

EFFECT OF DIFFERENT pH LEVELS ON THE GROWTH AND SPORULATION OF *FUSARIUM OXYSPORUM* SCHLECHT. F. SP. *LENTIS* (VASUDEVA AND SRINIVASAN) THE CAUSAL ORGANISM OF WILT DISEASE OF LENTIL

HIMANSHU BHUSHAN JARUHAR* AND AJAY PRASAD¹

Department of Botany, S. P. D. College, Garhwa - 822 114, Jharkhand

¹Department of Botany, S. S. J. S. N. College, Garhwa - 822 114, Jharkhand

E-mail: hbjaruhar@gmail.com

KEY WORDS

Fusarium oxysporum f. sp. *lentis*
Macro conidia
Micro conidia
Chlamydospores

Received on :

16.02.2011

Accepted on :

19.04.2011

*Corresponding author

ABSTRACT

Effect of pH on the growth and sporulation of *Fusarium oxysporum* Schlecht. f. sp. *lentis* (Vasudeva and Srinivansan) after incubation of two weeks *in vitro* culture in sucrose nitrate medium was studied. pH level 6.0 was found optimum for the growth as well as sporulation of the fungus. Sporulation of chlamydospore was however found best in the pH level 4.0. Further increases in the pH level show retarding effect on growth and sporulation. Size of the spores increases with increase in the pH range.

INTRODUCTION

Fusarium oxysporum Schlecht f. sp. *lentis* (Vasudeva and Srinivasan) causes a serious wilt disease in lentil (*Lens esculenta* Moench.) which is an important pulse crop of Palamu commissioner (23°52'North latitude and 84°17'East longitude). Due to this disease there is huge loss of the yield in lentil. Vasudeva and Srinivasan (1952) have reported that Wilt disease of lentil caused by *Fusarium* spp. is one of the serious disease and it causes huge loss of the standing crop throughout the world. The extent of the damage to the crop due to the disease ranges from 20-24% annually, (Saxena and Johansen, 1990 and Ali, 2007). There were various reports that indicate *Fusarium* spp grow at different pH levels for growth and sporulation (Wilson, 1946; Srobar, 1978; Prasad et al., 1992; Souramma and Singh, 2004; Groenewald, 2005). The pathogen is soil born and infects the host during seedling stage through root and blocks the vascular system (Vasudeva and Srinivasan, 1952). For the successful cultivation of the lentil it is necessary to investigate the physiology of the fungus. Therefore, effect of different pH level on the growth and sporulation of *Fusarium oxysporum* Schlecht. f. sp. *lentis* was undertaken.

MATERIALS AND METHODS

The fungus was isolated from the infected plant of lentil (*Lens*

esculenta Moench). Monospore culture as described by Prasad and Chaudhary (1966, 1967) was employed in the present study. From the culture of the isolates, pure culture was obtained using 10⁻⁵ decimal level dilution plate technique. Sucrose nitrate medium consisting of sucrose (50.0g), KNO₃ (10.0g), KH₂PO₄ (5.0g), MgSO₄·7H₂O (2.5g) and 1000 mL double distilled water used as basal medium. There were 10 different pH level ranging from 2.0 to 6.5 with a difference of 0.5 were prepared by using pocket size pH meter HANNA Instrument Co. Mauritius by using either N/10 HCl or NaOH before autoclaving. 100 mL of the medium was taken in 250 mL conical flask and five replicate sets were used in each case. The solution was autoclaved at 15 psi for 15 minutes. The inoculation was done with 3 mm discs of the fungus culture cut with a sterilized cork borer from a margin of 10 days old colony growing in Potato Dextrose Agar (PDA) medium. Flasks were then inoculated at 26°C ± 2°C for two weeks. The mean dry weight of the mycelium was determined as described by Prasad and Chaudhary (1966). The number of spores was counted using known depth of Haemocytometer slide (0.01 cm) using the formula:
Where,

$$\text{Number of spores / 100 mL} = V/N \times 100$$

N = Average number of spores per square of the four corner square of haemocytometer counted.

Table 1: Effect of different pH levels of mycelia growth, spore population and size of spore change of *Fusarium oxysporum* Schlecht. *F. sp. Lentis* (Vasudeva and Srinivasan). Incubation period: 2 Weeks

S. No.	pH range	Dry wt. in mg.	Spores in millions /100mL medium*			Size of spores in μ^*		
			Macro conidia	Micro conidia	Chlamydo spores	Macro conidia	Micro conidia	Chlamydo spores
1	2.0	37.00	0.67	2.62	0.39	26.20	7.20	4.90
2	2.5	53.00	0.78	3.63	3.51	27.70	7.30	4.90
3	3.0	73.00	1.17	4.06	3.64	28.20	8.10	6.10
4	3.5	93.00	1.55	7.42	5.20	28.90	8.90	6.30
5	4.0	124.00	1.69	9.37	4.03	29.20	9.60	7.30
6	4.5	169.00	1.95	13.68	3.71	30.70	9.80	7.50
7	5.0	198.00	2.19	19.68	3.71	31.20	10.70	7.50
8	5.5	276.00	2.62	69.14	2.73	33.30	12.30	8.20
9	6.0	476.00	5.86	154.03	2.19	34.20	12.90	8.30
10	6.5	210.00	3.69	132.42	3.51	31.10	9.20	8.50
SE \pm		15.24	1.50	5.78	1.74	0.05	0.18	0.24
C.D. at p= (5%)		30.97	3.14	11.75	3.54	0.11	0.36	0.48

*mean of five replicates

V = Volume of haemocytometer (0.256×10^{-5}) cc
 Length of the spores was measured by calibrated ocular micrometer under compound microscope ($10 \times 45 \times$ of magnification).

RESULTS AND DISCUSSION

The mean dry weight of mycelium and sporulation of three spore forms of the fungus on different pH levels is recorded in Table 1. The perusal of the results showed that *Fusarium oxysporum* Schlecht. f. sp. *lentis* grew maximum in pH 6.0. Very high acidic range of pH showed very poor growth of mycelium. Mycelial mat accumulation increased with increase in pH but declined after pH 6.0. Sporulation of the macroconidia and microconidia was observed to be maximum at pH 6.0. Least sporulation occurred in pH 2.0. Chlamydo spores produced were noted to be the maximum at pH 4.0. Length of macroconidia and microconidia were the maximum at pH 6.0 and thereafter it started to decline. Diameter of chlamydo spores increased with increase in pH level.

The growth and sporulation of many fungi have been studied under the influence of various pH levels. Mix (1933) found that pH range from 4 to 8 showed good growth for *Phyllosticta solitaria*. Wilson (1946) observed acid soil (pH 4.2) support growth of *Fusarium* spp. where as a pH near neutrality prevents growth. Chaudhary (1971) and Prasad et al. (1992) reported 6.0 pH level as the best for the growth and sporulation of *Fusarium moniliforme* v *subglutinans* Wr. and Rg. Munjal and Gautam (1977) observed that maximum fungal growth and sporulation occurred at pH 6.0 and 5.5 respectively for *Septoria humuli*. Srobar (1978) found pH 6 to be the most suitable for the growth of all species while a highly acidic medium was unsuitable for sporulation of all species causing fusarioses disease in wheat. Jat and Goyal (1978) found that growth and sporulation of *Claviceps microcephala* to be optimum at pH 7.5 and 6.0 respectively. Nair(1957); Souramma and Singh (2004); Groenewald (2005); Kishore et al. (2009) also found pH 5.5 to 7.0 to be the best for growth and sporulation of *Fusarium oxysporum* f. sp. *lini* (Belley). Such reports are in agreement with present findings for this pathogen. This indicates that unusual acidity badly hampered the growth

and sporulation of the fungus. On the other hand sporulation of chlamydo spores at more acidic range indicates that this spore is characteristically different from other two spores and more tolerable to stressed condition than the macro and micro conidia.

ACKNOWLEDGEMENT

The authors are happy to record the grateful cooperation of the persons concerned of the G.L. A. College, Medininagar, in the completion of this work.

REFERENCES

- Ali, M. 2007. Augmentation of Pulses Production. Mission2007: IIPR Kanpur-24: pp: 16-21
- Chaudhary, S. K. 1971. Studies of the Physiology of the *Fusarium oxysporum* f. *udum* (Butler) Snyder and Hansen (Causal Organism of Wilt of *Cajanus cajan* (Linn.) Millsp) Ph.D. Thesis, Deptt. of Bot. Ranchi University Ranchi, p. 297.
- Groenewald, S. 2005. Biology, Pathogenicity and Diversity of *Fusarium oxysporum* f. sp. *cubense*. M.Sc. (Agri.) Thesis. Faculty of Natural and Agricultural Science, University of Pretoria etd, Pretoria. p. 176.
- Jat, R. G. and Goyal, J. P. 1978. Physiological Studies of *Claviceps microcephala* Causing Ergot Diseases of Bajra. *Ind. J. Mycol. and Pl. Pathol.* **8**: 21.
- Kishore, R., Pandey, M., Dubey, K. and Kumar, Y. 2009. Effect of Temperature and pH on Growth and Sporulation of *Fusarium oxysporum* f. sp. *lini* (Bolley) Snyder and Hensan Causing Linseed Wilting. *Prog. Agric.* **9(1)**: 147-149.
- Mix, A. J. 1933. Factors Affecting the Sporulation of *Phyllosticta solitaria* in Culture. *Phytopath.* **23**: 503.
- Munjal, R. L. and Gautam, S. R. 1977. Effect of vitamins on growth and sporulation of *Septoria humuli*. *Indian Phytopath.* **30**: 566-567.
- Nair, P. N. 1957. Factors Affecting Resistance of Flax to *Fusarium oxysporum lini* (Belley). *Dis. Abstr.* **17(5)**: 942.
- Prasad, A., Chaudhary, R. S. and Chaudhary, S. K. 1992. Effect of Different pH levels on Growth and Sporulation of *Fusarium moniliforme* V. *subglutinans* Wr. and Rg., The Causal Organism of Wilt of Maize. *Bio. J.* **4(1 and 2)**: 75-78.

- Prasad, M. and Chaudhary, S. K. 1966.** Studies on The Effect of Different Phosphorus Concentrations on the Production of Chlamydo spores, Microconidia and Macroconidia in the Culture of *Fusarium udum* Butler. *Proc. Nat. acad. Sci. India.* **36B:** 43 – 48.
- Prasad, M. and Chaudhary, S. K. 1967.** Effect of Sulphur on Sporulation of *Fusarium udum* Butler. *J. Ind. Bot. Sci.* **46:** 45 – 51.
- Saxena, N. P. and Johansen, C. 1990.** Advances in Biotic and Abiotic Stress in Pulse Crops. I.I.P.R. Kanpur - **24:** 211-219.
- Souramma, V. and Singh, J. 2004.** Effect of Temperature and pH on Growth and Sporulation of Wilt Causing Pathogen *Fusarium oxysporum* f sp *lini* (Bolley) Snyder and Hensan in linseed (*Linum usitatissimum* L.). *J. Oilseed Research.* **21(1):** 206-207.
- Srobar, S. 1978.** The influence of temp. and pH on the growth of mycelium of the causative agents of Fusarioses in wheat in Slovakia ,Czechoslovakia.UVTI (Ustav Vedecko Technickych informaci) Ochrana Rostlin. **14:** 269-274.
- Vasudeva, R. S. and Srinivasan, K. V. 1952.** Studies on the Wilt Disease of Lentil (*Lens esculenta* Moench). *India Phytopath.* **5:** 23-32.
- Wilson, I. M. 1946.** Observations on wilt disease in flax. *Transactions of the British Mycological Society.* **29:** 221-231.

