

CHANGES IN CARBOHYDRATE CONTENT IN THE SEEDS OF SCHLEICHERA OLEOSA (KUSUM) DUE TO BIODETERIORATION BY PATHOGENIC FUNGI DURING STORAGE

A. K. SRIVASTAVA*, S. KUMAR¹ AND G. K. PANDEY²

Department of Botany, St. Xavier's College, Ranchi - 834 001

¹Department of Biotechnology, St. Xavier's College, Ranchi - 834 001

²Department of Botany, St. Columba's PG College, Hazaribag - 825 301

E-mail: ajaysrivastava11@gmail.com

KEY WORDS

Schleicheria oleosa
Colonising fungi
Carbon sources
Mycodwellers

Received on :

24.03.2011

Accepted on :

19.07.2011

*Corresponding author

ABSTRACT

The unsanitary and humid condition during storage of seeds of *Schleicheria oleosa* f. *Sapindaceae* (Kusum) makes it prone to a faster biodeterioration due to colonization of mycoflora namely *Aspergillus fumigatus*, *Aspergillus flavus*, *Aspergillus niger*, *Fusarium solani*, *paccilomyces variotii* and *mucar* spp. These internal mycodwellers significantly decreased the carbohydrate content in comparison with the control group not infested with the fungi. In their prolonged association with the seeds for one year the glucose level dropped by as much as 50.15% of its initial value while control group by just 2.15%. Similarly, total sugar also registered a loss of 40.25 per cent with control group at 1.48 per cent. The starch in the stored seeds reduced steadily by 48.88% but the control group lost only 2.04%.

INTRODUCTION

Seeds of *Schleicheria oleosa* f. *Sapindaceae* (Kusum) are of economic importance in Jharkhand. Its oil is used in burning lamps, varnishing, massage and medicine while oil-cake is good manure. Kusum seeds are collected just after onset of rainy season i.e. by the end of month of May. Conventionally seeds are stored in gunny bags in the village houses and godowns. In either case the condition is unsanitary and humid and is conducive for fungal growth leading to good scale destruction.

There are reports indicating that fungi are associated with seed surface as well as internal mycoflora (Dutta and Roy, 1987). Changes in glucose content with pathogenic association have been reported. Chaudhary and Prasad (1974) found a depletion of glucose among other nutrients in the shoot tissues of *Cajanus cajan* infected with *Fusarium oxysporum* f. sp. *udum*. It was worthwhile to seek the extent of biodeterioration occurring in the seeds in terms of carbohydrates in different forms viz. glucose, total sugars and starches.

MATERIALS AND METHODS

Kusum seeds were obtained from the godown of Ranchi forest department for the preparation of oil and oilcake on every month from June 2001 to May 2002. The experimental group seeds were kept in fifty gunny bags in godown while control group seeds were kept in three air tight container under same

environmental conditions. The variation of temperature, rain fall and humidity were monitored on daily basis. Every month few seeds from all the three airtight containers and three randomly picked gunny bags were sampled for glucose, total sugar and starch estimation. Glucose content was estimated by titration method using Fehling solutions (Cole's method, 1914). Total sugar was estimated by colorimetric method using anthrone reagent (Dubois *et al.*, 1951). Estimations were done in triplicate and standard deviation was calculated.

RESULTS AND DISCUSSION

Kusum seeds were collected just after onset of rainy season. The high humidity and moderate temperature (Table 1) in unsanitary condition favour colonization of fungi namely *Aspergillus fumigatus*, *Aspergillus flavus*, *Aspergillus niger*, *Fusarium solani*, *paccilomyces variotii* and *mucar* spp. The effect of continued fungal activity on the seeds manifested the lowered carbohydrate content. The glucose level dropped by as much as 50.15 percent in the span of one year of its initial value but the control group lost only 2.15 percent, shown in Table 2. Similarly, total sugar also registered a loss of 40.25 per cent with control group at 1.48 per cent (Fig. 1). The starch in the stored seeds reduced steadily by 48.88% but the control group lost only 2.04% (Fig. 2).

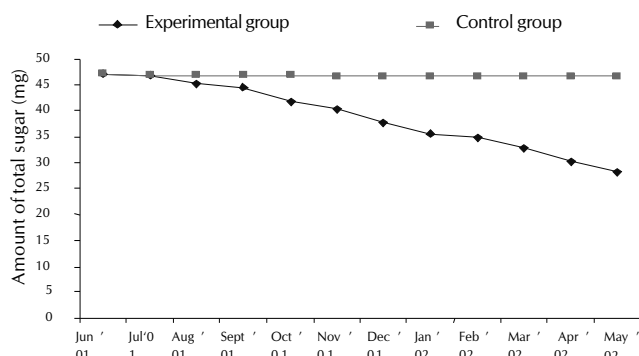
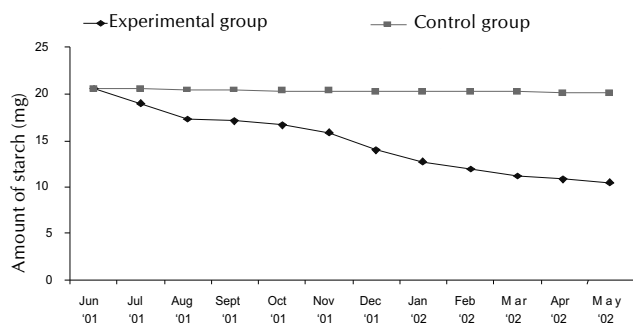
The effect of continued fungal activity on the seeds manifested in the lowered glucose, total soluble sugars and starch contents as seen in oil content (Srivastava and Pandey, 2000). Changes in glucose content with pathogenic association have been

Table 1: Environmental factors prevailing through June'01 to May'02

Month	Rain fall mm/month	Temperature°C		Humidity %
		min. ± S.D.	max. ± S.D.	
Jun-01	133.2	22.1 ± 0.23	34.6 ± 0.11	66.3
Jul-01	254.1	20.1 ± 0.14	30.1 ± 0.21	84.6
Aug-01	152.4	21.9 ± 0.10	31.3 ± 0.22	86.5
Sep-01	43.9	23.1 ± 0.31	34.8 ± 0.24	82.5
Oct-01	20.7	21.6 ± 0.24	33.2 ± 0.19	73.4
Nov-01	2.0	18.7 ± 0.19	28.9 ± 0.20	61.2
Dec-01	0.0	13.5 ± 0.16	22.1 ± 0.21	54.8
Jan-02	1.1	11.4 ± 0.26	24.3 ± 0.09	46.4
Feb-02	0.0	14.3 ± 0.09	28.2 ± 0.11	39.6
Mar-02	3.0	21.8 ± 0.22	32.1 ± 0.14	46.9
Apr-02	0.0	29.2 ± 0.25	35.6 ± 0.11	30.8
May-02	6.0	27.4 ± 0.17	38.7 ± 0.18	23.4

reported. Chaudhary and Prasad (1974) found a depletion of glucose among other nutrients in the shoot tissues of *Cajanus cajan* infected with *Fusarium oxysporum* f. sp. *udum*. Chattopadhyaya and Nandy (1978) found carbohydrate loss due to *F. moliniforme* var *subglutinum* infection. Loss in amount of glucose in fruits have been reported for tomato-*Drechslera australiense* (Kapoor and Tandon, 1970); tomato-*Alternaria solani* (Mehta *et al.*, 1975); banana- *Gloeosporium musarum* (Wang, 1960) and citrus- *Xanthomonas campestris pvcitri* (Vidhyasekaran and Durairaj, 1971) host pathogen systems.

Changes in total sugar content in the host tissues due to pathogenic actions have been reported by Craig and Hooker

**Figure 1: Amount of total carbohydrate present in 1 g of Kusum seed powder samples through one year of biodeterioration due to fungal attack, June '01 - May '02.****Figure 2: Amount of starch present in 1 g of Kusum seed powder samples through one year of biodeterioration due to fungal attack, June '2001 - May '02.****Table 2: Amount of glucose present in 1g of Kusum seed powder samples through one year of biodeterioration due to fungal attack, June'01 to May'02 (n = 3)**

Month	Amount of glucose in mg per g of seed control group M ± S.D.	Amount of glucose in mg per g of seed experimental group M ± S.D.
Jun '01	16.75 ± 0.008	16.75 ± 0.011
Jul '01	16.69 ± 0.010	16.25 ± 0.009
Aug '01	16.64 ± 0.009	16.25 ± 0.013
Sept '01	16.61 ± 0.009	15.75 ± 0.008
Oct '01	16.60 ± 0.011	14.90 ± 0.014
Nov '01	16.56 ± 0.012	14.31 ± 0.012
Dec '01	16.53 ± 0.009	13.13 ± 0.009
Jan '02	16.51 ± 0.013	12.47 ± 0.008
Feb '02	16.49 ± 0.017	11.90 ± 0.013
Mar '02	16.45 ± 0.015	10.31 ± 0.010
Apr '02	16.41 ± 0.009	09.52 ± 0.008
May '02	16.39 ± 0.011	08.35 ± 0.010

(1961); Dhanvantari (1967); Dayal and Joshi (1968) and Padmanabhan *et al.* (1988) on different host pathogen systems.

Similarly the depletion in the starch content is commensurate with the activation of starch degrading enzymes especially beta – *amylase* (Schipper and Mirocha, 1977). The drop is due to the sugar being used by the fungal pathogens as respiratory substrates (Baker, 1965; Wu, 1973). In pigeon pea seeds infested with *Aspergillus flavus*, Sinha and Prasad (1977) found a depletion of starches. Likewise Bilgrami *et al.*, (1979) recorded a considerable decrease in the amount of starch in paddy seeds during 60 and 90 days of fungal infestation of an aflatoxin producing strain of *Aspergillus parasiticus*. Sinha *et al.*, (1981) found a considerable reduction in the starch content of *Cajanus cajan* seeds infested with *Aspergillus flavus* and *A. Niger*. However, these seeds, when infested with *Alternaria alternata* and *Curvularia lunata*, showed a moderate reduction in the starch contents. On the contrary, *Cajanus* seeds infested with *Fusarium moniliforme* and *Drechslera hawaiiensis* exhibited a minimum level reduction in starch contents. Later on, Singh and Sinha (1985) confirmed that infestation of *Cajanus* seeds by *Aspergillus parasiticus* caused a considerable decline in their starch contents. Prasad (1989) reported a loss of starch in fungi infested seeds of *Coriandrum indicum* and the maximum loss in starch was due to *Aspergillus flavus* followed by *Curvularia lunata*.

All these reports are concurrent to our findings. It is thus apparent that one year of seed infestation of *Schleichera oleosa* predominantly and jointly by *Aspergillus fumigatus*, *A. flavus*, *A. niger*, *Fusarium solanii*, *Paecilomyces variotii* and *Mucor* sp. causes biodeterioration of its seeds manifested in the diminishing of carbohydrate contents.

REFERENCES

- Baker, J. 1965. Study in the respiratory and carbohydrate metabolism of plant tissues. XVIII. The effect of oxygen on starch formation and dissolution in potatoes. *New Phytol.* **64**: 201-209.
- Bilgrami, K., Jamaluddin, S., Sinha, R. K. and Prasad T. 1979. Changes in seed contents of paddy (*Oryza sativa* L.) due to fungal flora. *Phytopath. J.* **96**: 9- 14.
- Chattopadhyaya, N. C. and Nandy, B. 1978. Changes in total contents

of saccharides proteins and chlorophyll in malformed inflorescence induced by *Fusarium moniliforme* var *subglutinans*. *Biol. Plantarum*. **20**: 468-471.

Chaudhary, S. K. and Prasad, M. 1974. Variation in sugar contents of healthy and *Fusarium oxysporum*, *Fusarium udum* infected plants of *Cajanus cajan*. *Phytopath.* **80**: 303-305.

Cole, S. W. 1914. The estimation of lactose and glucose by the copper-iodide method. *Biochem. J. Apr.* **8(2)**: 134-142.

Craig, J. and Hooker, A. L. 1961. Relation of sugar trends and pith density to Diplodia stalk rot in dent corn. *Phytopathology*. **51**: 376-382

Dayal, R. and Joshi, M. M. 1968. Post infection changes in the sugar content of leaf spot infected barley. *Indian Phytopath.* **21**: 221-222.

Dhanvantari, B. N. 1967. The leaf scorch disease of Strawberry, *Diplocarpon earlianum* and the nature of resistance to it. *Can. J. Bot.* **45**: 1525-1543.

Dubois, M., Gilles, K., Hamilton, J. K., Rebers, P. A. and Smith, F. 1951. A colorimetric method for the estimation of sugar. *Nature*. **168**: 167.

Dutta, G. R. and Roy, A. K. 1987. Mycoflora associated with *Strychnos* seeds and deterioration of their active principles under storage. *Indian Pathology*. **40**: 520-524.

Kapoor, I. J. and Tandon, R. N. 1970. Post infection changes in sugar content of tomato fruits caused by *Dreschlera australiense*. *Indian Phytopath.* **23**: 133-135.

Mehta, P., Vyas, K. M. and Saksena, S. P. 1975. Metabolic changes during pathogenesis of fruit rot disease of tomato. *Indian Phytopath.* **28**: 253-255.

Padmanabhan, P., Alexander, K. C. and Shanmugam, N. 1988. Some metabolic changes induced in sugarcane by *Ustilago scitaminea*. *Indian Phytopath.* **41**: 229-232.

Prasad, M. 1989. Plant - bacteria interaction - Its dynamics and dimensions. In *Plant Microbe Interactions*. Ed: KS Bilgrami. Narendra Publishing House, Delhi. pp. 143-161.

Schipper, A. L. and Mirocha, C. J. 1977. Mechanism of starch hydrolysis in bean leaves during infection by bean rust fungus. *Phytopathology*. **58**: 1066.

Singh, P. and Sinha, K. K. 1985. Changes in the seed content of arhar infected with *Aspergillus parasiticus*. *Indian Phytopathol.* **38**: 560.

Sinha, M. K. and Prasad, T. 1977. Deterioration of arhar seeds by *Aspergillus flavus*. *Indian Phytopathol.* **30**: 70-72.

Sinha, M. K., Singh, K. K. and Prasad, T. 1981. Changes in starch contents of Arhar seeds due to fungi. *Indian Phytopathol.* **34**: 269-271.

Srivastava, A. K. and Pandey, G. K. 2000. Chemical changes in properties of Kusum (*Schleicheria oleosa*) oil during its seed infestation by fungi. *J. Mycopathol. Res.* **38(1)**: 29-32.

Vidhyasekaran, P. and Durairaj, P. 1971. Quality of the citrus fruits infected by *Xanthomonas citri*. *Indian Phytopath.* **24**: 781-782.

Wang, M. C. 1960. Physiological studies on *Gloeosporium musarum* Cook. At Mass, the causal organism of banana anthracnose. Changes in the carbohydrate composition of banana pulp with reference to the adaptive secretion of amylase. *Bot. Bull. Acad. Sin. N.S.* **1**: 59-75.

Wu, L. C. 1973. Changes in some enzymes of moong bean seeds germinated on mycelia macerate of *Rhizoctonia solani*. *Physiol. Plant Pathol.* **3**: 19-27.

