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# Molecular Characterization and Fungicidal Efficacy Against *Rhizoctonia* Isolates from Rice and Potato

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#### **KEYWORDS**

Rhizoctonia solani, rice,

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#### **ABSTRACT**

This study aimed to characterize *Rhizoctonia* isolates from rice sheath blight (R-1) and potato (P-1) infections and evaluate the efficacy of diverse fungicides against them. Molecular analysis identified the rice isolate (R-1) as belonging to the AG-1 group, displaying a distinct 265 bp band, while the potato isolate (P-1) exhibited amplification at 500 bp using Rs1F2(F)/Rs2R1(R) markers. Fungicide efficacy testing against R-1 revealed significant inhibition by Bavistin 50 EC, Titan Classic 5 EC, Orius 25 EC, and Emesto Prime 240 ES across various concentrations. Tilt 25 EC and Monceren 250 SC exhibited inhibition at 10 ppm and 100 ppm, while Fujita 40 EC showed the least inhibition (88.4% at 100 ppm). For P-1, Emesto Prime 240 ES and Monceren 250 SC consistently inhibited growth across 5–100 ppm. These findings highlight differential responses of *Rhizoctonia* isolates to fungicides, identifying potential candidates for effective disease management in rice and potato cultivation. Understanding fungal behavior and fungicide interactions is crucial for sustainable disease control. Further research should explore the mechanisms driving these responses to develop targeted management strategies, ultimately improving crop yield and quality.

#### INTRODUCTION

Rice (Oryza sativa L.) is a major cereal crop and a staple food for a large segment of the global population. In India, rice was cultivated over 47.83 million hectares with a record production of 137.82 million metric tonnes during the 2023-24 agricultural year (Anonymous, 2024). To meet the increasing global food demand, enhancing rice productivity remains a priority. However, various biotic and abiotic stresses significantly hinder rice cultivation, leading to substantial yield losses. Among fungal diseases, sheath blight, caused by *Rhizoctonia solani*, is one of the most damaging, with yield losses ranging from 4% to 40%, depending on crop stage, disease severity, and environmental conditions (Zheng et al., 2013; Bhunkal et al., 2015b). The disease severity is exacerbated by higher tiller density, temperatures between 25-30°C, and relative humidity of 80-100% (Bhunkal et al., 2015a).

Similarly, *R. solani* also causes black scurf disease in potato, which can lead to yield losses of up to 30% (Carling et al., 1989). This economically significant disease affects potato quality, yield, and plant productivity (Ahmad et al., 1995; Khan et al., 1995). The pathogen survives as mycelium in decomposing

material or sclerotia on potato tubers, and its infection of stolons results in malformed tubers (Weinhold et al., 1978; Escande & Echandi, 1991; Jeger et al., 1996). Such infections impair nutrient and water translocation, further diminishing productivity.

Due to the absence of resistant cultivars, fungicide application remains the primary disease management strategy. However, concerns regarding maximum residue limits (MRLs), environmental contamination, and food safety have led to strict regulatory scrutiny. India, as a leading exporter of agricultural commodities, faces challenges from the USA and European Union, where excessive fungicide residues have resulted in consignment rejections (Kumar et al., 2018). The indiscriminate use of fungicides contributes to ecological imbalances and potential health risks.

Several studies have evaluated fungicidal efficacy against *R. solani* (Moni et al., 2010; Singh et al., 2016) and assessed fungicide residues in soil, rice grains, potato tubers, and straw (Kundu et al., 2011; Arora et al., 2014; Chen et al., 2015). However, there is limited information on developing new fungicidal formulations, their bio-efficacy, and residue management. Addressing these gaps is essential for sustainable

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disease control, reducing agricultural losses, and ensuring food security.

The present study aims to evaluate the in vitro efficacy of different fungicides against sheath blight and black scurf disease, providing insights for enhanced disease management strategies in rice and potato cultivation.

#### 2. MATERIAL AND METHODS:

#### 2.1 Collection and isolation:

Sheath blight and black scurf disease samples were collected from Punjab region of India during 2019-2020 and were designated as R-1 and P-1, respectively. Sheath blight infected samples showing typical symptoms were collected, were cut into small bits and sterilized in 1% sodium hypochlorite for 1 minute.

Table 1: Primer sequences and detailed information

Sterilized bits were placed on the Petri plates containing potato dextrose agar (PDA) medium. Potato tubers infected with black scurf were cleaned under tap water. Later, sclerotia pieces were scraped off using a sterilized knife. The isolation was made further, as mentioned above. Both the isolates were incubated at  $25\pm2^{\circ}C$  to obtain pure cultures and they were used for the analysis.

#### 2.2 Molecular characterization:

Two *Rhizoctonia* isolates collected from rice and potato infected samples each, were grown on potato dextrose broth (PDB) for genomic DNA isolation. The CTAB technique was used to isolate DNA from fungal mat and the procedure followed as stated by Murray and Thompson (1980).

Primer name		Primer sequence (5'- 3')	Annealing temp. (°C)	Reference
R. solani AG common Primer (Forward)		5'-CTCAAACAGGCATGCTC-3'	50	
	AG1-1A	5'-CAGCAATAGTTGGTGGA-3'	48	
AG-1	AG1-1B	5'-AAGGTCCTTTGGGGTTGGGG-3'	60	Matsumoto
	AG1-1C	5'-CTTTTTTGGGGGGCCTTGC-3'	60	(2002)
AG-2	AG2-1	5'-AGGCAATAGGTTATTGGACC-3'	56	
	AG2-2	5'-CATGGATGGGAGAACTTTTA-3'	54	
AG-3	Rs1F2	5'- TTGGTTGTAGCTGGTCTATTT-3'	51.6	Lees <i>et al</i> (2002)
	Rs2R1	5'- TATCACGCTGAGTGGAACCA-3'	55.9	Lees et al (2002)

PCR profile for AG-1 marker: For AG 1: PCR amplifications were performed at initial denaturation at  $94^{\circ}$ C for 5minute, followed by 30 cycles of Denaturation at  $94^{\circ}$  C for 1 minute, 2-minute primer Annealing at  $54^{\circ}$ C for, 3 minutes primer extension at  $72^{\circ}$ C, followed by final extension at  $72^{\circ}$ C for 10 minutes.

PCR profile for AG-3 PT marker: For AG 3-PT: PCR amplifications were carried at initial denaturation at 95°C for 2 minutes, followed by denaturation at 95°C for 45 seconds for 35 cycles, annealing at 65°C for 60 seconds, Extension at 72°C for 1 minute 30 seconds, followed by final extension at 72°C for 5 minutes.

#### 2.3 Poison food technique:

Serial dilutions of various concentrations of each fungicide were prepared from a stock solution of 100 millilitres. Three

replications were made for each concentration for testing. Along-side, suitable control was maintained without fungicide of both the isolate. The inoculated and control Petri plates were incubated at 25±2°C and the data was recorded after the growth of control plates were covered completely. Per cent inhibition in mycelial growth was calculated based on the formula given by Bliss *et al* (1934).

$$P.I = \frac{C - T}{C}$$

Where, P.I =Percent inhibition; C= Growth in control plate; T= Growth in treated plate

Table 2: Description of chemical fungicides along with mode of action

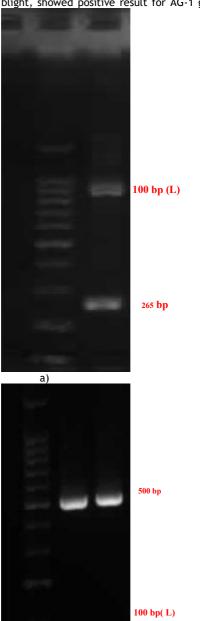
S.no.	FUNGICIDE	CONCENTRATIONS (ppm)	GROUP	TARGET SITE AND CODE
1	Bavistin 50 EC (Carbendazim)	1,5,10,50,100	B: Cytoskeleton and motor protein	ß-tubulin assembly in mitosis
2	Titan classic 5 EC (Hexaconazole)	5,25,50,100	G: sterol biosynthesis in membranes	G1 C14- demethylase in sterol biosynthesis (erg11/cyp51)
3	Tilt 25 EC (Propiconazole)	1,5,10,50	G: sterol biosynthesis in membranes	G1 C14- demethylase in sterol biosynthesis (erg11/cyp51)
4	Fujita 40EC(Isoprothiolane)	5,25,50,100	F: lipid synthesis or transport / membrane integrity or function	F2 phospholipid biosynthesis, methyltransferase
5	Pulsor 24 SC (Thifluzamide)	5,25,50,100	C. respiration	C2 complex II: succinate-dehydrogenase
6	Galileo Way 18.76 SC (Picoxystrobin + Propiconazole)	5,25,50,100	G: sterol biosynthesis in membranes	G1 C14- demethylase in sterol biosynthesis (erg11/cyp51)
7	Amistar 250EC (Azoxystrobin)	1,5,10,50,100	C. respiration	C3 complex III: cytochrome bc1 (ubiquinol oxidase) at Qo site (cyt b gene)
8	Orius 25 EC(Tebuconazole)	5,25,50,100	G: sterol biosynthesis in membranes	G1 C14- demethylase in sterol biosynthesis (erg11/cyp51)
9	Emesto prime 240 ES (Penflufen )	5,25,50,100	C. respiration	C2 complex II:succinate-dehydrogenase
10	Monceren 250 SC(Pencycuron)	5,25,50,100	B: Cytoskeleton and motor protein	B4 cell division (unknown site)

#### FRAC CODE LIST (2025)

#### 3. RESULTS AND DISCUSSION:

**3.1 Molecular characterization of** *Rhizoctonia* **isolates** *Rhizoctonia* **isolate** (R-1) collected from infected rice sheath blight, showed positive result for AG-1 group with band size of

265bp and other markers belonging to different AGs did not show any amplification. This result confirmed that rice isolate belonged to AG-1 group. Whereas, potato isolate (P-1) amplified at a band size of 500 bp with markers Rs1F2(F)/ Rs2R1(R).



b)
Fig.1: Rhizoctonia isolate (R-1) collected from infected rice sheath blight, showed positive result for AG-1 group with band size of 265bp and other markers belonging to different AGs did

not show any amplification. This result confirmed that rice isolate belonged to AG1 group. Fig.2: Potato isolate was characterized to AG3- PT with an amplified product of 500bp.

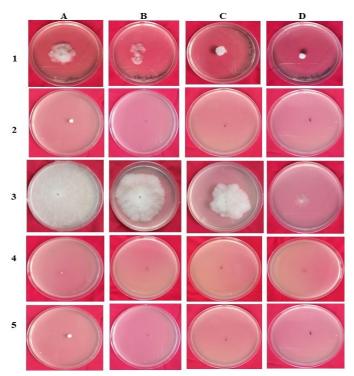
## Fig 1: PCR profile of *Rhizoctonia* solanigenerated by AG subgroups specific marker AG1-1A (265bp) and AG3-PT (500 bp) a) rice isolate b) potato isolate; Lane 1: 100 bp

**3.2** Screening of fungicides against *Rhizoctonia* isolates: The efficacy of ten fungicides tested against *Rhizoctonia* isolate (R-1) were tested as represented in table 3. The percent inhibition of fungus was calculated at different concentration. Out of all the fungicides tested, Bavistin 50 EC, Titan classic 5 EC, Orius 25 EC, Emesto prime 240 ES showed maximum inhibition

at all the concentrations used in this experiment. Followed by,

ppm and 100 ppm. The least inhibition of fungus showed by Fujita 40EC even at 100 ppm (88.4 %). Similarly, two fungicides were tested against *Rhizoctonia* isolate (P-1) as represented in table 2. Both Emesto prime 240 ES and Monceren 250 SC inhibited fungus at maximum at all the concentration ranging from 5 to 100 ppm.

Tilt 25 E C and Monceren 250 S C started inhibiting fungus at 10



#### Fungicides:

- 1. Propiconazole(Ppm =1,5,10,50)
- 2. Hexaconazole (Ppm=5,25,50,100)
- 3. Isoprothiolane=(Ppm=5,25,50,100)
- 4. Tebuconazole (Ppm=5,25,50,100)
- 5. Thifluzamide(Ppm=5,25,50,100)

Isolate: Rice (R-1)



Control

Plate 1: Effect of different fungicides against rice isolate (R-1) under in vitro condition

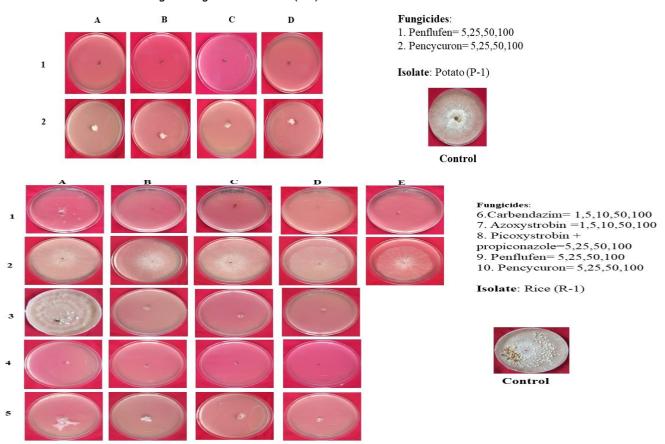


Plate 2: Effect of different fungicides against potato isolate (P-1) under in vitro condition

Table 3: Effect of different fungicides against rice isolate (R-1) under in vitro condition

Table 3. El	rect of different fungicides against	rice isolate (K	(- i) under in vic	ro condition				
C no	Fungicide	Percent Inhib	ition					
S.no.	rungicide	1 ppm	5 ppm	10 ppm	25 ppm	50 ppm	100 ppm	

1	Bavistin 50 EC	100.0	100.0	100.0	-	100.0	100.0
2	Titan classic 5 E C	-	100.0	-	100.0	100.0	100.0
3	Tilt 25 E C	77.3	85.6	100.0	-	100.0	-
4	Fujita 40EC	-	27.1	-	45.2	69.1	88.4
5	Pulsor 24 SC	-	85.5	-	89.8	94.9	100.0
6	Galileo Way 18.76 SC	-	0.0	-	87.4	93.1	97.0
7	Amistar 250EC	0.0	0.0	0.0	-	0.0	0.7
8	Orius 25 EC	-	100.0	-	100.0	100.0	100.0
9	Emesto prime 240 ES	-	100.0	-	100.0	100.0	100.0
10	Monceren 250 SC	-	83.3	-	86.9	94.2	100.0
Control		0.0	0.0	0.0	0.0	0.0	0.0

Table 4: Effect of different fungicides against potato isolate (P-1) under in vitro condition

S. no.	Fungicide	Percent inhibition					
	rungicide	1 ppm	5 ppm	10 ppm	25 ppm	50 ppm	100 ppm
1	Emesto prime 240 ES	-	100.0	-	100.0	100.0	100.0
2	Monceren 250 S C	=	100.0	=	100.0	100.0	100.0
Control		0.0	0.0	0.0	0.0	0.0	0.0

Matsumoto (2002) standardized PCR profile for direct identification of *Rhizoctonia* isolates using AG-specific markers belonging to AG-1 and AG-2 subgroups. Lees *et al* (2002) standardized PCR profile for AG3-PT subgroup identification with markers Rs1F2(F)/ Rs2R1(R). Goswami *et al* (2017) identified one hundred and twelve isolates causing sheath blight in rice as AG-1-1A subgroup, indicating its dominancy over other subgroups. El-Zaidy *et al* (2018) found twenty-six *Rhizoctonia* isolates infecting potato when amplified with AG-3 PT specific primers produced a band size of 500bp.

Sheath blight of rice is not controlled by resistance breeding till date. So, the better way to manage of sheath blight is by chemical control. Hunjan et al (2010) reported that Trifloxystrobin+ tebuconazole inhibited the sheath blight disease in both field and lab conditions. They confirmed that newly available fungicides like tebuconazole, pencycuron and thifluzamide were found effective in managing the disease. Similarly, Lore et al (2012) reported that under both lab and field conditions, propiconazole, hexaconazole and carbendazim showed maximum inhibition of fungus. Prakash et al (2013) found that carbendazim and hexaconazole showed maximum growth inhibition of 86.62 and 71.22 per cent, but propiconazole and tebuconazole found to have moderate inhibition. Datta et al (2017) studied the comparative effectiveness of the triazoles group on Rhizoctonia solani and reported that among four fungicides (hexaconazole, propiconazole, difenoconazole, tebuconazole), hexaconazole found to have maximum inhibitory effect with lowest EC50 value.

Treatment of Moncern fungicide for tubers affected with black scurf greatly reduced the scleotial viability (Campion *et al* 2003). Different anastomosis groups (AG 3, AG 2-1 and AG 5) showed difference in percent inhibition against Monceren fungicide with AG 5 isolates showing moderate resistant with highest EC50 value. But with respect to AG 3, Monceren showed inhibition of 84.75 per cent (Malik *et al* 2014). Agarwal *et al* (2008) reported that different potato isolates showed variation in inhibition of mycelial growth with Monceren fungicide, the maximum inhibition found was 77.1 per cent and minimum was 70.4 per cent.

#### CONCLUSION

The study confirms the rice isolate (R-1) as part of the AG-1 group based on positive amplification at 265bp. The potato isolate (P-1) differs, displaying a distinct genetic profile with a 500bp band using specific markers Rs1F2(F)/Rs2R1(R). These

distinct genetic signatures highlight unique characteristics of each isolate, crucial for tailored disease management in rice and potato crops. The study highlighted varied responses of fungicides against *Rhizoctonia* isolates R-1 and P-1. Bavistin 50 EC, Titan classic 5 EC, Orius 25 EC, Emesto prime 240 ES, and Monceren 250 SC exhibited high inhibition for both isolates across different concentrations. Fujita 40EC showed the least efficacy against R-1. This emphasizes the need for tailored fungicide selection based on isolate-specific responses for effective disease control.

#### 5. ACKNOWLEDGEMENT

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#### 6. CONFLICT OF INTERESTS

The authors declare no conflict of interest in relation to this publication. All forms of help and financial assistance have been duly acknowledged.

#### REFERENCE

- Ahmad, I., Khan, M. A. & Javed, N. 1995.
   Pathogenicity and host range of Rhizoctonia solani causing black scurf of potato. Pakistan Journal of Phytopathology, 7(1): 40-45.
- Anonymous. 2024. Final estimates of foodgrain production for 2023-24. Press Information Bureau, Government of India.
- Arora, S., Mukherjee, I. & Kumar, A. 2014. Dissipation of tricyclazole and validamycin in rice ecosystem. Environmental Monitoring and Assessment, 186(2): 835-841.
- Bhunkal, N., Singh, R. & Mehta, N. 2015a. Progression and development of sheath blight of rice in relation to weather variables. Journal of Mycology and Plant Pathology, 45: 166-172.
- Bhunkal, N., Singh, R. & Mehta, N. 2015b. Assessment of losses and identification of slow blighting genotypes against sheath blight of rice. Journal of Mycology and Plant Pathology, 45: 285-292.
- Bliss, C. 1934. The method of probits. Science, 79: 38.

- Campion, C., Chatot, C., Perraton, B. & Andrivon, D. 2003. Anastomosis groups, pathogenicity and sensitivity to fungicides of Rhizoctonia solani isolates collected on potato crops in France. European Journal of Plant Pathology, 109(9): 983-992.
- Carling, D. E., Leiner, R. H. & Westphale, P. C. 1989.
   Symptoms, significance, and biology of Rhizoctonia disease in potato. American Potato Journal, 66(9): 693-701.
- Chen, W., Li, M., Li, W., Wu, X. & Han, L. 2015.
   Dissipation and residue level of thifluzamide in rice field ecosystem. Journal of Chemistry, 2015: 1-6.
- Datta, S., Dey, P., Sarkar, A., Tarafdar, J. & Chowdhury, A. 2016. Comparison among four triazole fungicides on growth and development of sheath blight of rice pathogen Rhizoctonia solani Kühn AG1-1A. Archives of Phytopathology and Plant Protection, 49(9-10): 239-251.
- El-Zaidy, M., Mahmoud, M. A., Al-Othman, M. R. & Abdelazim, N. S. 2018. Molecular identification and characterization of Rhizoctonia solani AG-3 isolates causing black scurf of potato. Pakistan Journal of Botany, 50(1): 415-420.
- Escande, A. & Echandi, E. 1991. Protection of potato from Rhizoctonia canker with binucleate Rhizoctonia fungi. Plant Pathology, 40: 197-202.
- FRAC. 2025. FRAC (Fungicide Resistance Action Committee): Code list Fungicides. Available at: https://www.frac.info/
- Goswami, S. K., Singh, V. & Kashyap, P. L. 2017. Population genetic structure of Rhizoctonia solani AG1-IA from rice field in North India. Phytoparasitica, 45: 299-316.
- Hunjan, M. S., Lore, J. S., Pannu, P. P. S. & Thind, T. S. 2011. Performance of some new fungicides against sheath blight and brown spot of rice. Plant Disease Research, 26: 61-67.
- Jeger, M. J., Hide, G. A., van den Boogert, P. H. J. F., Termorshuizen, A. J. & van Baarlen, P. 1996. Pathology and control of soil-borne fungal pathogens of potato. Potato Research, 39: 437-469.
- Khan, R. A., Iftikhar, S., Rafi, A., Riaz, S. & Ahmad, I. 1995. Distribution and incidence of tuber diseases of potato in Swat Valley. NARC, PSPDP, PARC, Islamabad.
- Kumar, P., Ahlawat, S., Chauhan, R., Kumar, A., Singh, R. & Kumar, A. 2018. In vitro and field efficacy of fungicides against sheath blight of rice and postharvest fungicide residue in soil, husk, and brown rice using gas chromatography-tandem mass spectrometry. Environmental Monitoring and Assessment, 90(9): 1-9.
- Kundu, C., Goon, A. & Bhattacharyya, A. 2011. Harvest residue study of fungicide tebuconazole EC formulation in groundnut and paddy. Journal of Environmental Protection Science, 2: 424-428.
- Moni, Z. R., Ali, M. A. & Meah, M. B. 2010. Efficacy of fungicides and plant extracts against sheath blight (Rhizoctonia solani) of rice. Bangladesh Journal of Agricultural Research, 35(3): 367-374.
- Murray, M. G. & Thompson, W. F. 1980. Rapid isolation of high molecular weight plant DNA. Nucleic Acids Research, 8(19): 4321-4326.
- Prakash, G., Singh, U. D., Sharma, P. & Pandian, R. T.
   P. 2013. Evaluation of pesticides against rice sheath blight caused by Rhizoctonia solani. Indian Phytopathology, 66(4): 351-355.
- Singh, R., Singh, A. & Singh, U. S. 2016. Management of sheath blight of rice with combination of fungicides and biocontrol agents. Indian Phytopathology, 69(1): 56-60.
- Weinhold, A. R., Bowman, T. & Hall, D. H. 1978. Rhizoctonia disease of potato in California. American Potato Journal, 55: 56-57.
- Zheng, A., Lin, R., Zhang, D., Qin, P., Xu, L., Ai, P., Ding, L., Wang, Y., Chen, Y. & Liu, Y. 2013. The

evolution and pathogenic mechanisms of the rice sheath blight pathogen. Nature Communications, 4: 1424.