

Epidemiological studies on the Incidence and Distribution of Leaf Spot Disease in Ashwagandha (*Withania somnifera*) Caused by *Alternaria alternata*

Sudhanshu Vats¹, Ram Suman Mishra², Abhishek Katiyar³

¹M.Sc., Department of Plant Pathology, Acharya Narendra Deva University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India.

² Associate Professor, Department of Plant Pathology, Acharya Narendra Deva University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India.

³Ph.D. Research Scholar, Department of Plant Pathology, Acharya Narendra Deva University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India.

Corresponding author Email: sudhanshuvats27@gmail.com

<https://doi.org/10.63001/tbs.2025.v20.i03.pp345-354>

Received on: 22-06-2025

Accepted on: 09-07-2025

Published on: 19-07-2025

ABSTRACT

Medicinal and aromatic plant *Withania somnifera* (Ashwagandha), is integral to Indian traditional medicine and the herbal pharmaceutical industry, owing to their rich content of bioactive compounds like alkaloids, glycosides, flavonoids. However, their commercial cultivation is severely impacted by fungal diseases, particularly foliar infections, which reduce yield, compromise quality, and alter the efficacy of pharmacologically important metabolites. Ashwagandha showed early disease symptoms in the 43rd SMW, with severity peaking at 38.77% in the 13th SMW due to rainfall and sustained humidity. In Ashwagandha, T9 achieved over 81% control for both metrics at early stages.

Introduction -

Epidemiology has made significant contributions to plant pathology by elucidating the general principles underlying the development of disease epidemics and provided analytical prospective of theoretical and empirical understanding of the dynamics of disease epidemics in time and space (Hughes and

Burnett, 2017), predictions of disease outbreaks or the need for disease control in real-time basis, and tactical and strategic solutions to disease problems (Ojiambo, 2017). Medicinal and aromatic plants constitute a major segment of the flora, which provides raw materials for use in the pharmaceuticals, cosmetics and drug

industries (Jeelani- *et. al.*2018). The global market for plant-based drugs is estimated at \$100billion, projected to reach \$5 trillion by 2050, it is because of about 80% of the population relying on herbal medicine (Andrea, 2011). Medicinal and aromatic plants are attacked by number of fungal, bacterial, viral and nematode pathogens which affect its growth as well as alter its therapeutics properties. Biotic pathogens are spread rapidly throughout the crop under wet and humid environmental conditions. Among the several diseases of Ashwagandha the leaf spot of is one of the devastating diseases.

Materials and Methods:

The research experiment entitled **“Epidemiological studies on disease occurrence in medicinal and aromatic plants especial reference to Basil , Kalmegh, Ashwagandha and Opium poppy”** was carried out at Medicinal and Aromatic Plant farm, Main Horticulture Experiment Station, Department of Plant pathology, Acharya Narendra Deva University of Agriculture and Technology, Kumarganj, Ayodhya (U.P.) India during, Kharif and *Rabi* 2024-25.

Collection of disease samples

The diseased samples were collected from the experimental site. Infected leaves were gathered in sterilized polybags. The infected parts showing the typical symptom of the associated was brought to the laboratory for isolation of the pathogens on PDA media or distilled water. The pure culture was maintained on PDA slant and distilled water for further studies.

Visual examination

On the basis of visual observations of plant, the typical symptoms of diseases on Kalmegh (leaf spot), Ashwagandha (leaf spot), Basil (downy mildew) and Opium (downy mildew) were recorded and noted in the field.

Disease incidence and severity

Weekly data of disease incidence and disease severity of leaf spot of Ashwagandhan was collected from 1st week of December to 2nd week of March in both the year.. The weekly disease incidence and severity data were recorded using 0 - 5 point scale after the appearance of downy mildew.

Downy mildew disease rating scale (Natti *et al.*, 1967)

Score/Grade	Description of disease symptoms
0	Cotyledons/ leaves free from infection
1	Small creamy white to light brown spots on cotyledons/leaves covering 1-10% area
2	Small creamy white to light brown spots with cottony growth on lower surface covering 10.1-25% cotyledons/leaves area
3	Creamy white to light brown spots with cottony growth covering 25.1-50% cotyledons/leaves area
4	Creamy white to light brown spots with cottony growth covering 50.1-75% cotyledons/leaves area
5	Creamy white to light brown spots with cottony growth covering >75% cotyledons/leaves area

The percent disease incidence was calculated using formula given by Ginting and Maryono, 2009.

$$\text{Percent Disease Incidence} = \frac{\text{Total no.of infected plant}}{\text{Total no.of plant planted}} \times 100$$

Disease severity was calculated on weekly intervals with the following formula given by Shutong *et. al.*, 2007.

$$\text{Percent Disease Severity} = \frac{\text{Sum of all numerical rating}}{\text{Number of plants scored} \times \text{maximum score on scale}} \times 100$$

Experimental Details.

Season	Kharif
Crop	Ashwagandha
Design	RBD
Replications	3
Treatment	10
Plot Size	3×2.40 m²
Spacing	50×30cm

Table.1 Treatment of Integrated Disease Management for downy mildew (Ashwagandha)

Treatment	
T₁	Seed treatment with <i>Pseudomonas fluorescence</i> (5g/kg) + <i>Trichoderma harzianum</i> (10g/kg) each + Soil application of FYM enriched with <i>Trichoderma</i> + <i>Pseudomonas</i> (2.5kg formulation in 400kg of FYM) + first foliar spray of jeevamrit @ 10% with the onset of disease followed by 2nd and 3rd at 15days interval.
T₂	Seed treatment with <i>Pseudomonas fluorescence</i> (5g/kg) + <i>Trichoderma harzianum</i> (10g/kg) each + Soil application of FYM enriched with <i>Trichoderma</i> + <i>Pseudomonas</i> (2.5kg formulation in 400kg of FYM) + first foliar spray of <i>Pseudomonas fluorescence</i> + <i>Trichoderma harzianum</i> each @ 1% onset of disease followed by two sprays at 15 days interval.
T₃	Seed treatment with <i>Pseudomonas fluorescence</i> (5g/kg) + <i>Trichoderma harzianum</i> (10g/kg) each + Soil application of FYM enriched with <i>Trichoderma</i> + <i>Pseudomonas</i> (2.5kg formulation in 400kg of FYM) + first foliar spray of Azadiractin 1500ppm @ 2.5 ml/liter onset of disease followed by two sprays at 15 days interval.
T₄	Seed treatment with <i>Pseudomonas fluorescence</i> (5g/kg) + <i>Trichoderma harzianum</i> (10g/kg) each + Soil application of FYM enriched with <i>Trichoderma</i> + <i>Pseudomonas</i> (2.5kg formulation in 400kg of FYM) + first foliar spray of Chitosan @0.1% with the onset of disease followed by two sprays at 15days interval.
T₅	Seed treatment with jeevamrit @10ml/l + Soil application of FYM enriched with <i>Trichoderma</i> + <i>Pseudomonas</i> formulation each (2.5kg formulation in 400kg of FYM) + first foliar spray of Azadiractin 1500ppm @ 2.5 ml/liter onset of disease followed by two sprays at 15 days interval.

T₆	Seed treatment with jeevamrit @10ml/l + Soil application of FYM enriched with <i>Trichoderma</i> + <i>Pseudomonas</i> formulation each (2.5kg formulation in 400kg of FYM) + first foliar spray of jeevamrit @10% with the onset of disease followed by 2nd and 3 rd at 15 days interval.
T₇	Seed treatment with jeevamrit @10ml/l + Soil application of FYM enriched with <i>Trichoderma</i> + <i>Pseudomonas</i> formulation each (2.5kg formulation in 400kg of FYM) + first foliar spray of Chitosan @0.1% with the onset of disease followed by two sprays at 15 days interval.
T₈	Seed Treatment with chitosan 0.1% + first foliar spray of Chitosan @ 0.1% with the onset of disease followed by two sprays at 15days interval.
T₉	Chemical control (50% Tebuconazole and 25% Trifloxystrobin)
T₁₀	Control

Percent Disease Incidence

$$\text{Percent Disease Incidence} = \frac{\text{Total no.of infected plant}}{\text{Total no.of rhizome planted}} \times 100$$

Percent Disease Severity

$$\text{Percent Disease Severity} = \frac{\text{Sum of all numerical rating}}{\text{Number of plants scored} \times \text{maximum score on scale}} \times 100$$

Results and Discussion:

Effect of Integrated disease management on leaf spot of Aswhagandha

Leaf spot of Aswhagandha

Experimental data revealed that all the treatments were significantly minimized the Leaf spot disease of aswhagandha (table -3). The highest disease incidence was recorded in T₁₀ (47.29%) followed by T₆ (36.39%) ,T₂ (33.34 %) and T₁ (30.49%) and lowest in T₉ (8.73%) followed by T₈ (10.11%), T₅ (10.89%) and T₄ (14.37%) at 30 days of transplanting(DAT). In case of 45 DAT, maximum disease incidence was recorded in T₁₀ (55.20%) followed by T₆ (45.44%) ,T₂ (43.99 %) and T₁ (43.63%) and lowest in T₉ (18.22%) followed by T₈ (23.28%), T₅ (27.44%) and T₄ (28.48%) at 60 days of transplanting (DAT). In case of 75 DAT, maximum disease incidence was recorded in T₁₀ (59.40%), followed by T₆ (53.88%) ,T₂ (51.44 %) and T₁ (49.75%) and lowest in T₉ (22.87%) followed by T₈(27.38 %), T₅ (31.10%) and T₄ (31.28%) .

Where as in disease severity it was found maximum in T₁₀ (23.05%) followed by T₆ (15.05%), T₂ (14.08%) and T₁ (13.15%) and minimum in T₉ (4.22%) followed by T₈ (5.89%), T₅ (6.19%) and T₄ (7.21%) at 30 days of transplanting (DAT). In case of 45 DAT, the highest disease severity T₁₀ (28.44%) followed by T₆ (20.75%), T₂ (20.35%) and T₁ (18.88%) and lowest in T₉ (8.06%) followed by T₈ (10.10%), T₅ (11.87%) and T₄ (12.77%). Disease severity it was found maximum in T₁₀ (33.77%) followed by T₆ 27.60%), T₂ (26.49%) and T₁ (25.08%) and minimum in T₉ (11.55%) followed by T₈ (14.66%), T₅ (16.22%) and T₄ (19.09%) at 60 days of transplanting (DAT). In case of 75 DAT, the highest disease severity T₁₀ (38.77%) followed by T₆ (32.09%), T₂ (30.00%) and T₁ (28.99%) and lowest in T₉ (16.38%) followed by T₈ (18.83%), T₅ (21.57%) and T₄ (24.80%). Sarkar (2021) reported that the highest disease (30.61%) was in control treatment and application of Carbendazim recorded the lowest percent disease incidence (14.75%) in comparison to control (74.99 %). These results are similar to previous researcher like as Mishra (2021) and Kathal (2023).

Table 2: Effect of Integrated disease management on leaf spot of Aswhagandha

Treatments	Disease appearance Days after transplanting	Percent Disease Incidence Days of transplanting (DAT)				Percent Disease Severity Days of transplanting (DAT)			
		30	45	60	75	30	45	60	75
T₁	30	30.49 (33.50)	37.55 (33.77)	43.63 (41.32)	49.75 (44.84)	13.15 (21.25)	18.88 (25.73)	25.08 (30.04)	28.99 (32.56)
T₂	30	33.34 (35.25)	39.85 (39.12)	43.99 (41.53)	51.44 (45.80)	14.08 (22.02)	20.35 (26.80)	26.49 (30.95)	30.00 (33.19)
T₃	31	26.56 (31.00)	31.20 (33.94)	38.67 (38.43)	45.68 (42.50)	12.70 (20.86)	17.42 (24.65)	21.25 (27.43)	25.83 (30.53)
T₄	32	14.37 (22.26)	21.43 (27.56)	28.48 (32.24)	31.28 (33.99)	7.21 (15.56)	12.77 (20.96)	19.09 (25.89)	24.80 (29.85)
T₅	32	10.89 (19.25)	20.22 (26.70)	27.44 (31.57)	31.10 (33.87)	6.19 (14.39)	11.87 (20.14)	16.22 (23.51)	21.57 (27.65)
T₆	30	36.39 (37.08)	40.77 (39.63)	45.44 (42.36)	53.88 (47.20)	15.05 (22.81)	20.75 (27.08)	27.60 (31.68)	32.09 (34.48)
T₇	30	29.82 (33.08)	35.29 (36.43)	41.01 (39.80)	46.87 (3.19)	13.02 (21.14)	17.72 (24.87)	23.05 (28.67)	28.43 (32.20)
T₈	32	10.11 (18.52)	18.89 (25.75)	23.28 (28.83)	27.38 (31.53)	5.89 (14.02)	10.10 (18.51)	14.66 (22.50)	18.83 (25.69)
T₉	34	8.73 (17.14)	11.88 (20.15)	18.22 (25.25)	22.87 (28.55)	4.22 (11.83)	8.06 (16.46)	11.55 (19.84)	16.38 (23.85)
T₁₀	30	47.29 (43.42)	49.93 (44.94)	55.20 (47.96)	59.40 (50.39)	23.05 (28.68)	28.44 (32.21)	33.77 (35.51)	38.77 (38.49)
C.D. (P=0.5%)		1.858	1.379	1.588	1.720	1.022	1.477	1.335	1.837
SE(m) ±		0.620	0.460	0.530	0.574	0.341	0.493	0.446	0.614
C.V. %		4.333	2.598	2.514	2.371	5.158	5.136	3.535	4.000

(Data in the parenthesis is transformed values)

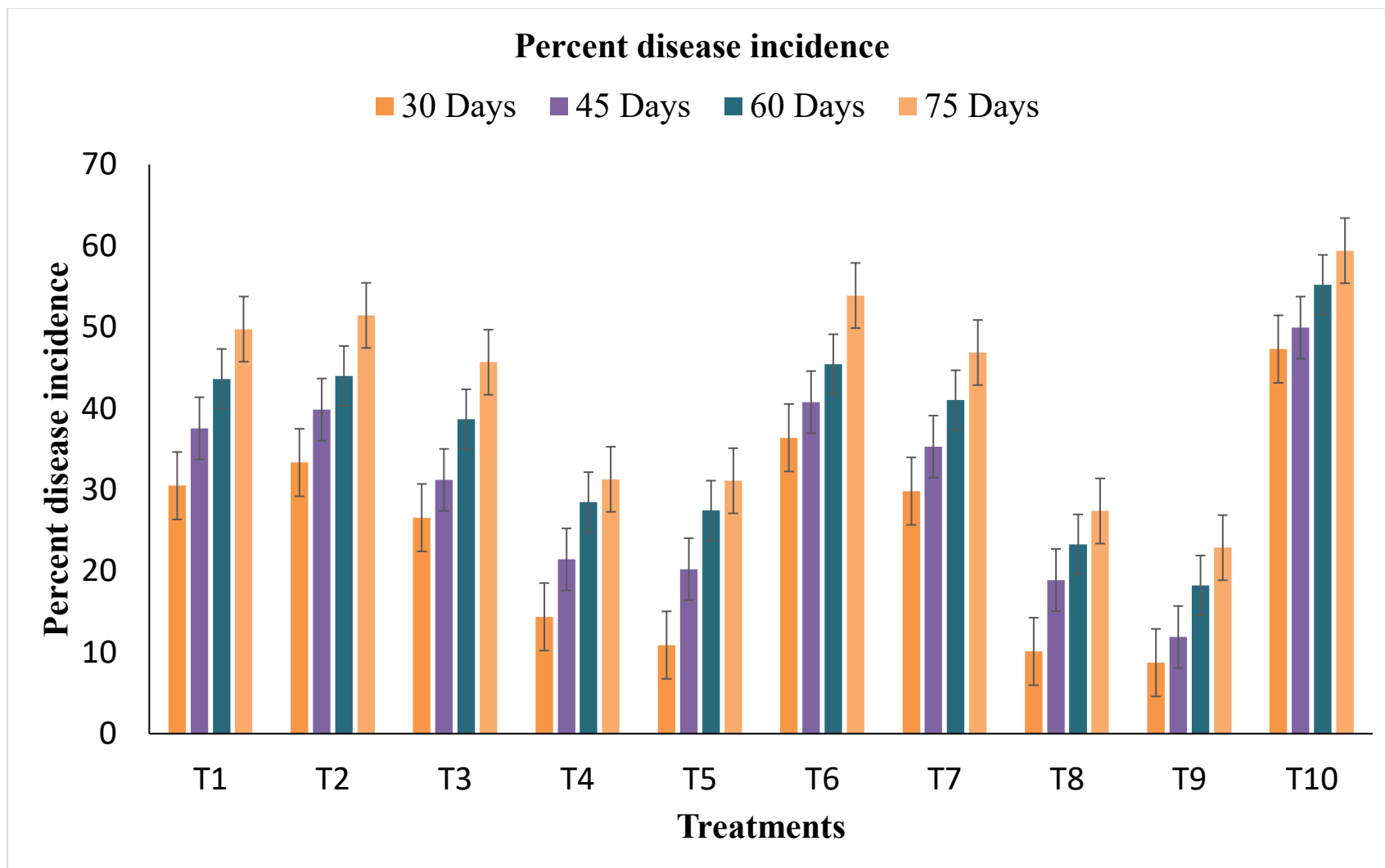


Fig 1: Effect of Integrated disease management on leaf spot of Ashwgandha

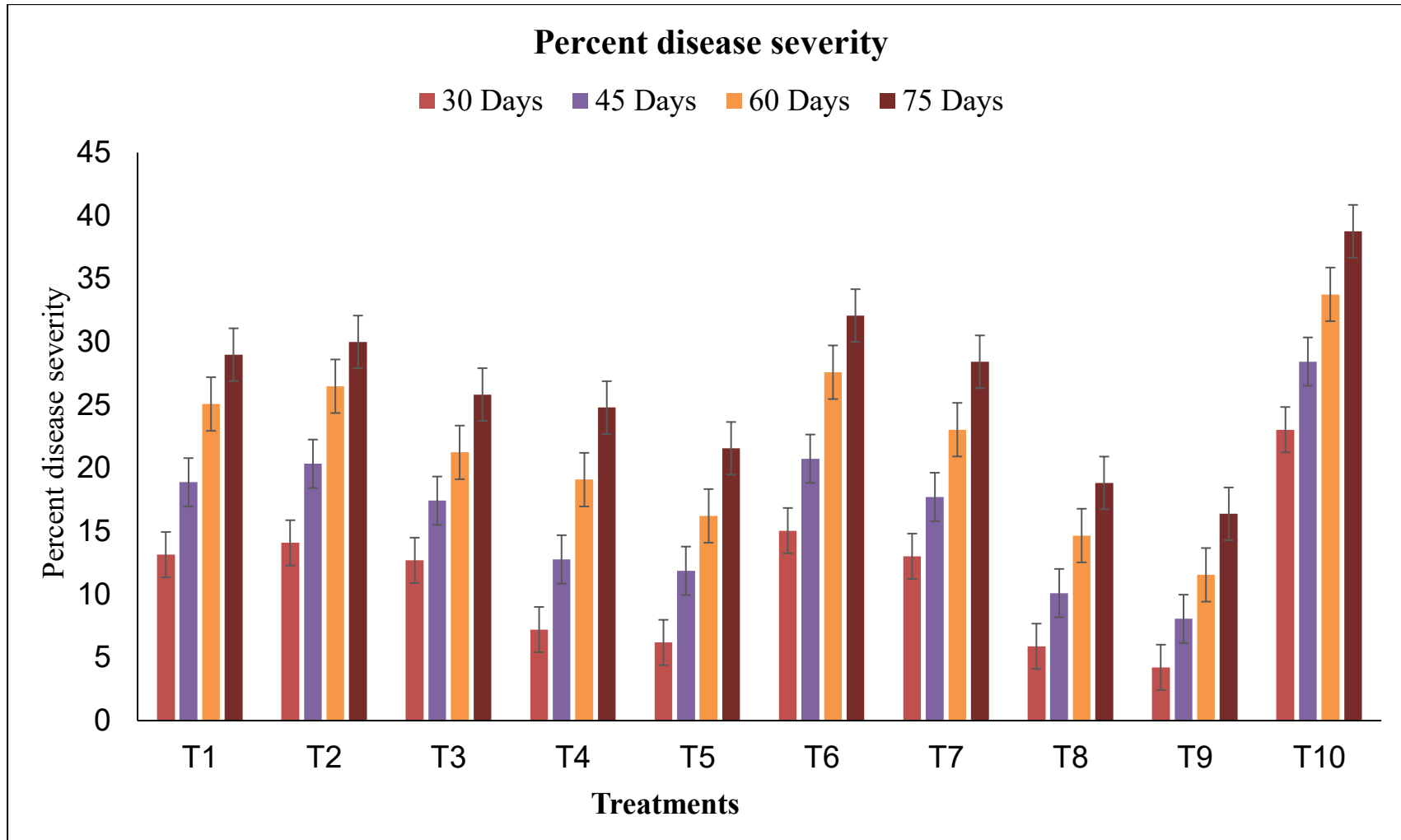


Fig 2: Effect of Integrated disease management on leaf spot of Ashwagandha

Summary and Conclusion

Ashwagandha (*Withania somnifera*) – Alternaria Leaf Spot

1. Leaf spot of Ashwagandha was found to be affected by *Alternaria alternata*, First symptoms recorded at 34 DAS in T₉ as large, zonate, concentric necrotic lesions with brown margins. The lesions appeared mostly on the older leaves but later covered the entire
2. Microscopic observation showed the presence of muriform, beaked conidia with longitudinal and transverse septa, confirming the identity of *A. alternata*.
3. Lowest disease incidence was observed in T₉ 8.73 %, 11.88%, 18.22% and 22.87% followed by T₈ 10.11%, 18.89% , 23.28% and 27.38%, T₅ 10.89%, 20.22%, 27.44% and 31.10% at 30,45,60 and 75 DAS respectively .
4. Minimum disease severity was recorded in T₉ 4.22%, 8.06% ,11.55%, and 16.38% followed by T₈ 5.89%, 10.10 %, 14.66% and 18.83%, T₅ 6.19%, 11.87%, 16.22% and 21.57% at 30,45,60 and 75 DAS respectively .
5. Maximum per cent disease incidence and severity control was observed in T₉ 61.48, 57.78

followed by T₄ 53.91, 51.47 at 75DAT.

6. Weekly increase rate of leaf spot in Ashwagandha (0.46) was recorded at 50th meteorological week when average maximum and minimum temperature was 23.1°C and 4.0°C, with relative humidity 63.4%.
7. The leaf spot disease reached its peak severity 38.77%. in the 13th SMW when maximum and minimum temperatures were 35.5°C and 14.8°C, 63.6 mm of rainfall,
8. Total evaporation exhibited the strongest positive correlation with disease severity ($r = 0.5826$), with an R^2 value of 0.1666 and regression equation $y = 0.4112x + 21.553$.

Reference

- Ojiambo PS, Yuen J, van den Bosch F, Madden LV (2017). Epidemiology: Past, Present, and Future Impacts on Understanding Disease Dynamics and Improving Plant Disease Management- A Summary of Focus Issue Articles. Phytopathology.
- Jayakumar, T., Hsieh, C. Y., Lee, J. J., & Sheu, J. R. (2013). Experimental and clinical pharmacology of andrographis

- paniculata and its major bioactive phytoconstituent andrographolide. *Evidence-Based Complementary and Alternative Medicine*, 2013, 846740.
- Kumar, S., Singh, B., & Bajpai, V. (2021). *Andrographis paniculata* (Burm. f.) Nees: Traditional uses, phytochemistry, pharmacological properties and quality control/quality assurance. *Journal of Ethnopharmacology*, 275, 114-054.
- Mani, M.T. and Hepziba, S.J., 2009. Integrated disease management of Pear millte downy mildew caused by *Scleropora graminicola*. *Archives of Phytopathology and plant protection* 42(2):136-141.
- Meena, R.P. and Roy, S. (2020). Morphological and molecular characterization of *Fusarium* sp. causing wilt disease of Isabgol (*Plantago ovata* Forsk.) and its management strategies, *Journal of Applied Research on Medicinal and Aromatic Plants*. 16: 100-244.
- Martínez-Arias, C., Witzell, J., Solla, A., Martin, J.A., Rodríguez-Calcerrada, J., 2022. Beneficial and pathogenic plant-microbe interactions during flooding stress. *Plant. Cell Environ.* 45, 2875–2897.
- Mishra, R. S., Haridas, R. S., and Agrawal, P. (2021). High entropy alloys—Tunability of deformation mechanisms through integration of compositional and microstructural domains. *Materials Science and Engineering: A*, 812, 141085.
- Natti, J.J., Dickinson, M.H. and Atkin, J.D. (1967). Resistance of *Brassica oleracea* varieties to downy mildew. *Phytopathology*, 57: 144-147.
- Aceto, M.D., Harris, L.S., Abood, M.E. and Rice, K.C. (1999). Stereoselective mu- and delta- opioid receptor-related antinociception and binding with (+) - thebaine. *European Journal of Pharmacology*, 365(2-3): 143-147.
- Meena, R. P. and Kalariya, K. A. and Saran, P. L. and Ponnuchamy Manivel, Ponnuchamy Manivel, (2019). Evaluation of ashwagandha (*Withania somnifera* L.) Dunal accessions and breeding lines against leaf spot disease caused by *Alternaria alternata* under subtropical condition of India., *Journal of Applied Research on Medicinal and Aromatic Plants*. 100 (211): 2214-7861.