METABOLOMIC VARIATION STUDY ON THE GENETICALLY VARIANT MICROBIAL STRAINS OF *BORASSUS FLABELLIFER* SAP COLLECTED FROM

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KANYAKUMARI AND TIRUNELVELI DISTRICTS

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ABSTRACT

Borassus flabellifer (Palmyra tree) has a unique specificity with gallons of medicinal properties in it. Contemporary science has proved that consumption of fruits, leaves, bark, and roots has a role to play in boosting as well as disease-preventing benefits because of a few substances, namely phytochemicals, viz., polyphenols, vitamins, minerals, proteins, etc. (Prasad et al. 2016). Palm sap is a non-alcoholic, refreshing natural beverage that has a plethora of health benefits of various natures. As the sap is rich in microflora, it is vulnerable to fermentation instantaneously after the harvest. Toddy palm nectar (TPN) is a naturally fermented sap from young and matured inflorescences of Borassus flabellifer Linn. The palm sap fermentation process is controlled by a complex microbial community that evolved during natural fermentation. In the present research, the bacterial strains present in the Palmyra sap responsible for the fermentation of Palmyra sap to toddy were collected from two different geographical communities of Kanyakumari and Tirunelveli Districts, Tamilnadu and were investigated using metagenomics DNA molecular studies and their phylogenetic variation were analyzed using Bioinformatics Tools. The results reported that the Bacterial strains responsible for fermentation was present predominantly in the Palmyra sample collected from Kanyakumari District compared to the samples collected from Tirunelveli District. Comparative metagenomics analysis revealed the presence of beneficial probiotic bacteria in the sap, with a predominance of Proteobacteria and Firmicutes, highlighting the saps potential probiotic benefits.

Introduction

B. flabellifer, a dioecious monocotyledonous woody perennial tree in the family Arecaceae, is a massive palm with a single stem reaching 30 m in height and large fan-shaped leaves spanning 4–6 m in diameters. In India the wealth of

Palmyra palm is very rich with a population nearly 122 million palms (Vengaiah *et al.*, 2012). India's rich biodiversity and ethno culinary traditions offer a vast landscape for microbial bioprospecting (Tamang *et al.*, 2021). Among traditional beverages, palmyra palm nectar, locally known as



Pathaneer in Tamil Nadu, is a naturally fermenting sap extracted from the inflorescence of B. flabellifer Linn, a culturally significant palm species prevalent South and Southeast Asia. Fresh Pathaneer is a mildly sweet, translucent liquid with a near-neutral pH (6.0–7.0), rich in carbohydrates, amino acids, vitamins, organic acids, and polyphenols (Lasekan and Abbas, 2010; DebMandal and Mandal, 2011). The quality and quantity of palm sap collected in each cylinder vary based on factors such as hygiene maintenance, preservatives used during harvesting. environmental conditions, tapping duration, palm variety, and soil fertility (Hebbar et al., 2018; Sarkar et al., 2023). Fermented palm nectar is a relatively untapped source for exploring exopolysaccharide (EPS) production and β-galactosidase activity. The commercial interest in β-galactosidaseproducing probiotics has grown due to their potential application in managing lactose intolerance (Srinivash al.. 2023). **Probiotic** strains are often characterized by their ability to withstand harsh gastrointestinal conditions, including low pH, bile salts, and digestive enzymes. In addition, functional their efficacy evaluated through properties such as coaggregation, auto-aggregation, cell surface hydrophobicity, antioxidant function. adhesion to intestinal epithelial cells, possession of inhibitory effects towards

enteric pathogens, and immunomodulatory potential (Saadat et al., 2019). During natural fermentation, the composition of microbiota in palm saps may change their dynamics (Kouamé et al., 2020) due to the fluctuation of micro-environments (pH, alcohol content, sugar profile and organic acids) (Karamoko et al., 2012), Predictive functional profiles of microbial communities in fermented foods is an appropriate approach to annotate predictive metabolic pathways in gene sequences of bacteria and fungi (Ortiz-Estrada et al., 2019). Hence, in our present study, we aimed to study the bacterial community present in the natural fermentation of Palmyra sap collected from Kanyakumari and Tirunelveli Districts of Tamilnadu in India by high-through put sequencing tool and predict the functional profiles of bacterial genes by using the bioinformatics pipelines.

Materials and Methods

Collection of Palmyra palm sap samples

Fresh palmyra palm sap samples were collected from two geographically distributed areas, namely Kanyakumari and Tirunelveli districts Tamil of Borassus flabellilfer is extensively Nadu. found in Kanyakumari the region, particularly in Karungal, where the

extraction of sap reaches its peak from August to November. In the Tirunelveli region, it is widely distributed in the area of Aral, and the extraction of sap is high from April to June. Palm sap is tapped from the mature, unopened inflorescence of the palm. The palm sap was collected by cutting the head of the inflorescence. In rural areas, palm sap was traditionally collected from palmyra trees by organized practice for its local consumers.

The fresh samples were collected in the early morning by professional tappers. As soon as the collected fresh palmyra sap samples were brought down from the palmyra tree, they were transferred to sterilized bottle, packed in a sterile ice box to avoid fermentation during transportation, and transported to the laboratory for analysis. On reaching the laboratory, the palmyra sap samples were stored in the refrigerator at about 4°C for further analysis.



Figure. 1. Collection of sap from Borassus flabellifer

Metagenomic Analysis

Metagenomic analysis was performed to detect the presence and diversity of microbes in the Palmyra sap sample collected from two different geographical locations viz., Kanyakumari and Tirunelveli Districts. Initially, the total DNA was isolated from the Palmyra sap sample using the soil DNA isolation kit from MagGenome. Following this, the DNA was isolated using the manufacturer's



protocol (http://www.maggenome.com/wp-content/uploads/2022/04/XpressDNA-Soil-kit.pdf). The quality of the obtained DNA was analysed using the absorbance readings in a nanodrop. A₂₆₀/A₂₈₀ ratio of 1.8 was considered of high quality and a total yield of 1 μg was considered fit for sequencing.

Sequencing of the isolated DNA

The analysis of the sequenced fragments was done by Avant 3100 Gene Analyzer. DNA sequences were determined by the chain termination method using an ABI Prism Dye Terminator Cycle Sequencing kit (Applied Biosystems). Gene sequence of closely related species retrieved from GenBank was used to compare the 16S rRNA gene sequence of strain.

Analysis of the metagenome sequences

The tool used for comparison of 16S rRNA analysis of organisms isolated from palmyra sap sample and classification using the Quantitative Insights into Microbial Ecology (QIIME) software package (Cole et al. 2014). The gene sequences of 16S rRNA reads from palmyra sample sap metagenomes were compared with the computational resources and speed of q2-feature-classifier **QIIME** generating the three main 16S rRNA

databases: Green genes, SILVA, and RDP.

Result and Discussion

A metagenomic approach was used to find out the phylogenetic relationships of the sequenced gene with taxonomic groups of microorganisms known in the database. In this case, phylogenetic clusters like the 16S rRNA gene sequence are targeted where operational taxonomic units (OTUs) (Franzén et al. 2015) are compared against their amplitude to estimate the microbial species abundance in that particular environment (Edgar, 2018). Probiotics, which are live beneficial bacteria or yeasts, have gained significant attention in recent their numerous healthyears due to promoting properties, such as competitive inhibition of pathogenic bacteria, immune response regulation, assistance in nutrient metabolism, and even improvement of cognitive function (Rianda al.2019). In the past decades, probiotic research has rapidly increased due to the improvement of multi-omics approaches such culturomics, genomics, and



transcriptomics (Rebollar et al. 2016). Palmyra palm sugar is a local beverage produced by palmyra palm (Borassus flabellifer Linn.). The juice is popular in tropical Asian countries, including the south of Thailand (Naknean et al. 2010). The juice and products from palmyra palm sugar are an important source of natural bacteria, including LAB. Lactic acid bacteria isolated from fermented palmyra palm sap are considered to be probiotics. (Sornsenee et al. 2021). Several studies have investigated the diversity of Lactobacillus species present in palmyra sap. For instance, study conducted by Mitsuwan et al. (2022) characterized different lactic acid bacteria species from palmyra sap collected from 30 Palmyra sugar samples. The study identified three different lactobacillus species, namely Lactobacillus plantarum, Lactobacillus brevis, and Lactobacillus fermentum, from the palmyra sap samples. Das and Tamang (2021) carried out computational analysis of metagenomes based on KEGG and MetaCyc databases in the fresh palm saps and

fermenting saps during toddy fermentation and found the presence of Firmicutes (78.25%) was the most abundant phylum, followed Proteobacteria (21.57%).by Leuconostoc was the most abundant genus in the early stages of fermentation. Torulaspora, Lachancea, and Starmerella showed their heterogeneous distribution throughout the fermentation. Another study conducted by Pammi et al. (2021) also investigated the Lactobacillus diversity in palmyra sap from various regions in Tamil Nadu. The study identified four indigenous lactic acid bacteria strains, mainly belonging to the Lactobacillus plantarum group. Prathiviraj et al. (2022) revealed the bacterial diversity and metabolic profile of Indian palm wine using next-generation sequencing of the V3-V4 regions of the 16S rRNA gene. The metagenomic sequencing reveals the dominance of the phyla Proteobacteria, Firmicutes, and Tenericutes in the Indian palm wine.



The progress of research on metagenomics over the years aims in understanding the genome of uncultured microbes isolated from the environment. In the present study, the metagenome workflow was based on culture-based profiling of the bacteria present in the palmyra sap samples collected from two different localities, viz., Kanyakumari and Tirunelveli District. This study represents the experimental data generation and analysis of the 16S rDNA-based metagenome of the DNA extracted from the Palmyra sap samples for metagenomics analysis. culture-independent For metagenomics profiling of Palmyra sap samples, the region of 16SrRNA was amplified for the selected Palmyra sap

samples collected from Kanyakumari and Tirunelveli Districts. After pre-processing the raw data, the 16S amplicon analysis revealed 129,128 raw reads, 77,595,585 raw total bases, 77.60 raw data's in MB, 88,613 high-quality sequence reading region (HQ reads), 39,015,757 HQ total bases and 39.02 HQ Data in MB in palmyra sap samples isolated from Kanyakumari District and the 16s Amplicon analysis of the palmyra sap samples isolated from Tirunelveli District represent 127,120 raw reads, 76,583,582 raw total bases, 76.60 raw data's in MB, 87,632 high-quality sequence reading region (HQ reads), 38,023,768 HQ total bases and 38.02 HQ data. The result of the data statistics was represented in Table 1.

Table1.Data Statistics of 16s Amplicon analysis of probiotic bacteria present in the selected Palmyra samples.

| Palmyra Sap Sample | Kanyakumari District | Tirunelveli District |
|--------------------|----------------------|----------------------|
| Raw Reads | 129,128 | 127,120 |
| Raw Total Bases | 77,595,585 | 76,583,582 |
| Raw Data in MB | 77.6 | 76.6 |
| HQ Reads | 88,613 | 87.632 |
| HQ Total Bases | 39,015,757 | 38.023,768 |
| HQ Data in mb | 39.02 | 38.02 |

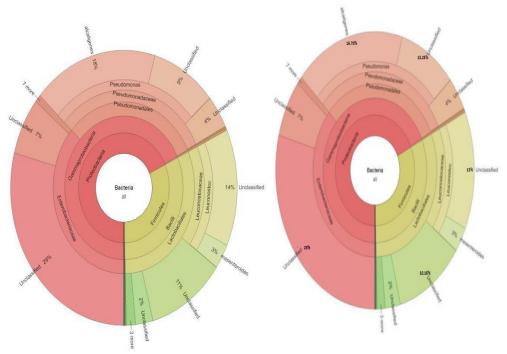


Krona Craft

The Krona chart was a multilayered pie chart that represented the hierarchical classification of probiotic bacteria present in the Palmyra sap samples collected from Kanyakumari and Tirunelveli Districts based on the phylum level, class level, order level, family level, genus level, and species level distribution. Krona Craft displayed the multilevel distribution of probiotic bacteria

present in the Palmyra sap sample collected from Kanyakumari and Tirunelveli District. The comparative results of the Krona craft revealed a slight variation in the percentage of probiotic bacteria distribution in the selected Palmyra sap samples. Figure 2 of Krona Craft displayed the multilevel distribution of probiotic bacteria present in the Palmyra sap sample collected from Kanyakumari District.

Fig. 2. Krona Craft displayed the multilevel distribution of the Probiotic bacteria isolated from Palmyra sap sample collected from Kanyakumari and Tirunelveli District



Finally, the metagenomic analysis based on the 16-s rRNA sequencing of the selected



palmyra sap sample reported the presence of different probiotic bacteria in the samples collected from Kanyakumari and Tirunelveli Districts at different hierarchical levels. Especially the phylum level, order level, class level, family level, genus level, and species level distribution. The overall metagenomics result of the Palmyra sample collected from sap Kanyakumari district distributed 233 OTUs and the most predominant taxonomy represented in the analysis was in the phylum level Proteobacteria, which was higher at 68.11%, followed by the class level Gammaproteobacteria at 67.83%, Enterobacteriales at 36.72% in the order level of 36.72% of Enterobacteriaceae at the family level, Klebsiella at 29.08% in the genus level, and unclassified species from the genus *Klebsiella* at 29.08%. metagenomics result of the Palmyra sap

sample collected from Tirunelveli district distributed 229 OTUs and the most predominant taxonomy represented in the analysis was at the phylum level. Proteobacteria was higher at 65.14%, followed the class level by Gammaproteobacteria 65.23%, at Enterobacteriales at 34.52% in the order level, 34.52% of Enterobacteriaceae at the family level, Klebsiella at 28.28% in the genus level, and unclassified species from the genus *Klebsiella* at 28.28%. The overall results of metagenomic analysis revealed that the probiotic bacteria belongs to the genus Klebsiella were predominant in the selected Palmyra sap samples compared to other probiotic bacteria distributed. The overall distribution of probiotic bacteria in the Palmyra sap sample of Kanyakumari and Tirunelveli District was described in Table 2.

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Table 2. The Overall Hierarchical Level Distribution of Probiotic Bacteria isolated from the Palmyra sap sample of Tirunelveli District

| | KANYAKUMARI DISTRICT | TIRUNELVELI DISTRICT |
|----------------|------------------------------|-------------------------------|
| NUMBER OF OTUS | 229 | 233 |
| PHYLUM | Proteobacteria (65.14%) | Proteo bacteria (68.11%) |
| CLASS | Gammaproteobacteria (65.23%) | Gamma proteobacteria (67.83%) |
| ORDER | Enterobacteriales (34.52%) | Enterobacteriales (36.72%) |
| FAMILY | Enterobacteriaceae (34.52%) | Enterobacteriaceae (36.72%) |
| GENUS | Klebsiella (28.28%) | Klebsiella (29.08%) |
| SPECIES | Unclassified (28.28%) | Unclassified (29.08%) |

The comparative metagenomics study of probiotic bacteria in the Palmyra sap samples of Kanyakumari and Tirunelveli districts through 16S rDNA-based metagenome analysis revealed that the distribution percentage of probiotic bacteria was predominant in the Palmyra sap samples collected from Kanyakumari District compared to the Palmyra sap samples collected from Tirunelveli District.

Conclusion

Palmyra sap is a low-cost plantbased mild alcoholic drink for rural Indian people, which is traditionally prepared by natural fermentation of fresh palm saps. Our study revealed the co-existence of bacteria in different environmental conditions in natural fermentation of fresh palm saps. The present study has authenticated the ethnic of traditional knowledge people preserving the essential microbial resources during natural fermentation of fresh saps to mild-alcoholic drink. The predictive profiles of both functional bacterial communities collected from Tirunelveli and Kanyakumari District in Palmyra fermentation showed significant differences the microbial synthesis of in some secondary metabolites, vitamins and aromatic amino acids between the fresh palm saps and fermented Palmyra sap

(toddy), which predict the health-promoting benefits in the product.

References

- Á.M. Ortiz-Estrada, T. Gollas-Galván, L.R. Mart ínez-Córdova, M. Martínez-Porchas, Predictive functional profiles using metagenomic 16S rRNA data: a novel approach to understanding the microbial ecology of aquaculture systems, Rev. Aquacult., 11 (1) (2019), pp. 234-235, 10.1111/raq.12237.
- Cole, JR, Wang, Q, Fish, JA, Chai, B, McGarrell, DM, Sun, Y, Brown, CT, Porras-Alfaro, A, Kuske, CR & Tiedje, JM 2014, 'Ribosomal database project: data and tools forhigh throughput rRNA analysis. Nucleic Acids', Res., vol. 42,pp. 633–42.
- D. Karamoko, N.T. Djeni, K.F. N'gu essan, K.P. Bouatenin, K.M. Dje, The biochemical and microbiological quality of palm wine samples produced at different periods during tapping and changes which occurred during their storage, Food Contr., 26 (2) (2012), pp. 504 511, 10.1016/j.foodcont.2012.02.018.
- DebMandal, M. and Mandal, S.

- (2011). Coconut (*Cocos nucifera* L.: Aceraceae): in health promotion and disease prevention. Asian Pac. J. Trop. Med. 4, 241–247. doi: 10.1016/S1995-7645(11)60078-3.
- Edgar, RC 2018, 'Accuracy of taxonomy prediction for 16S rRNA and fungal ITS sequences', PeerJ,vol. 6, p. 4652.
- Franzen, O, Hu, J, Bao, X, Itzkowitz, SH, Peter, I & Bashir, A 2015, 'Improved OTU-picking using long-read 16S rRNA gene amplicon sequencing and generic hierarchical clustering', Microbiome, vol. 3, p.43.
- H.K. Kouamé, M.D.F. Aké, N.M.C. Assohoun, M.K. Djè, N.T. Djéni, Dynamics and species diversity of lactic acid bacteria involved in the spontaneous fermentation of various palm tree saps during palm wine tapping in Côte d'Ivoire, World J. Microbiol. Biotechnol. 36 (2020), p. 64, 10.1007/s11274-020-02832-3.
- K.B. Hebbar, M. Arivalagan, M.R. Manikantan, A.C. Mathew, C. Tham ban, G.V. Thomas, P. Chowdappa, Coconut inflorescence sap and its value addition as sugar – collection techniques, yield, properties and



market perspective, Current Science, 109 (8) (2015), pp. 1-7, 10.18520/v109/i8/1411-1417.

- Kovoor A. The Palmyrah palm: potential and perspectives (FAO plant production and protection paper) Rome: FAO; 1983.
- Lasekan, O. and Abbas, K. A. (2010).
 Flavour chemistry of palm toddy and palm juice: a review. Trends Food Sci. Technol. 21, 494–501.
 doi: 10.1016/j.tifs.2010.07.007
- Mitsuwan, W, Sornsenee, P &Romyasamit, C 2022, 'Lacticaseibacillus spp.; Probiotic candidates from Palmyra palm sugar possesses antimicrobial and antibiofilm activities against methicillinresistant Staphylococcus aureus', Vet World,vol. 15, no. 2, pp. 299-308.
- Naknean, P, Meenune, M & Roudaut, G 2010, 'Characterization of palm sap harvested in Songkhla province, Southern Thailand', International Food Research Journal, vol. 17, pp. 977-986.
- Pammi, N, Bhukya, KK, Lunavath,
 RK &Bhukya, B2021,
 Bioprospecting of Palmyra Palm
 (Borassusflabellifer) Nectar:
 Unveiling the Probiotic and

- Therapeutic Potential of the Traditional Rural Drink', Front Microbiol, vol. 12, p. 683996.
- Prasad, G, Jamkhande, Vikas, A,Suryawanshi,Tukaram,M,Kaylank ar &Shailesh L Patwekar 2016, 'Biological activities of leaves of ethnomedicinal plant, **Borassus** flabellifer Linn. (Palmyra palm): An antibacterial, antifungal and antioxidant evaluation', Bulletin of of Pharmacy, Faculty Cairo University, vol. 54, no. 1, pp. 59-66.
- Prathiviraj, R, Dr, Rajeev, Riya, Jose, Chris, Begum, Ajima, Selvin, Joseph, Kiran &Seghal 2022, 'Fermentation microbiome and metabolic profiles of Indian palm wine', Gene Reports, vol. 27.
- Rebollar, EA, Antwis, RE, Becker, MH, Belden, LK, Bletz, MC. Brucker, RM, Harrison, XA, Hughey, MC, Kueneman, JG & Loudon, AH2016, 'Using "omics" and integrated multi-omics approaches to guide probiotic selection to mitigate chytridiomycosis and other emerging infectious diseases', Front. Microbiol, vol. 7, p. 68.
- Rianda, D, Agustina, R, Setiawan, EA &Manikam, NRM 2019,Effect of probiotic supplementation on cognitive function in children and



adolescents: A systematic review of randomised trials, Benef. Microbes, vol. 10, pp. 873–882.

- Saadat, Y. R., Khosroushahi, A. Y., and Gargari, B. P. (2019). A comprehensive review of anticancer, immunomodulatory and health beneficial effects of the lactic acid bacteria exopolysaccharides. Carbohydr. Polym. 217, 79–89. doi: 10.1016/j.carbpol.2019.04.025.
- Sornsenee, P, Singkhamanan, K, Sangkhathat, S, Saengsuwan, P &Romyasamit, C 2021, 'Probiotic Propertiesof*Lactobacillus*species isolated from fermented palm sap in Thailand', Probiotics Antimicrob. Proteins, vol. 13, no. 4, pp. 957 969.
- Srinivash, M., Krishnamoorthi, R.,
 Mahalingam, P. U., Malaikozhundan,
 B., and Keerthivasan, M. (2023).
 Probiotic potential of exopolysaccharide producing lactic acid bacteria isolated from

homemade fermented food products. J. Agric. Food Res. 11, 100517.

doi: 10.1016/j.jafr.2023.100517.

- T. Sarkar, M. Mukherjee, S. Roy, R. Chakraborty, Palm sap an unconventional source of sugar exploration for bioactive compounds and its role on functional food development, Heliyon, 9 (2023), p. e14788, 10.1016/j.heliyon.2023.e 14788
- Tamang, J. P., Cotter, P. D., Endo,
 A., Han, N. S., Kort, R., Liu, S. Q.,
 (2020). Fermented foods in a global age: East meets West. Compr. Rev.
 Food Sci. Food Saf. 19, 184–217.
 doi: 10.1111/1541-4337.12520.
- Vengaiah P.C., Murthy G.N., Prasad K. R., Kumari K.U. (2012). Post harvest technology of Palmyra (*Borassusflabellifer* L.) present practices and scope. International conference on food processing by Omics group, India.