.20**	3.33**	-0.54	5.60**		1.99**		1.54	7.21**	0.05	1.02		-0.88		0.73		-0.47			
Dry matter efficiency	þi	0.38	1.37	-0.04	-0.57	-0.11	0.47	1.47	1.70**	1.46	0.63	0.55	96.0		2.62**		5.06	1.34	3.11**	1.24		0.62		0.82		-0.07			
Dry matte	×	15.72	12.82	16.77	18.38	15.99	15.4	13.51	20.21	12.99	14.33	14.68	17.69		14.65		16.56	14.55	15.51	15.14		14.51		15.4		16.07		1.1112	15.545
	S2di	3.73**	-2.31	0.84	11.62** 1	-2.2	28.57**	2.96** 1	11.76**	12.87** 12.99	4.78**	-1.14	8.10**		2.23 **		3.42 **	9.91	2.13 **	2.55		-1.11		1.64**		-1.17			
Harvest index	þi	0.61	1.53	-0.13	-0.25	-0.07	0.12	1.18	1.67**	1.02	92.0	0.61	1.05		2.48**		1.74**	1.41	2.87**	1.51		0.85		1.1		-0.04			
Harv	×	19.25	15.58	20.26	22.23	19.41	18.78	16.22	24.46	15.81	17.39	17.56	21.2		17.71		19.79	17.7	18.7	19.72		17.6		18.87		19.31		1.0685	18.876
Characters	Varieties	DRMRLEJ902	DRMR150-35	NRCDR2	RH8814	TM151	TM128	KRANTI	KMR10-1	MAYA	ROHINI	PKRS28	PUSA MUST	ARD 25	PUSA MUST	ARD 28(NPJ-124)	RGN-13	RAURD 212	RAURD 78	VARUNA	(CHECK)	PUSA BOLD	(CHECK)	PUSA MAHAK	(JD-6) (CHECK)	RAJENDRA SUF	LAM(CHECK)	SE(bi)	

Table 3: Stable genotypes (in poor, average and rich environments) for morpho-physio-biochemical and quality traits in Indian mustard under different moisture regimes

S.NO.	CHARACTER	STABLE IN POOR ENVIRONMENT	STABLE IN AVERAGE ENVIRONMENT	STABLE IN RICH ENVIRONMENT
		$_{\rm X}^- > \mu$ , bi < 1, NS S2di	$_{\rm X}^- > \mu$ , bi = 1, NS S2di	$_{\rm X}^- > \mu$ , bi > 1, NS S2di
1 2	Days to first flower open Days to physiological maturity	DRMR 150-35,MAYA,PUSA MAHAK KRANTI,PKRS 28,RGN-13,RAURD 78	- PUSA MUSTARD -28	RH8814,PUSA BOLD NRCDR-2,RH8814,TM 151,KMR10-1,PUSAMUST ARD -25, PUSA BOLD, RAJENDRA SUPHLAM
3	Days to cessation of flowering	TM151, TM128, KRANTI, KMR10-1, RAJENDRA SUFLAM	-	RH8814, RAURD 78
4	Height of first primary branch			
a	BB TYPE	PKRS28, RGN-13 VARUNA, RAJENDRA SUFLAM	-	-
b	NBB TYPE	-	-	DRMR150-35
5	Primary branches per plant	PKRS28, PUSA MAHAK, RAJENDRA SUFLAM	-	RAURD 212
6	Secondary branches per plant	RAJENDRA SUFLAM	-	ROHINI, PUSA MUSTARD 25
7	Number of siliqua on Primary Mother Axis	PUSA MAHAK	-	PUSA MUSTARD 28(NPJ-124)
8		RAJENDRA SUFLAM	-	DRMR150-35, PKRS28, PUSA MUSTARD 25
9	Siliqua density	TM128, RAJENDRA SUFLAM	MAYA	TM151, KRANTI, PUSA MUSTARD 25
10	Length of siliqua	KRANTI, KMR10-1, MAYA, ROHINI, VARUNA, RAJENDRA SUFLAM	-	PUSA MUSTARD 25, PUSA MUSTARD 28 (NPJ-124), RGN-13, PUSA BOLD
11	Number of seeds siliqua-1	DRMRLEJ902, KMR10-1, RAJENDRA SUFLAM	-	MAYA, ROHINI, PUSA BOLD
12	Tap root length	RAURD 212	_	_
13	Root volume	-	-	DRMR150-35, NRCDR2, TM151, PUSA MAHAK,
				RAJENDRA SUFLAM
14	Root mass	MAYA, ROHINI, PUSA MAHAK, RAIENDRA SUFLAM	-	RH8814, TM151
15	Relative growth rate	NRCDR2, TM128, ROHINI, RAJENDRA SUFLAM	-	KMR10-1, PKRS28
16	Leaf area index	NRCDR2, TM151, TM128, KRANTI, KMR10-1, ROHINI, PKRS28, PUSA	-	RH8814
17	Specific leaf weight	MUSTARD 28(NPJ-124), RAJENDRA SUFLAM NRCDR2, TM128, ROHINI, PUSA MUSTARD 28(NPJ-124), RAJENDRA SUFLAM	-	RH8814, TM151, KMR10-1PKRS28

and OC in rich environment.

Kranti had yield stability for poor environment and also exhibited stability for DPM, DCF, LS, LAI, OC and OY in poor environment, in average environment for CC and for SD, PRO, and BY in rich environment.

PKRS-28 exhibited poor environment yield stability and also reflected similar stability for DPM, HFPB, PBP, LAI, PRO, TSW, BY; RWC and OY in average environment and LPMA, RGR, SLW and CC in rich environment( Mariotti et al., 1975).

Rajendra Suphlam a popular mustard variety in Bihar exhibited high grain yield ha<sup>-1</sup> and stability in average environment and also reflected stability for OY in similar environment; for DCF, HFPB, PBP-1, SBP-1, LPMA, SD, LS, SS, RM, RGR, LAI, SLW,

TSW, HI, DME in poor environment and in rich environment for DPM, RV, CC, DTI, SI, PRO and BY may be a good option for farmers for cultivation in both agro- ecologies as average stable genotype.

Rohini showed yield stability in rich environment and also exhibited stability in poor environment for DCF, LS, RM, RGR, LAI, SLW, RWC and OC; stability for PBP-1, SBP-1, SS, CC, PERO,PRO and OY in rich environment.

RH-8814 reflected stability for yield in rich environment, stability for PRO in poor environment and for DFFO, DPM, DCF, RM, LAI, SLW, CC, RWC, DTI, SI, TSW and OY in rich environment. This genotype is only, which showed DFFO including LAI and SLW stable in rich environment and as third high yielder reflected that this genotype could perform

18	Chlorophyll content	-	-	NRCDR2, RH8814, TM151, KRANTI,
				KMR10-1, MAYA, ROHINI, PKRS28, RAJENDRA SUFLAM
19	Leaf membrane stability	_	_	KMR10-1, MAYA
'	index			KIVIKTO 1, MIXTIX
20	Relative water content	TM128, MAYA, ROHINI	-	KMR10-1, PKRS28
21	Excised leaf water loss	-	-	PUSA MUSTARD 25, PUSA MAHAK
22	Drought tolerance index	DRMRLEJ902, KMR10-1, RGN-13,	-	DRMR150-35, RH8814, ROHINI,
		VARUNA		RAURD 212, RAJENDRA SUFLAM
S.NO.	CHARACTER	STABLE IN POOR ENVIRONMENT	STABLE IN AVERAGE	STABLE IN RICH ENVIRONMENT
			ENVIRONMENT	
		$_{\rm X}^- > \mu$ , bi < 1, NS S2di	$_{\rm X}^{-} > \mu$ , bi = 1, NS S2di	$_{\rm X}^- > \mu$ , bi > 1, NS S2di
23	Stress intensity	DRMRLEJ902, KMR10-1RGN-13,	-	DRMR150-35, RAURD 212,
	,	VARUNA		RAJENDRA SUFLAM
24	Catalase activity	-	-	-
25	Peroxidase activity	-	-	-
26	Proline accumulation	RH8814, TM151, KMR10-1, PKRS28	-	NRCDR2, TM128, KRANTI,
				ROHINI,
				PUSA MUSTARD 28(NPJ-124),
	1000	DIVEGGO DALLED TO DALENDRA CLIELAN		RAJENDRA SUFLAM
27	1000 seed weight	PKRS28, RAURD 78, RAJENDRA SUFLAN	1 -	DRMR150-35, RH8814, KMR10-1,
				RAURD 212, PUSA BOLD, PUSA MAHAK
28	Biological yield	PKRS28		NRCDR2, TM151, TM128, KRANTI
20	Diological yield	1 KK320	-	TARCERZ, TWIST, TWIZE, REMAINT
29	Harvest index	NRCDR2, TM151, RAJENDRA SUFLAM	-	-
30	Dry matter efficiency	NRCDR2, TM151, RAJENDRA SUFLAM	-	-
31	Oil content	KRANTI, KMR10-1, ROHINI, , PUSA	PUSA MUSTARD 25	TM151, PUSA BOLD
		MUSTARD 28(NPJ-124), RAURD 78		·
32	Grain yield ha-1		rajendra suflam,	RH8814, ,ROHINI
		PKRS28, PUSA MUSTARD 28(NPJ-124),	KMR10-1	
		RAURD 78		
33	Oil yield ha-1	, , , , , , , , , , , , , , , , , , , ,	PKRS28	RH8814, KMR10-1, ROHINI,
		PUSA MUSTARD 28(NPJ-124),		RAJENDRA SUFLAM
		RAURD 78		

Table 4: Stability parameters for grain yield and its morpho-physio-biochemical and quality traits under poor, average and rich environments

S.NO.	Genotypes/ stability	Bran ching	Pooled yield overenvironments	Eberhert russell (three parameters) model Stable in	Stable in average	Stable in rich environment
		behaviour	(E1 + E2 + E3 + E4)	poor environment	environment	$_{\rm X}^- > \mu$ , bi > 1, NS S2di
				$\frac{-}{X} > \mu$ , bi < 1, NS S2di	$\frac{-}{X} > \mu$ , bi = 1, NS	5 S2di
Stable i	n poor environment $(\frac{-}{X} > \mu)$	, bi < 1, NS S2	di): 7 Genotypes			
1	NRCDR-2	NBB	1676.22*	RGR, LAI, SLW, HI, DME, OY	-	DPM, HFPB, RV, CC, PRO, BY
2	TM-151	NBB	1620.21*	DCF, LAI, SL, PRO, HI, DME	-	DPM, SD, RV, RM, SLW,CC,BY,OC
3	TM-128	NBB	1555.40*	DCF, SD, RGR, LAI, SLW, RWC, OY	-	PRO, BY
4	KRANTI	NBB	1601.69*	DPM, DCF, LS, LAI, OC, OY	CC	SD, PRO, BY
5	PKRS-28	BB	1578.55*	DPM, HFPB, PBP, LAI, PRO, TSW, BY	RWC, OY	LPMA, RGR, SLW, CC
6	PM-28	BB	1566.97*	LAI, SLW, CC, OC, OY	DPM	SPMA, LS, PRO
7	RAURD78	NBB	1532.25*	DPM, DCF, TSW, OC, OY	-	=
Stable i	n average environment ( $\frac{1}{X}$	$>\mu$ , bi = 1, NS	S2di): 2 Genotypes			
8	RAJENDRA SUPHLAM	BB	1897.96**	DCF, HFPB, PBP,SBP, LPMA, SD, LS, SS, RM, RGR, LAI, SLW, TSW, HI, DME	OY	DPM, RV, CC, DTI, SI, PRO, BY
9	KMR-10-1	BB	1620.21*	DCF, LS, SS, LAI, DTI, SI, PRO, OC,	RWC, OY	DPM, RGR, SLW, CC, TSW
Stable i	n rich environment ( $\frac{-}{X} > \mu$ ,	bi > 1, NS S2c	li) : 2 Genotypes			
10	RH8814	NBB	1698.90*	PRO	-	DFFO, DPM, DCF, RM, LAI, SLW, CC, RWC, DTI, SI, TSW, OY
11	ROHINI	NBB	1816.95*	DCF, LS, RM, RGR, LAI, SLW, RWC, OC	DTI, SI	PBP, SBP, SS, CC, PERO, PRO, OY
12	VARUNA	BB	1342.46	DTI, SI, LS, HFPB	- '	-
	$\frac{-}{X}$ < $\mu$ , bi = 1, NS S2di					

better in environment which is well irrigated(Rashid et al., 2002; Chaudhary et al., 2004; Gupta and Pratap, 2007; Yadava et al., 2010; Priyamedha et al., 2017 for stability of

various morpho-physio-biochemical traits and Dar et al., 2011 and Moghaddam and Pourdad (2011) for stability of oil yield along with seed yield/plant).

Rajendra Suphlam showed superiority for all flowering — maturity characteristics, height of first primary branch and siliqua, root parameters and grain yield ha-1, Rohini showed superiority for late DFFO, DFF, DPM, low IL, dwarfness and NRCDR-2 for late DFFO, DPM were crossed to get three divergent crosses namely, Rohini/ Rajendra Suphlam, Rajendra Suphlam/ NRCDR-2 and Rohini / NRCDR-2. Among these crosses, only two crosses Rohini/ Rajendra Suphlam and Rajendra Suphlam/ NRCDR-2 they involve Late × Early (days to first open and days to physiological maturity), Basal/Non-Basal, High × Low placed siliqua, Low×High (harvest- index and dry matter efficiency) and Rich× Average and Average × Poor (stability for grain yield ha-1) parents could through transgressive seggregants which may be stable across the environments.

It can be suggested to the farmers of Bihar that under moisture stress condition, NRCDR-2 (Variety 2007) is the best option for poor environment stability but, average stable genotype, Rajendra Suphlam (popular variety in Bihar) with highest grain yield ha<sup>-1</sup> in individual (E1, E2, E3 and E4) as well as pooled over environments, may give choice of cultivation across the moisture regime environments in Bihar. However, under rich environment Rohini has provided second highest yield performance after Rajendra Suphlam( Dhillon et al., 1997).

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# **REFERENCES**

- **Abou El-Nasr, T.H.S., Ibrahim, M.M. and Aboud, K.A. 2006.** Stability Parameters in Yield of White Mustard (*Brassica Alba* L.) in Different Environments. *World J. Agricultural Sciences.* **2(1):** 47-55.
- **Baker, R.J. 1969.** Genotype environment interaction in yield of wheat. *Canadian J. Plant Science*.**49:** 743-751.
- Barr, H. D. and Weatherley, P. E. 1962. A re-examination of the relative turgidity technique for estimating water deficit in leaves. *Australian J. Biological Science*. 15: 413-428.
- Bates, L., Waldren, R.P. and Teare, I.D. 1973. Rapid determination of free proline for water-stress studies. *Plant and Soil*. 39: 205-207
- Bibi, T., Rauf, S., Mustafa, H.S.B., Mahmood, T. and Salah-ud-Din.2016. Selection of stable mustard (Brassica junceaL.) genotypes through genotype × environment interaction and stability analysis suitable for Punjab, Pakistan. *J. Agriculture and Basic Sciences*. Vol. 01(1):14-17.
- Byth, D. E., Caldwell, B. E. and Weber, C. R. 1969. Specific and nonspecific index selection in soybeans. *Glycine max* L. (Merrill). *Crop Science*. 9:702-705.
- Campbell, C. A., Zentner, R. P., McConkey, B. and Selles, F. 1992. Effect of nitrogen and snow management on efficiency of water use by spring wheat grown annually on zero-tillage. *Canadian J. Plant Science*. 72: 271–279.

- Chaudhary, S.P.S., Choudhary, A.K., Singh, R.V., Singh, N.P. and Shrimali, M.K.2004.Genotype X environment interaction for yield contributing characters in Indian mustard [*Brassica juncea* (L.)Czern&Coss].Research on crops. **5(2&3)**:232-239.
- Comstock, R. E. and Moll, R. H. 1963. Genotype-environment interactions. Pages 164-196 in W. D. Hanson and H. F. Robinson, eds. Statistical genetics in plant breeding. NASNRC, Washington, D.C. Publ. 982.
- Dar, Z. A., Gulzaffar, S. A. W., Ahmad. I., Sheikh, F. A., Ishfaq, A., Razvi, S. M. and Habib, M. 2011.GxE interaction for seed yield and oil content in brown sarson (*Brassica rapaL*.) under temperate conditions. Cruciferae Newsletter. vol. 30.PP.25-27.
- **Dhillon, S.S., Singh, K. and Brar, K.S. 2000.** Stability analysis of elite strains in Indian mustard. In: 10thInternational rapeseed Congress, Canberra, Australia (1999), *Indian J. Agricultural Science.* **70(8):** 554-558.
- **Donald, C.M. and Hamblin, J. 1976.** The biological yield and harvest index of cereals as agronomic and plant breeding criteria. Adv. Agron. **28:** 361-405.
- **Euler, H. and Josephson, K. 1927.**Photometric determination of catalase activity. Annual Chemistry. **462:**158
- **Fischer, R.A. and Maurer, R. 1978.** Drought resistance in spring wheat cultivars. I. Grain yield responses. *Australian J. Agricultural Research.***29:** 897–912.
- Gazal, A., Dar, Z.A., Zafar, G. and Habib, M. 2013. Stability analysis for yield and its contributing traits in Brown sarson(Brassica rapal.) under Kashmir conditions in India. *J. Oilseed Brassica*. 4(1): 33-38.
- **Gupta, S.K.** and **Pratap, A. 2007.** Phenotypic stability of Indian mustard (Brassica juncea) genotypes developed from intervarietal and intergeneric crosses. *Indian J. Crop Science.* **2(2):** 379-382.
- Jinks, J.L. and Jones, R.M. 1958. Estimation of the components of heterosis. *Genetics*. 43:223-234.
- **Knight, R. 1970.**The measurement and interpretation of genotype environment interactions. *Euphytica*. **19:** 225–35.
- Kumar, V., Arunachalam, V. and Chakrabarty, S.K. 1996. Genetic variability for plant type traits in Brassicaspecies *.J. Oilseeds Research*. 18(2):161-163.
- Langer, S., Frey, K.J. and Bailey, T. 1979. Association among productivity production response and stability indices in oat varieties. *Euphytica*. 28:17-24.
- Lewis, F.B. 1954. Gene-environment interaction. Heredity. 8: 333-356
- Malik, T.A. 1995. Genetics and Breeeding for Drought Resistance in Wheat: Physio-molecular Approaches. Ph.D. thesis. University of Wales. UK.
- Mariotti, J.A., Oyarzabal, E.S., Osa, J.M., Bulacia, A.N. R. and Almada, G.H. 1975. Analysis of stability and adaptability in sugarcane genotype. *J.Interactions With inan experimentalsite*. Revista Agronomicadel NoroesteArgentino.13 (1/4): 105-118 (Es, en, 15 ref.)
- **Moghaddam, M. J. and Pourdad, S. S. 2011.**Genotype  $\times$  environment interactions and simultaneous selection for high oil yield and stability in rainfed warm areas rapeseed (*Brassica napus* L.) from Iran. Euphytica. **180:** 321–335.
- **Palmiano, E. P. and Juliano, B. C. 1973.** Changes in the Activity of Some Hydrolases, Peroxidase and Catalase in Rice Seed During Germination. *Plant Physiol.* **52:** 274-277.
- **Pearce, R.B., Brown, R.K. and Blaser, R.E. 1968.** Photosynthesis of Alfalfa leaves as influenced by age and environment. *Crop Science.* **8:** 677 680.

**Pfakler, P.L. and Linskens, H. L. 1979.** Yield stability and population diversity in Oat (Avena sp.). *Theory of Applied Genetics.* **54**:1-5.

Powel, W., Caligari, P.D., Phillips, M.S. and Jinks, J.L. 1986. The measurement and interpretation of genotype by environment interaction in spring barley (Hordeumvulgare). Heridity. 56: 255-262.

**Premchandra, G.S., Saneoka, H. and Ogata, S. 1990.** Cell membrane stability, an indicator of drought tolerance as affected by applied nitrogen in soybean. *J. Agri. Sci.* **115:** 63-66.

**Priyamedha, Kumar, A. and Haider, Z.A. 2017.** Stability for seed yield and component traits in Indian mustard (*Brassica junceal*..) under Jharkhand condition. *J. Oilseed Brassica*. **8(1):** 37-42.

**Ramagosa, I. and Fox, P.N. 1993.**Genotype × Environment interaction adaptation. In: M.D. Hayward, N.O. Bosemark and P.N. Fox(eds.) Plant Breeding Principles and Prospects, Chapman and Hall, London.pp.373-390.

Rashid, A., Hazara, G.R., Javid, N., Nawas, M.S. and Ali, G.M. 2002.Genotype x environment interaction and stability analysis in mustard. *Asian J. Plant Science*. Vol. 1(5): 591-592.

Ray, K., Dutta, J., Banerjee, H. Biswas, R., Phonglosa, A. and Pari, A. 2014. Identification of principal yield attributing traits of Indian mustard [*Brassica juncea* L czernj and cosson] using multivariate analysis. The Bioscan. 9(2): 803-809.

**Robertson, M. J. and Holland, J.F.2001.** Production risk of canola in the semi-arid subtropics of Australia. *Australian J. Agricultural Research.* **55:** 525-538.

Sagolsem, D., Singh, N.B., Devi, R., Wani, S.H., Haribhushan, A., Singh, N. G. and Laishram, J. M. 2013. Genotype x environment interaction in Indian mustard (*Brassica juncea* L. Czern and Coss) under Manipur valley conditions. *Indian J. Genetics*. 73(3): 332-334.

Sah, R. P., Kumar, A., Ghosh, J. and Prasad, K.2015. Stability study

in Indian mustard (Brassica juncea L.)J. Hill Agriculture.6(1): 40-44.

Sairam, R.K. 1994. Effect of moisture stress on physiological activities of two contrasting wheat genotypes. *Indian. J. Exp. Biol.* 32:594-597.

**Schuster, W. and Sra, S.S. 1979.** Income structure of various winter and summer rape raps locations. Field and crop production.**148:** PP.348-366.

Shekhawat, N., Jadeja, G. C., Singh, J. and Shekhawat, R.S. 2014 character association studies among yield and its component characters in Indian mustered (*Brassica juncea* L czern & coss). *The Bioscan.* 9(2): 685-688.

Tahira, Rashid, A., Khan, M.A. and Amjad.2016a. Stability analysis of canola (Brassica napus) genotypes in Pakistan. Global Advanced Research J. Agricultural Science .Vol. 2(10).pp. 270-275.

**Tahira, Rashid,A., Khan, M.A. and Amjad.2016b.** Stability Analysis of Rapeseed Genotypes Targeted Across Irrigated Conditions of Pakistan. *International J. Agriculture Innovations and Research.* Volume **2(2):** 208-212.

**Thomas, R. C., Grafius, J. E. and Hahn, S. K. 1971.** Stress: An analysis of its source and influence. Heredity. **27:** 423–432.

**Watson, D.J. 1947.** Comparative physiological studies in the growth of field crops. I. Variation in net assimilation rate and leaf area between species and varieties, and within and between years. Annals of Botany. **11:** 41-76.

**Williams, R.F. 1946.** The physiology of plant growth with special reference to the concept of net assimilation rate. Annals of Botany. **10:** 41 - 42.

Yadava, D. K., Giri, S. C., Vasudev, S., Yadav, A.K., Dass, B., Raje, R. S., Vignesh, M., Singh, R., Mohapatra, T.andPrabhu, K. V. 2010. Stability analysis in Indian mustard (Brassica juncea) varieties. *Indian J. Agricultural Sciences*. 80(9): 761-765.