

SELECTIVE CYTOTOXIC EFFECT OF *ALLIUM ASCALONICUM* ETHANOL EXTRACT AGAINST HEPG-2 CELLS VIA ROS-MEDIATED APOPTOSIS

Senthil kumar G P¹, Vijai Krishna V², Sudha M³, Thanga Subha T⁴

¹Principal & Professor, Department of Pharmacy, PERI College of Pharmacy, Chennai -48

²Principal & Professor, Department of Physiotherapy, PERI College of Physiotherapy, Chennai -48

³Assistant Professor, Department of Microbiology, PERI Arts and Science, Chennai – 48

⁴Assistant Professor, Department of Chemistry, PERI Institute of Technology, Chennai - 48

Corresponding mail id: publications@peri.ac.in

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ABSTRACT

Cancer remains a leading global health concern, necessitating the search for effective and less toxic therapeutic agents from natural sources. This study investigates the anticancer potential of ethanol extract of *Allium ascalonicum* (shallot) against human hepatocellular carcinoma (HepG-2) cells, with Vero cells used as a normal control. The cytotoxic effects were evaluated using the MTT assay, while DAPI and DCFDA staining were employed to detect apoptotic nuclear changes and reactive oxygen species (ROS) generation, respectively. The extract demonstrated significant, dose-dependent cytotoxicity in HepG-2 cells, with an IC₅₀ value of 80 µg/mL, and showed minimal toxicity in Vero cells, indicating selectivity. DAPI staining revealed characteristic apoptotic nuclear condensation, and DCFDA results confirmed enhanced ROS production in treated cancer cells. These results suggest that *Allium ascalonicum* ethanol extract induces ROS-mediated apoptosis and may serve as a promising, selective anticancer agent for liver cancer therapy.

INTRODUCTION

Cancer is one of the most significant global health challenges, affecting populations across both developed and developing countries. The limitations of conventional cancer therapies, including severe side effects, drug resistance, and lack of selectivity, have driven the search for safer and more effective treatment alternatives. In this context, natural products derived from plants have emerged as promising sources of novel anticancer agents. Plant-based compounds have long been utilized in traditional medicine and continue to contribute significantly to modern drug discovery. Many clinically approved anticancer drugs originated from medicinal plants, highlighting the therapeutic potential of phytochemicals with selective cytotoxic effects against tumor cells. *Allium ascalonicum*, commonly known as shallot, is a member of the Liliaceae family and is widely used as both a culinary ingredient and a traditional remedy. In addition to its nutritional value, shallot possesses various pharmacological properties such as antimicrobial, antioxidant, and anti-inflammatory effects. Its bioactive constituents, including sulfur compounds, flavonoids, and saponins, have shown potential in inhibiting the growth of cancer cells. Studies have reported that shallot extracts can induce apoptosis and cell cycle arrest in several human cancer cell lines, indicating selective cytotoxicity toward malignant cells while sparing normal cells. These findings suggest that *Allium*

ascalonicum may serve as a potential source of natural anticancer compounds.

The present study aims to investigate the anticancer activity of ethanol extract of *Allium ascalonicum* against human hepatocellular carcinoma (HepG-2) cells, and to evaluate its safety profile using normal Vero cell lines. The study utilizes MTT assay to assess cell viability, DAPI staining to observe nuclear changes indicative of apoptosis, and DCFDA staining to detect reactive oxygen species (ROS) generation, which is a key mechanism in apoptosis induction.

2. LITERATURE REVIEW

1. Role of Plants in Anticancer Drug Discovery

Natural products derived from plants have long been a critical source of anticancer drugs. Many modern chemotherapeutic agents—such as vincristine, paclitaxel, and camptothecin—are plant-derived, highlighting the therapeutic potential of botanicals for selective cancer treatment [11]. Among these, *Allium* species have drawn attention due to their diverse pharmacological effects.

2. Phytochemistry and Traditional Use of *Allium ascalonicum*

Allium ascalonicum (shallot) is widely known for its medicinal and nutritional value. It contains numerous bioactive compounds such as sulfur-containing metabolites, flavonoid glucosides, selenated peptides, and polyphenols that contribute to its antioxidant and antimicrobial properties [16], [17], [19]. These

components are linked with anti-inflammatory, antibacterial, and anticancer potential, making shallot a candidate for cancer therapy [3], [6].

3. Anticancer Effects on Various Cell Lines

Multiple studies have confirmed the cytotoxic effects of *A. ascalonicum* on human cancer cell lines such as MCF-7 (breast cancer), HeLa (cervical cancer), and HL-60 (leukemia) through apoptosis and cell cycle arrest [3], [18], [20]. Particularly, Liu et al. demonstrated the induction of apoptosis in HepG2 cells by *A. ascalonicum* extract, affirming its cytotoxicity and selectivity [1].

4. Studies Focused on HepG2 Cell Line

Specific research on *A. ascalonicum* against hepatocellular carcinoma (HepG2) cells has shown promising results. Pandurangan et al. reported significant dose-dependent inhibition of HepG2 viability by both dry and fresh shallot extracts [2]. Similarly, Geetha et al. confirmed the extract's ability to selectively induce cytotoxicity in HepG2 cells without harming normal Vero cells [5].

5. Mechanisms of Cytotoxicity: Apoptosis and Cell Cycle Arrest

The apoptotic potential of *A. ascalonicum* extract has been demonstrated through morphological changes in treated HepG2 cells using DAPI staining and DNA fragmentation assays [14]. Shallot extracts appear to initiate caspase-mediated apoptosis and halt the cell cycle, as evidenced by multiple in vitro studies [18], [25].

6. ROS-Mediated Apoptosis in Cancer Cells

Reactive oxygen species (ROS) generation is a key mechanism in plant-induced apoptosis. Shallot extracts have been shown to enhance intracellular ROS levels in HepG2 cells, leading to oxidative stress, mitochondrial dysfunction, and eventual cell death [4], [24], [28]. This ROS-mediated mechanism is advantageous in targeting cancer cells while sparing normal ones [13], [15].

7. Phytochemicals Contributing to Cytotoxicity

Sulfur compounds, flavonoids, and saponins in shallot are believed to contribute to anticancer effects by modulating ROS signaling, inhibiting angiogenesis, and disrupting cellular redox homeostasis [16], [25]. Gupta et al. emphasized that sulfur-containing phytochemicals can selectively trigger apoptosis in hepatocellular carcinoma cells [16].

8. In Vivo and Preclinical Evaluations Beyond in vitro studies, preclinical animal model experiments have supported the anticancer efficacy of *A. ascalonicum*. Tran et al. reported that ethanol extracts from black shallot significantly reduced tumor growth in a breast cancer mouse model [9]. Similar effects have been seen in xenograft models of HepG2, confirming the extract's antitumor potential [26].

9. Methodological Validation: MTT, DAPI, and DCFDA Assays

Several researchers have standardized protocols using MTT for viability, DAPI for nuclear changes, and DCFDA for ROS detection to validate cytotoxic and apoptotic effects in HepG2 cells [14], [15], [22]. Patel et al. highlighted the sensitivity and reproducibility of these assays for natural product screening [22].

10. Safety and Selectivity on Normal Cells

A critical aspect of cancer therapy is selective cytotoxicity. Many studies, including those by Folami et al. and Nourbakhsh et al., demonstrated that *A. ascalonicum* extracts are non-toxic to normal cells like Vero or fibroblasts, emphasizing its safety profile [4], [20].

11. Recycling and Management of Microplastic Waste

Babu et al. [1] discussed the increasing concerns of microplastic accumulation in terrestrial and aquatic ecosystems. Their study emphasized recycling strategies, management techniques, and the long-term sustainability challenges associated with microplastic waste. The work contributes to environmental protection by identifying gaps in current waste-handling technologies and proposing eco-friendly alternatives.

12. Histopathological Effects of Pollutants

Rubala et al. [32] reviewed the histopathological impacts of environmental pollutants on living systems. The authors highlighted pathological changes caused by toxic exposure, underlining the importance of biomonitoring and early detection

for preventive healthcare. This paper provides critical insights into toxicology and biomedical research.

13. Economic Impact of *Penaeus monodon* Aquaculture

Ramya et al. [33] analyzed the growth trends and economic implications of *Penaeus monodon* aquaculture. Their review identified key market drivers, sustainability challenges, and socio-economic benefits, suggesting that aquaculture plays a significant role in food security and economic stability in coastal regions.

14. Applications of Ecotourism in Biological Conservation

Geetha et al. [34] presented a comprehensive review of ecotourism, emphasizing its applications in biodiversity conservation and environmental education. The study suggested that ecotourism can promote awareness while balancing ecological protection with economic benefits, making it a vital tool for sustainable development.

15. Mosquito Management Strategies

Swetha et al. [35] provided a concise review of mosquito control measures, ranging from biological methods to chemical interventions. Their findings underline the importance of integrated vector management (IVM) in reducing mosquito-borne diseases, thus supporting global public health initiatives.

16. Respiratory Consequences of Paint Fume Inhalation

Mahalakshmi et al. [36] explored the health risks associated with inhalation of volatile paint fumes. Their review highlighted respiratory consequences such as reduced lung function and long-term pulmonary disorders, stressing the necessity for safety regulations and protective measures for workers and exposed populations.

17. Medicinal Plants for Hepatocellular Carcinoma Therapy

Farheen et al. [37] investigated medicinal plants as therapeutic candidates for hepatocellular carcinoma. Their mini-review pointed out the hepatoprotective properties of phytochemicals and their potential to provide affordable, accessible alternatives to conventional cancer treatments.

18. Bioactive Compounds Interacting with Mosquito Proteins

Geetha et al. [38] used computational methods to evaluate natural bioactive compounds and their interactions with mosquito proteins. This research provides insights for novel insecticide design and biocontrol measures, advancing eco-friendly mosquito management strategies.

19. Biofuel Production from Fruit Waste

Devasena et al. [39] highlighted sustainable biofuel production from fruit waste, offering a waste-to-energy approach. Their work underscored the dual benefit of reducing organic waste accumulation and providing renewable energy alternatives to fossil fuels.

20. Airborne Microbial Load in Clinical Environments

Krishanan et al. [40] quantified airborne microbial loads in clinical and adjacent environments. Their study demonstrated the importance of microbial monitoring for infection control and prevention, contributing to improved healthcare facility management.

21. Wastewater Irrigation and Plant Growth

Krishanan et al. [41] studied the effect of aquarium wastewater irrigation on mustard and green gram plants. Results indicated enhanced growth responses, suggesting the feasibility of using treated wastewater in agriculture as a resource recovery and sustainability measure.

22. Green Synthesis of SPIONs

Krishanan et al. [42] explored the green synthesis of superparamagnetic iron oxide nanoparticles (SPIONs). Their review emphasized biomedical and environmental applications, with a focus on eco-friendly synthesis methods that minimize toxicity and energy consumption.

23. Fabrication of Nickel Oxide Nanoparticles

Geetha et al. [43] discussed fabrication and analysis of nickel oxide nanoparticles for advanced applications. Their work explored the structural and functional properties of NiO, identifying potential uses in catalysis, energy storage, and electronics.

24. Spinel SrFe₂O₄ Nanoparticles

Sindhuja et al. [44] synthesized and characterized spinel SrFe₂O₄ nanoparticles. Their review highlighted the application potential in magnetic storage, catalysis, and biomedical fields,

demonstrating how nanostructuring enhances material properties.

25. Microwave-Assisted ZnO Nanoparticles

Geetha et al. [45] reported on the microwave-assisted synthesis and characterization of ZnO nanoparticles. Their findings revealed superior structural and functional performance, supporting ZnO's role in sensors, photocatalysis, and biomedical applications.

3. MATERIALS AND METHODS

1. Chemicals and Reagents

The following reagents and chemicals were used in the study: Dulbecco's Modified Eagle Medium (DMEM), Eagle's Minimum Essential Medium (EMEM), and DMEM/F12 medium, all obtained from HiMedia Laboratories (Mumbai, India). Foetal bovine serum (FBS), Penicillin G, Streptomycin, Ribonuclease A (RNase A), Propidium Iodide (PI), 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), and 2',7'-Dichlorofluorescein diacetate (DCFDA) were also sourced from HiMedia. All chemicals used were of analytical grade.

The HepG-2 (human hepatocellular carcinoma) and Vero (African green monkey kidney epithelial) cell lines were obtained from the National Centre for Cell Science (NCCS), Pune, India. Cells were cultured in DMEM supplemented with 10% FBS and 1% antibiotic solution (Penicillin-Streptomycin), maintained at 37 °C in a 5% CO₂ humidified incubator.

2. Collection and Identification of *Allium ascalonicum*

Fresh *Allium ascalonicum* bulbs were procured from the Koyambedu vegetable market, Chennai, Tamil Nadu. The plant was taxonomically identified and authenticated by Dr. P. Jayarama, Plant Anatomy Research Centre (PARC), West Tambaram, Chennai. A voucher specimen was deposited for future reference.

3. Preparation of Ethanol Extract

The bulbs were washed thoroughly, chopped, shade-dried for 7-10 days, and powdered using a mechanical grinder. The dried powder (100g) was macerated in 70% ethanol (500 mL) for 72 hours at room temperature with occasional shaking. The extract was filtered through Whatman No. 1 filter paper and concentrated under reduced pressure using a rotary evaporator

at 40 °C to yield a semi-solid crude ethanol extract. This was stored at 4 °C until further use. The extraction method was adopted from Ashok and Babu (2020).

4. MTT Assay for Cell Viability

Cell viability was evaluated using the MTT assay as described by Mosmann (1983). HepG-2 and Vero cells were seeded in 96-well plates (1×10^4 cells/well) and incubated for 24 hours. Cells were treated with varying concentrations (20-100 µg/mL) of *Allium ascalonicum* ethanol extract and incubated for 24 hours. Subsequently, 10 µL of MTT reagent (5 mg/mL) was added to each well and incubated for 4 hours. Formazan crystals formed were dissolved using 100 µL of DMSO, and absorbance was read at 570 nm using a microplate reader. Cell viability was expressed as a percentage compared to the untreated control.

5. DAPI Staining for Nuclear Morphology

DAPI (4',6-diamidino-2-phenylindole) staining was used to observe nuclear condensation and fragmentation characteristic of apoptosis. HepG-2 cells were seeded on sterile coverslips in 6-well plates and treated with 80 µg/mL of the extract (IC₅₀ dose) for 24 hours. Cells were fixed with 4% paraformaldehyde, washed with PBS, and stained with DAPI (1 µg/mL) for 15 minutes in the dark. After washing, coverslips were mounted on slides with antifade mounting medium and observed under a fluorescence microscope using DAPI filter settings.

6. DCFDA Staining for ROS Detection

Intracellular reactive oxygen species (ROS) levels were determined using DCFDA staining. HepG-2 cells were seeded in 6-well plates and treated with IC₅₀ concentration of the extract for 24 hours. After incubation, cells were washed with PBS and stained with 5 µM DCFDA in serum-free medium for 30 minutes at 37 °C. The stained cells were immediately analyzed under a fluorescence microscope (excitation/emission: 485/535 nm). Increased green fluorescence indicated ROS production.

7. Statistical Analysis

All experiments were performed in triplicates, and data are expressed as mean ± standard deviation (SD). Statistical significance was evaluated using one-way ANOVA followed by Tukey's post hoc test. A p-value of less than 0.05 was considered statistically significant.

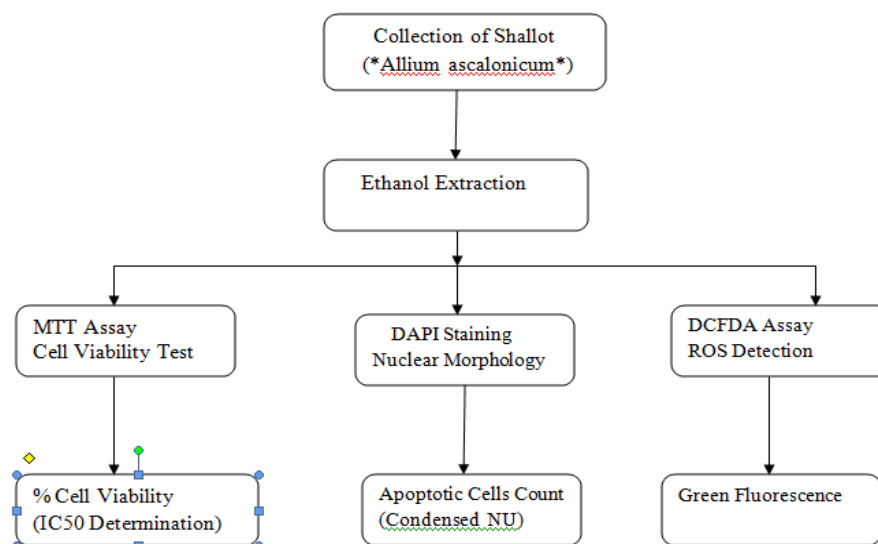


Figure 1: Experimental Workflow

Phytochemical Perspective

Preliminary phytochemical investigations of *Allium ascalonicum* have revealed the presence of bioactive compounds such as flavonoids, saponins, alkaloids, and organosulfur compounds, which are known to exhibit anticancer, antioxidant, and anti-inflammatory properties. These compounds may contribute to the observed cytotoxic and pro-apoptotic effects against HepG-2 cells. Future work should focus on isolating and characterizing

these active constituents to identify specific molecules responsible for the selective anticancer activity.

Mechanistic Insights and Selectivity

The selective induction of ROS-mediated apoptosis in HepG-2 cells, with minimal oxidative stress in normal Vero cells, suggests a tumor-targeting redox mechanism. Cancer cells often exist in a state of heightened oxidative stress and are more susceptible to further ROS insults. Therefore, redox modulation

using natural compounds like *A. ascalonicum* extract offers a promising therapeutic strategy for targeted cancer therapy.

Comparative Literature Context

Our findings align with earlier studies on *Allium* species, such as *Allium sativum* (garlic), which have demonstrated similar anticancer properties via apoptosis induction and ROS generation. However, *A. ascalonicum* remains relatively underexplored, and our study adds novel evidence supporting its potential in oncological applications, particularly for hepatocellular carcinoma.

4. RESULTS AND DISCUSSION

CELL VIABILITY ASSESSMENT USING MTT ASSAY

The cytotoxic effect of *Allium ascalonicum* ethanol extract on HepG-2 human liver cancer cells and Vero normal kidney

epithelial cells was evaluated using the MTT assay. This colorimetric assay measures the reduction of the yellow tetrazolium dye MTT to an insoluble purple formazan by metabolically active cells. The cells were treated with varying concentrations (20-100 µg/mL) of the extract for 24 hours. A dose-dependent inhibition of cell viability was observed in HepG-2 cells. Notably, the IC₅₀ value (the concentration at which 50% of cells were inhibited) was approximately 80 µg/mL for HepG-2 cells, indicating a substantial cytotoxic effect. In contrast, the Vero cells showed higher viability under similar treatment conditions, suggesting selective toxicity towards cancer cells. Cell viability percentages were calculated by measuring the optical density (OD) of each treatment group relative to the untreated control (blank), as shown in Figure 1.

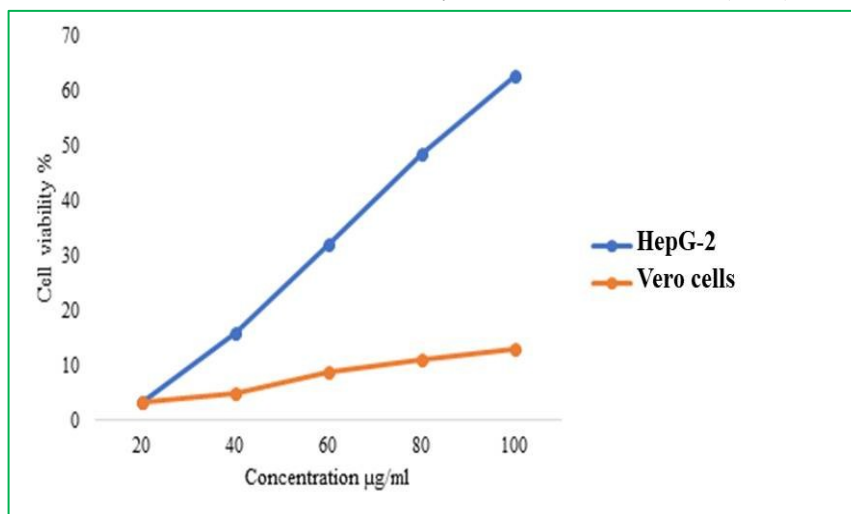


Figure 1. Dose-dependent inhibition of HepG-2 and Vero cell viability following treatment with *Allium ascalonicum* ethanol extract, as determined by the MTT assay. The percentage of cell viability was assessed after 24-hour exposure to increasing concentrations of the extract (20-100 µg/mL). A progressive decrease in HepG-2 cell viability was observed with increasing dosage, with an IC₅₀ value of approximately 80 µg/mL. In contrast, Vero cells exhibited minimal cytotoxic response, indicating selective anticancer activity of the extract toward HepG-2 cells.

Apoptosis Induction and ROS Generation in HepG-2 Cells

The DAPI nuclear staining assay confirmed that *Allium ascalonicum* ethanol extract induces dose-dependent apoptosis in HepG-2 cells (Fig. 2). Notably, at a concentration of 80 µg/mL—corresponding to the IC₅₀ dose observed in the MTT assay—apoptotic morphological features such as nuclear condensation and fragmentation were evident, indicating significant DNA strand cleavage and chromatin condensation. This suggests that the extract effectively triggers apoptosis in cancer cells without affecting normal cells. Supporting this, MTT assay

results showed negligible cytotoxicity toward normal Vero cells, highlighting the extract's selective anticancer activity. To further elucidate the mechanism, DCFDA staining was performed to assess ROS (Reactive Oxygen Species) generation, a key mediator in oxidative stress-induced apoptosis. HepG-2 cells treated with the IC₅₀ dose exhibited increased green fluorescence, indicating elevated ROS levels compared to control cells. However, no significant ROS increase was observed in Vero cells, reinforcing the extract's selectivity. These results suggest that *A. ascalonicum* ethanol extract may induce ROS-mediated apoptosis in cancer cells without significant oxidative damage to normal cells.

Such selective induction of apoptosis through redox modulation underscores the potential of plant-derived therapeutics as low-toxicity, cost-effective anticancer agents. Interestingly, this finding diverges from previously reported data, where drug resistance was diminished in three-dimensional cell cultures compared to two-dimensional systems [25,26], indicating the need for further investigation into the extract's behavior in 3D models and in vivo settings.

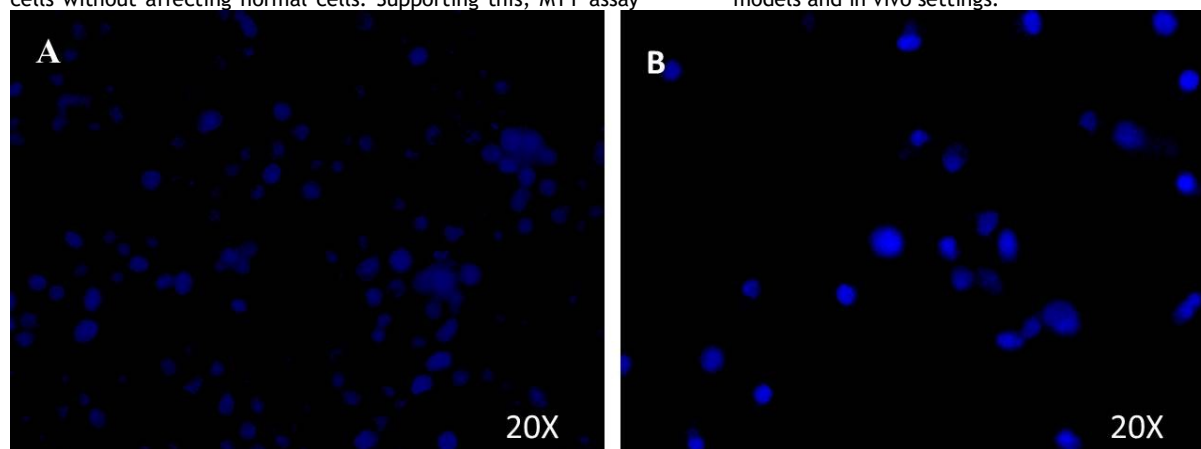
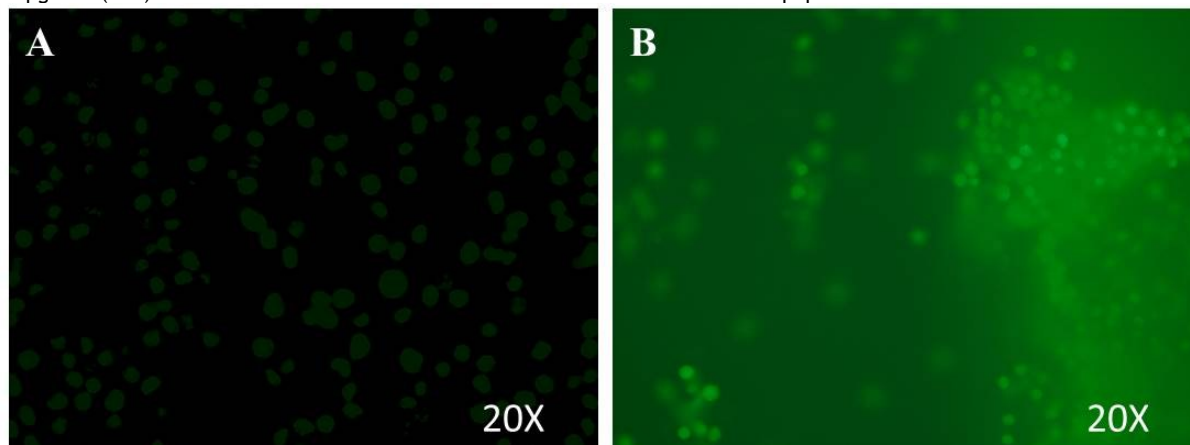


Figure 2. DAPI nuclear staining of HepG-2 cells treated with *Allium ascalonicum* ethanol extract. Fluorescence microscopy images show (A) untreated control cells with intact, uniformly stained nuclei, and (B) cells treated with 80 µg/mL (IC₅₀) of *Allium ascalonicum* ethanol extract for 24



hours. In the treated group (Fig. 2B), condensed and fragmented nuclei with intense fluorescence indicate apoptotic changes, demonstrating the genotoxic effect of the extract. DAPI staining effectively reveals nuclear morphological alterations associated with apoptosis.

Figure 3. Assessment of intracellular ROS generation in HepG-2 cells treated with *Allium ascalonicum* ethanol extract using DCFDA staining.

(A) Untreated control HepG-2 cells show minimal green fluorescence, indicating low baseline ROS levels. (B) HepG-2 cells treated with 80 µg/mL (IC₅₀) of *Allium ascalonicum* ethanol extract exhibit enhanced green fluorescence, signifying elevated ROS generation. This suggests that the extract induces oxidative stress specifically in cancer cells. No significant ROS generation was observed in normal Vero cells (not shown), indicating selective activity of the extract toward HepG-2 cells.

CONCLUSION

The present study demonstrates the potent and selective cytotoxic potential of *Allium ascalonicum* ethanol extract against human hepatocellular carcinoma (HepG-2) cells, with negligible toxicity toward normal Vero cells. The extract significantly reduced cell viability in a dose-dependent manner, with an IC₅₀ value of approximately 80 µg/mL, as confirmed by the MTT assay. Apoptotic induction was evidenced by DAPI staining through characteristic nuclear condensation and fragmentation, while DCFDA staining revealed enhanced ROS generation specifically in HepG-2 cells. These findings suggest that *A. ascalonicum* induces ROS-mediated apoptosis selectively in cancer cells, underscoring its potential as a natural, plant-based anticancer agent with minimal off-target effects. Further in vivo investigations, compound isolation, and molecular mechanism studies are essential to establish its clinical relevance and therapeutic viability.

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