

BIOCHEMICAL ESTIMATION OF PRIMARY METABOLITES IN OCIMUM TENUIFLORUM

ARCHANA SHARMA*, ANJU MEENA AND RISHI KESH MEENA¹

Department of Botany, Vedic P. G. Girls College, Raja Park, Jaipur - 302 004

¹Department of Botany, Govt. Birla College, Bhawani Mandi - 326 502

E-mail: drarchanasharma11@gmail.com

KEY WORDS

Ocimum tenuiflorum
Primary metabolites
Total soluble sugar
Lipid, Starch, Phenols

Received on :

16.05.2011

Accepted on :

07.08.2011

*Corresponding author

ABSTRACT

Ocimum tenuiflorum is an important medicinal herb belonging to family Lamiaceae. It is commonly known as 'Shyama tulsi'. In the present investigation, various *in vivo* plant parts (leaves, stem and root) and *in vitro* (callus) of *ocimum tenuiflorum* was carried out for biochemical estimation of primary metabolites viz. total soluble sugar, starch, lipid, protein and phenol. The plant parts varied in composition of their primary metabolites. Results showed that the maximum content of total soluble sugar (3.5 ± 0.08 mg/gfw), lipid (2.6 ± 0.11 mg/gfw), protein (3.6 ± 0.65 mg/gfw) and phenol (1.8 ± 0.456 mg/gfw) and maximum starch found in root (2.1 ± 0.014 mg/gfw).

INTRODUCTION

Ocimum tenuiflorum commonly known as holy basil (English), or Shyama Tulsi (local language), is a herbaceous sacred plant found throughout India. Indian material medica describes the use the plant in a verity of ailments. In Indian mythology the plant is considered to extirpate all sins and purify the body when touched. It is often grown outside dwellings and worshipped daily. It is said to daunt Yama, the god of death, but has a close affinity with Lord Krishna who is reputed to have grown the herb. If offered to Lord Krishna it is said to have mystical powers of protection from death, disease and misfortune. (Williamson, 2002) Different parts of plant like stem, flower, seed, leaves, root etc are known to possess therapeutic potential and have been used, by traditional medicinal practitioners, as analgesic, antibacterial (Khanna and Bhatia, 2003), anticancer (Karthikeyan et al., 1999) and hypoglycemic agent (Rai et al., 1997) etc. early studies have been reported its wound-healing activity (Shetty et al., 2006), Anti oxidant activity (Samjon et al., 2007).

Primary metabolites directly involved in growth and development of plants. Primary metabolites viz, chlorophyll, amino acid, nucleotides and carbohydrates have a key role in metabolic process such as photosynthesis, respiration and nutrient assimilation. They are used as raw material and food additives. Many plant such as *Gloriosa superba*, *Ricinus communis* *Euphorbia hirta* and *Moringa oleifera* Lam have been evaluated for their composition of primary metabolites (Rishi and Sarin, 2009; Vijayvergia et al., 2009 and Sharma et al., 2010). The present study was conducted to investigate

biochemical estimation of primary metabolites viz, total soluble sugar, starch, lipid, protein and phenol in *ocimum tenuiflorum*.

MATERIALS AND METHODS

Plant parts (stem, leaves and root) of *Ocimum tenuiflorum* were collected from the campus of university of Rajasthan, Jaipur and callus developed by tissue culture technique. The *in vivo* plant material were separately washed with running water to remove dust and powdered with motor and pestle.

The quantitative estimation of primary metabolites was carried out using different protocols. The powdered *in vivo* plant parts (leaves, stem and root) and *in vitro* callus of *ocimum tenuiflorum* was used for estimation of carbohydrate (Dubois et al., 1956), protein (Lowry et al., 1951), lipid (Jayaraman, 1981), starch, (Dubois et al., 1956) and Phenol (Bray and Thorpe, 1954) respectively. All experiment repeated in triplicate and means (\pm SD) were calculated.

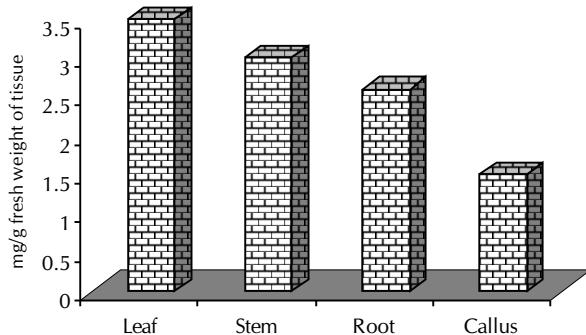
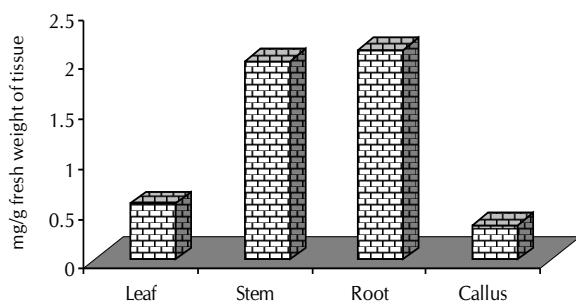
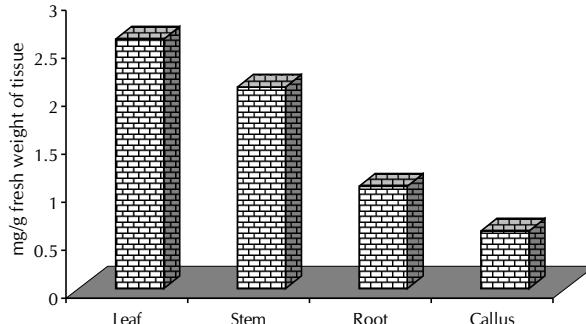
RESULTS AND DISCUSSION

In the present investigation, various *in vivo* plant parts (leaves, stem and root) and *in vitro* (callus) of *Ocimum tenuiflorum* was carried out for biochemical estimation of primary metabolites viz, total soluble sugar, starch, lipid, protein and phenol (Table 1). The various plant parts (stem, leaves and root) and *in vitro* callus of *Ocimum tenuiflorum* varied in composition of primary metabolites studied. Maximum content of total soluble sugar was observed in leaf of *Ocimum tenuiflorum* (3.5 ± 0.08 mg/gfw) and minimum in callus (Fig.

Table 1: Estimation of primary metabolites (mg/gfw) in *Ocimum tenuiflorum*

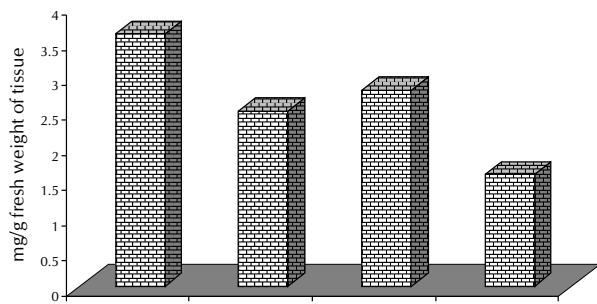
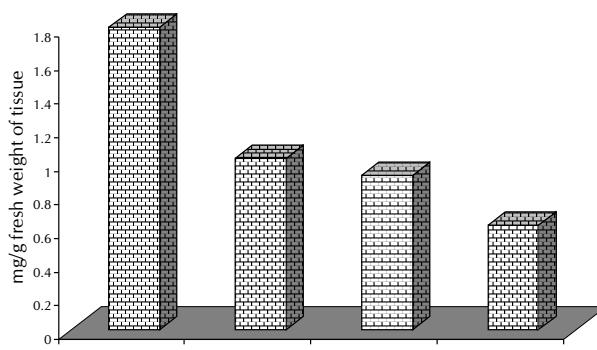
Plant parts	Total soluble sugar	Starch	Lipid	Protein	Phenol
Leaf	3.5 ± 0.08	0.56 ± 0.33	2.6 ± 0.11	3.6 ± 0.65	1.8 ± 0.46
Stem	3.0 ± 0.01	1.98 ± 0.26	2.1 ± 0.45	2.5 ± 0.33	1.02 ± 0.03
Root	2.6 ± 0.40	2.1 ± 0.14	1.07 ± 0.21	2.8 ± 0.34	0.92 ± 0.11
Callus	1.5 ± 0.22	0.34 ± 0.05	0.6 ± 0.02	1.6 ± 0.22	0.62 ± 0.02

Abbreviation: mg/gfw = milligrams per g fresh weight of tissues

**Figure 1: Estimation of total soluble sugar in *Ocimum tenuiflorum*****Figure 2: Estimation of starch in *Ocimum tenuiflorum*****Figure 3: Estimation of lipid in *Ocimum tenuiflorum***

1). Higher content of total soluble sugar was reported in leaves of *M. indica* (Vijayvergia and Shekhawat, 2009). Plant sugars can be used as artificial sweetener and they can even help in diabetes by supporting the body in its rebuilding (Freeze, 1998).

Among all *in vivo* plant parts root had higher starch level (2.1 ± 0.14 mg/gfw) as compared to stem and leaf. *In vitro* callus sowed minimum starch (0.34 ± 0.05) (Fig. 2). According to Sharma *et al.* (2006), the presence of higher levels of starch in intact plant part might be due to more storing capacity to

**Figure 4: Estimation of protein in *Ocimum tenuiflorum*****Figure 5: Estimation of total phenol in *Ocimum tenuiflorum***

escape the drought conditions.

The highest amount of lipid (2.6 ± 0.11 mg/gfw) protein (3.6 ± 0.65 mg/gfw) and phenol (1.8 ± 0.456 mg/gfw) was also recorded in the leaf of *Ocimum tenuiflorum*, and minimum in callus (Figs. 3, 4 and 5).

Lipid a diverse group of primary metabolites, include reserve plant material such as fats, essential oils, waxes terpenoids and oleoresin. Plant lipid have developed products that work with diverse requirements, be it culinary, medicinal or cosmetic (Yadav and Tyagi, 2006).

In the present studies, among all the samples (*in vivo* and *in vitro*) tested, leaf showed higher protein level protein (3.6 ± 0.65 mg/gfw) followed by, root, stem, and callus (Fig. 4). In similar studies carried out protein content was in leaves of *M. indica* (Vijayvergia and Shekhawat, 2009). The presence of higher protein level in the plant parts towards their possible increase in food value or that a protein based bioactive compounds could also be isolated in future (Thomsen *et al.*, 1991).

The total phenolic contents was also recorded maximum (1.8 ± 0.456 mg/gfw) in the leaf of *Ocimum tenuiflorum*) and minimum in callus (Fig. 5). The higher amount of phenols is

important in regulation of plant growth, development and disease resistance. Plant phenol may interfere with all stages of cancer process, potentially resulting in a reduction of cancer risk (Hollman, 2001) Phenolic compounds have been studied in different plant species like *Arachis hypogaea* and *Simmondsia chinensis* (Ali, 2000); *Chlorophytum borivilianum* (Singh, 2005), *Balanites aegyptica* L. (Vijayvergiya and Vijay, 2006); *Terminalia catappa* (Nagesh et al., 2007).

REFERENCES

- Ali, D.J. 2000.** Biotechnological techniques of morphogenetic regulation of some economically important plants. Ph. D. Thesis, University of Rajasthan, Jaipur.
- Bray, H. G. and Thorpe, W. V. 1954.** Analysis of phenolic compounds of interest in metabolism. In: Method of Biochemical Analysis (Ed.) Glick, D., Interscience Publishers, Inc. 1-27.
- Dubois, M. K., Gilles, J. K., Robers, P. A. and Smith, F. 1956.** Calorimetric determination of sugar and related substance. *Analyt. Chem.* **26:** 351-356.
- Freeze, H. 1998.** Disorder in protein glycosylation and protein therapy. *The J. Pediatrics.* **133(5):** 553-600.
- Hollman, P. C. 2001.** Evidence for health benefits of plant phenol. *J. Sci. of food and Agr.* **89:** 842-852.
- Jayaraman, J. 1981.** Laboratory manual in biochemistry. Wilsey eastern limited, New Delhi. pp. 96-97.
- Karthikeyan, K., Ravichandran, S. and Govindasamy, S. 1999.** Chemopreventive effect of *Ocimum sanctum* on DMBA-induced hamster buccal pouch carcinogenesis. *Oral. Oncol.* **35:** 112-9.
- Khanna, N. and Bhatia, J. 2003.** Antibacterial activity of *Ocimum sanctum* L.fixed oil. *Indian J. Exp. Biol.* **43(9):** 835-7
- Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. 1951.** Protein measurement with the folin phenol reagent. *J. Biol. Chem.* **193:** 265.
- Nagesh, D., Swarna, R., Srinivas, P., Sriram, S., Sundara Rajan, S. and Nagamani, T. S. 2007.** Biochemical studies in leaves of *Terminalia catappa*. In: Proc. 94th Indian Science Congress Part II (Abstracts). Annamalainagar. Jan 3-7. pp. 129.
- Rai, V., Iyer, U. and Mani, U. V. 1997.** Effect of Tulasi (*Ocimum sanctum*) leaf powder supplementation on blood sugar levels, serum lipids and tissue lipids in diabetic rats. *Plant Foods Hum. Nutr.* **50:** 9-16.
- Rishi, A. and Sarin, R. 2009.** Estimation of primary metabolites From Gloriosa superbaL. *in vivo* and *in vitro*. *Int. J. Mendel.* **26(1-4):** 87.
- Samjon, J., Sheeladevi, R. and Ravindran, R. 2007.** Oxidative stress in brain and antioxidant activity of *Ocimum sanctum* in noise exposure. *Neurotoxicology*, Article in press.
- Sharma, R. A., Meena, R. C., Jain, S. C. and Saxena, M. K. 2006.** Physio-chemical analysis of two species of *Cassia* : *C. obtusifolia* Linn. and *C. siamea* Lam. In: Proc. of International Conference, Bot. Expo. 25-27 March, Jaipur. pp. 274-276.
- Sharma, A., Yadav, A., Barman, N. and Malwal, M. 2010.** Quatification of primary metabolites of *Moringa oleifera* Lam. *The Bioscan.* **5(3):** 403-405.
- Shetty, S., Udupa, S., Udupa, L. and Somayaji, N. 2006.** Wound healing activity of *Ocimum sanctum* Linn with supportive role of antioxidant enzymes. *Indian J. Physiol. Pharmacol.* **50(2):** 163-8.
- Singh, R. 2005.** Biochemical changes in embryogenic callus cultures of *Chlorophytum borivilianum* Sant. Et Fernand. In :Abst Vol. of XXVIII conference of Indian Botanical Society, Oct. 24-26, B.S.I., Dehradun. pp. 56.
- Thomsen, S., Handen, H. S. and Nyman, V. 1991.** Ribosome inhibiting proteins from in vitro culture of phytolacea decandra. *Planta. Medica.* **57:** 232-236.
- Vijayvergiya, R. and Vijay, P. 2006.** Quantitative estimation of primary metabolites of *Balanites aegyptiaca* L. In: Abstract Vol. of XXIX. All India Botanical Conference, Oct. 9-11, Udaipur. pp. 16.
- Vijayvergiya, R. and Shekhawat, N. 2009.** Biochemical estimation of primary metabolites of *Madhuca indica* GMEL. *The Bioscan.* **2(3):** 203-206.
- Vijayvergiya, R., Sharma, S. and Singh, T. 2009.** Biochemical estimation of primary metabolites of some medicinal plants of Eupobiaceae family. *J. Indian Bot. Soc.* **88(1-2):** 116-119.
- Williamson, E. M. 2002.** Major herbs of Ayurveda. Londen, Churchill Livingstone.
- Yadav, P. R. and Tyagi, R. 2006.** Lipid Biotechnology. Discovery publishing house-New Delhi.

