

PLANT GROWTH PROMOTING RHIZOBACTERIA- IMPACTS ON CAULIFLOWER YIELD AND SOIL HEALTH

MANOJ KAUSHAL* AND RAJESH KAUSHAL

Department of Basic Science, Dr Y. S. Parmar University of Horticulture and Forestry, Nauni - 173 230, Solan (Himachal Pradesh) INDIA
e-mail:kaushal.mbg@gmail.com

KEYWORDS

Cauliflower
Disease suppression
Integrated
PGPR
P- solubilization
Soil health

Received on :

09.11.2012

Accepted on :

08.04.2013

*Corresponding author

ABSTRACT

The present investigation were conducted for two summer seasons during 2009 and 2010 in the field of Department of Soil science and Water Management, Dr. Y. S. Parmar University of Horticulture and Forestry, Nauni, Solan, Himachal Pradesh to study the effects of integrated nutrient management systems on growth, yield of cauliflower and soil health. The conjoint effect of plant growth promoting rhizobacteria (PGPR) at varying (50%, 75% and 100%) doses N and P fertilizers registered a significant increase in number of non-wrapper leaves, curd diameter, curd depth and curd weight of cauliflower, total microbial counts and available N and P contents of soil. The application of MK₂ isolate at 75% recommended dose of NP fertilizers not only increased the yields of cauliflower by 24% but also saved 31kg N/ha and 8 kg P/ha fertilizers over control (recommended doses of NPK). A significant increase in available N and P contents were also noted by the conjoint application of PGPR and chemical fertilizers. Hence the developed integrated nutrient module can be used for enhanced yields without destructing soil health in mid hills of North Western Himalayas.

INTRODUCTION

Plant growth promoting rhizobacteria (PGPR), are considered as efficient microbial competitors in the root zone to enhance plant growth directly and/or indirectly by reducing soil borne pathogens, enhancing efficiency of applied inputs and also helps in degrading xenobiotic compounds. In the context of increasing international concern for food and environmental quality, the use of plant growth-promoting rhizobacteria (PGPR) for reducing chemical inputs in agriculture is a potentially important issue. PGPR have been applied to various crops to enhance seed emergence, growth and crop yield, and only a few isolate have been commercialized. PGPR are also known to produce antibacterial compounds that are effective against certain plant pathogens and pests. PGPR are directly involved in increased uptake of nitrogen, synthesis of phytohormones, and solubilization of minerals such as phosphorus and production of siderophores that chelate iron and make it available to the plant root (Bowen and Rovira, 1999).

Cauliflower is grown throughout the year in different agro-climatic zones occupying an area of 2800 ha with annual production of 54,500 million tonnes (National Horticulture Board, nhb, 2010). The high-yielding cauliflower variety has resulted in an increase in cauliflower production but requires large amounts of chemical fertilizers which lead to health hazards and environmental pollution. In order to make cauliflower cultivation sustainable and less dependent on chemical fertilizers, it is important to have effective PGPR

isolate that can biologically fix nitrogen, solubilize phosphorus and induce some growth promoting substances like indole acetic acid (IAA) besides acting as biocontrol agents that can contribute to the production of cauliflower.

Therefore, the present investigations were undertaken to screen the PGPR isolates from its natural growing zones with multifarious activities, at various levels of N and P with other recommended package of practices for commercial cultivation of cauliflower.

MATERIALS AND METHODS

Soil and root samples were collected from the rhizosphere of cauliflower plants from three (Hamirpur, Bilaspur and Kangra districts) different naturally growing agro-climatic zones of Himachal Pradesh. The samples were stored at 4°C in the Soil Microbiology Laboratory, Dr. Y.S. Parmar University of Horticulture and Forestry, Nauni, Solan. For isolation of PGPR, one gram of the rhizosphere soil was placed in 9mL of sterilized distilled water under aseptic conditions. The soil suspension was diluted in 10 fold series and the microbial count was determined by the standard pour plate technique (Subba Rao, 1999). After incubation of 24 - 48h, Modified Replica plate technique was used for isolation.

Thereafter, screening of the bacterial isolates for the various plant growth promoting activities like P-solubilization, siderophore formation, HCN concentration, growth on N-free medium, auxin production and antagonism against *Fusarium* spp., *Rhizoctonia solani* and *Pythium* spp. were performed

by adopting the standard methods as given in Table 1. The optimization of growth conditions (physical, chemical and nutritional) of selected bacterial isolates were standardized by conducting separate experiments.

Out of 30 isolates, 5 efficient isolates designated as MK₂, MK₄, MK₅, MK₇ and MK₉ were selected and characterized after successful experiments under *in vitro* and net house conditions. On the basis their multifarious plant growth promoting activities, growth and yield attributes under controlled conditions (growth chamber and net house), three isolates (MK₅, MK₇ and MK₉) were selected for field experiments along with varying (50%, 75% and 100%) doses of N and P fertilizers. The experiment was conducted during summers of 2009 and 2010 at Nauni, Solan of Himachal Pradesh. Seeds of recommended variety were treated with bacterial inoculum for 8h and untreated seeds were treated with sterilized water for same time and designated as control. Bacterial cell suspension (O.D. 1.00 at 540nm) of 72h old culture grown in nutrient broth at the rate of 10 per cent was used as inoculum for field experimentation. The seeds were sown in nursery and one month old seedlings were transplanted in the field at the spacing 60 × 45cm. The treatments combinations viz.: T1 (Control), T2 (MK₅ + 50% NP), T3 (MK₅ + 75% NP), T4 (MK₅ + 100% NP), T5 (MK₇ + 50% NP), T6 (MK₇ + 75% NP), T7 (MK₇ + 100% NP), T8 (MK₉ + 50% NP), T9 (MK₉ + 75% NP) and T10 (MK₉ + 100% NP) were arranged in RBD design and replicated thrice. The sources of nitrogen and phosphorus were CAN (25% N) and SSP (16% P₂O₅) respectively. All phosphorus fertilizers were applied at the time of transplanting of seedlings and nitrogen fertilizer was applied in three split applications up to the curd formation stages. A booster dose of bacterial culture was added at every one month interval till harvesting. Weeding was done manually and the crop was irrigated when required.

Physico – chemical properties viz. pH, EC, organic matter (OM) and bulk density of soil sample was determined before start of the experiment and after the termination of the experiment and was determined as per the standard method adopted by Jackson (1973) while, available N, available P, available K was determined by the methods adopted by Subbiah and Asija (1956), Olsen et al., 1954 and Merwin and Peech (1951) respectively. The observations were recorded on different quantitative characters of cauliflower viz. (number of non-wrapper leaves, curd diameter, curd depth and curd weight and curd yield). Five plant samples at the time of harvest were also randomly collected from each plot and mixed separately to determine concentrations of N, P and K at harvest using procedure described by Jackson (1973). Statistical analysis was performed as per the design suggested by Gomez and Gomez (1984).

RESULTS AND DISCUSSION

On the basis of morphological, physiological and biochemical characteristics isolates were presumptively identified as *Bacillus* spp. All the five isolates were P-solubilizers, nitrogen fixers, auxin and siderophore producers. And only three (MK₅, MK₇ and MK₉) isolates were HCN producer and were also able to show antagonism towards *Rhizoctonia solani*, *Pythium*

spp. and *Fusarium* spp. A large group of researchers gave evidences to suggest that PGPR enhance the growth and crop yield, and known to produce antibacterial compounds that are effective against certain plant pathogens and pests (Kloepper et al., 2004). In our studies, bacterial isolate (MK₅) showed maximum antagonistic activity against the three tested fungal pathogens and all five isolates were able to solubilize phosphate in the rhizosphere soil.

Then these isolates (MK₂, MK₄, MK₅, MK₇ and MK₉) were further characterized and the activities were also quantified. The results represented in Table 2 reveal that MK₅ had highest phosphate solubilizing efficiency (PSE i.e. 172.21%) which was statistically at par with rest of the four isolates. In liquid medium maximum phosphate was solubilize by MK₅ (664.30 µg/ml), which was statistically at par with MK₄, MK₇ and MK₉. Growth inhibition shown by various bacterial isolates against *Fusarium* spp., *Rhizoctonia solani* and *Pythium* spp. were also summarized in Table 2. Also the isolate MK₅ produced a significantly higher concentration of IAA (29.67 µg/mL) after 72h of incubation as compared to other isolates. All the five bacterial isolates produced a bright zone with yellowish colour around the bacterial colony on Chrome-azurol-S medium. Quantitative estimation of siderophore using Chrome-azurol-S (CAS) liquid assay revealed that bacterial isolate MK₅ produced maximum (33.02 % siderophore unit) at 72h of incubation.

Physico-chemical properties and nutrient status of soil

The physico-chemical properties of soil were recorded at the start and termination of the experiment. The physico-chemical properties of soil were recorded at the start and termination of the experiment. The data on initial soil parameters are pH (6.76), EC (0.47 dSm⁻¹), bulk density (1.04mgm⁻³) and organic matter (1.16%). The initial available N (301.7 Kg/ha) and available K (195.3 Kg/ha) was medium, however available P (41.00 Kg/ha) was in high range. There was no significant change in basic physico-chemical properties of soil i.e. pH, EC, organic matter and bulk density after the termination of experiment.

However, the available nutrient contents N and P were increased by 2.56-32.00% and 6.66-40.72%, respectively over control as given in Table 3. Treatments also did not significantly influence the available K content of soil. The seed inoculation with different bacterial isolates not only improved the nutritional content particularly NPK of plants but also increased their uptake significantly over uninoculated control. This increase may be attributed to atmospheric nitrogen fixation, phosphate solubilization in the rhizosphere. Further due to enhanced uptake by increase in specific ion fluxes at the root surface in the presence of plant growth promoting rhizobacteria has also been reported by Bertrand et al. (2000). Apparently, synergistic effect of chemical fertilizers and PGPR could have brought significant improvement in soil available nutrients. The higher nutrient concentration with conjoint use of PGPR and chemical fertilizers may be attributed to well develop root system, significant improvement in soil physical properties, microbial and metabolic activity and higher photosynthesis rate, which might have improved absorption of nutrients by plants (Hazara et al., 1987).

Plant Parameters

The application PGPR isolates at different levels of N and P significantly increased number of non wrapper leaves, curd diameter, curd weight, curd depth and curd yield over uninoculated control as given in Table 4. The improved growth and yield of cauliflower as a result of integrated use of PGPR and chemical fertilizers might be due to improved photosynthetic and metabolic activity, which led to increase in various plant metabolites responsible for cell elongation

Table 1: Methods adopted to check multifarious plant growth promoting activities of the bacterial isolates.

| Activity | Methods employed |
|-------------------------|-----------------------------|
| Phosphate solubilizing | Pikovskaya (1948) |
| Nitrogen fixing ability | Jensen (1987) |
| Siderophore production | Schwyn and Neilands (1987) |
| HCN production | Bakker and Schippers (1987) |
| Auxin production | Gorden and Paleg (1957) |
| Antagonistic activity | Vincent (1947) |

Table 2: Plant growth promoting activities of selected bacterial isolates

| Isolates | P-solubilization %P-solubilization efficiency | Antifungal activity (% growth inhibition)* | | | Indole-3-acetic acid ($\mu\text{g}/\text{mL}$) | Siderophore activity | | |
|--------------------|---|---|--------------------------------|------------------|---|-------------------------------|-------------------|-----------------------|
| | | P-solubilization in liquid medium ($\mu\text{g}/\text{mL}$) | <i>Fusarium</i> <i>spp.</i> | <i>R. solani</i> | | <i>Pythium</i> <i>spp.</i> | Zone size (mm) | % siderophore unit |
| MK ₂ | 166.67 | 444.3 | 83.33 | 81.81 | 83.72 | 24.83 | 8.67 | 51.36 |
| MK ₄ | 147.22 | 567.67 | 90.38 | 78.40 | 81.39 | 24.67 | 11.33 | 19.40 |
| MK ₅ | 172.21 | 664.30 | 89.28 | 84.04 | 86.04 | 29.67 | 12.67 | 33.02 |
| MK ₇ | 151.36 | 640.33 | 85.71 | 77.27 | 81.39 | 25.50 | 13.33 | 19.86 |
| MK ₉ | 158.33 | 604.00 | 88.09 | 81.81 | 77.90 | 28.33 | 14.67 | 31.14 |
| CD _{0.05} | 76.03 | 131.98 | 5.43 | 7.05 | 10.07 | 3.26 | 2.77 | 15.94 |

Table 3: Effect of different treatments on physico-chemical properties and available nutrient contents of soil

| Treatments | Soil physico-chemical characteristics | | | | | Available N (kg/ha) | Available P (kg/ha) | Available K (kg/ha) |
|--------------------|---------------------------------------|--------------------------|--------------------------|--------|-------|---------------------|---------------------|---------------------|
| | pH | EC (dS m^{-1}) | BD (g cm^{-3}) | OM (%) | | | | |
| T1 | 6.78 | 0.43 | 1.57 | 1.09 | 304.6 | 43.39 | 212.2 | |
| T2 | 6.79 | 0.48 | 1.56 | 1.22 | 312.4 | 46.28 | 203.8 | |
| T3 | 6.81 | 0.49 | 1.54 | 1.28 | 345.6 | 52.88 | 213.3 | |
| T4 | 6.80 | 0.48 | 1.56 | 1.21 | 402.1 | 61.06 | 215.8 | |
| T5 | 6.75 | 0.39 | 1.55 | 1.12 | 313.7 | 46.39 | 213.1 | |
| T6 | 6.81 | 0.48 | 1.55 | 1.29 | 343.9 | 53.27 | 214.1 | |
| T7 | 6.81 | 0.45 | 1.49 | 1.25 | 395.3 | 59.56 | 205.7 | |
| T8 | 6.79 | 0.46 | 1.56 | 1.18 | 317.8 | 47.89 | 206.8 | |
| T9 | 6.80 | 0.43 | 1.50 | 1.22 | 355.0 | 53.05 | 206.8 | |
| T10 | 6.81 | 0.45 | 1.57 | 1.21 | 399.6 | 59.17 | 211.8 | |
| CD _{0.05} | NS | NS | NS | NS | 16.85 | 2.42 | NS | |

Table 4: Effect of different treatments on plant (growth and yield) parameters

| Treatments | Plant Parameters | | | | | | | |
|--------------------|---------------------------------|---------------------------|----------------------|-----------------------|---------------------|---------------------------------|---------------------------------|---------------------------------|
| | No. of non wrapper leaves | Curd diameter (cms) | Curd weight (gms) | Curd yield /ha (q) | Curd depth (cms) | Total N content in plant (%) | Total P content in plant (%) | Total K content in plant (%) |
| T1 | 9.46 | 10.84 | 750.8 | 278.0 | 7.33 | 3.15 | 0.52 | 2.52 |
| T2 | 9.83 | 13.01 | 820.0 | 303.7 | 7.73 | 3.21 | 0.56 | 2.76 |
| T3 | 10.37 | 14.27 | 930.3 | 344.6 | 8.55 | 3.25 | 0.57 | 2.78 |
| T4 | 10.97 | 15.35 | 965.0 | 357.4 | 9.85 | 3.25 | 0.56 | 2.58 |
| T5 | 10.17 | 12.95 | 808.0 | 299.3 | 7.37 | 3.23 | 0.53 | 2.60 |
| T6 | 10.17 | 13.73 | 881.7 | 326.5 | 8.65 | 3.29 | 0.54 | 3.02 |
| T7 | 10.67 | 14.10 | 964.2 | 357.0 | 8.68 | 3.20 | 0.53 | 2.54 |
| T8 | 10.03 | 12.50 | 786.7 | 291.3 | 7.63 | 3.20 | 0.54 | 2.65 |
| T9 | 10.30 | 14.23 | 870.8 | 322.5 | 8.45 | 3.29 | 0.55 | 2.77 |
| T10 | 10.67 | 14.10 | 944.2 | 349.6 | 8.72 | 3.23 | 0.55 | 2.67 |
| CD _{0.05} | 0.72 | 1.57 | 40.32 | 14.93 | 0.81 | 0.07 | 0.03 | 0.50 |

curd depth and curd weight of cauliflower, total microbial counts in soil and available N and P contents of soil. Taken together, the results suggested that PGPR are able to induce the production of IAA, solubilization of phosphorus and resistance to pathogens and pests, thereby improving growth of plants. The use of PGPR as inoculants biofertilizer is an efficient approach to replace chemical fertilizers and pesticides for sustainable cauliflower cultivation.

Thus, the selected isolate (MK₂) with optimum doses of N and P chemical fertilizers has good prospects to be used as integrated nutrient module not only for enhanced yield but also to sustain soil health under mid hill conditions of Himachal Pradesh.

REFERENCES

- Bakker, A. W. and Schippers, B. 1987.** Microbial cyanide production in the rhizosphere to potato yield reduction and *Pseudomonas* spp. mediated plant growth stimulation. *Soil Biology and Biochemistry*. **19**: 451-457.
- Bertrand, H., Plassard, C., Pinochet, X., Touraine, B., Normand, P. and Cleyet-marcel, J. C. 2000.** Stimulation of the ionic transport system in *Brassica napus* by a plant growth promoting rhizobacterium (*Achromobacter* spp.) *Canadian J. Microbiology*. **46**: 229-236.
- Bowen, G. D. and Rovira, A. D. 1999.** The rhizosphere and its management to improve plant growth. *Advances in Agronomy*. **66**: 1-102.
- Gomez, K. A. and Gomez, A. A. 1984.** *Statistical procedure for agricultural research* 11nd edition. J. Wiley and sons, New York. Inc. p. 680.
- Gorden and Paleg, L. G. 1957.** Quantitative measurement of IAA. *Plant Physiology*. **10**: 37-38.
- Hatwar, G. P., Gondane, S. U. Urkude, S. M. and Gahukar, O. V. 2003.** Effect of micronutrients on growth and yield of chilli. *Soils and Crops* **13**: 123-125.
- Hazara, P., Maity, T. K. and Mandal, A. R. 1987.** Effect of foliar application of micronutrients on growth and yield of okra (*Abelmoschus esculentum* L.). *Progressive Horticulture*. **19**: 219-222.
- Jackson, M. L. 1973.** *Soil chemical analysis* Prentice Hall of India Ltd. New Delhi. pp. 219-221.
- Jensen, E. S. 1987.** Inoculation of pea by application of *Rhizobium* in planting furrow. *Plant and Soil*. **97**: 63-70.
- Kloepper, J. W., Ryu, C. M. and Zhang, S. 2004.** Induced systemic resistance and promotion of plant growth by *Bacillus* spp. *Phytopathology*. **94**: 1259-1266.
- Merwin, H. D. and Peech, M. 1951.** Exchange ability of soil potassium in the sand, silt and clay fractions as influenced by the nature and complementary exchangeable cations. *Soil Science American Proceedings*. **15**: 125-128.
- Olsen S. R. C. V., Cole, D. S. Watanabe, D. S. and Dean, L. A. 1954.** Estimation of available phosphorus in soils by extraction with sodium bicarbonate. *USDS Circular*. p. 939.
- Pikovskaya, R. I. 1948.** Mobilization of phosphorus in soil in connection with the vital activity of some microbial species. *Mikrobiologiya*. **17**: 362-370.
- Schwyn, B. and Neilands, J. B. 1987.** Universal chemical analysis for the detection and determination of siderophores. *Analytical Biochemistry*. **160**: 47-56.
- Sharma, R. C. 1986.** Nitrogen management of potatoes in the presence of farmyard manure and P K fertilizers on acid hill soils of Shimla. *J. Agricultural Sciences, Cambridge*. **107**: 15-19.
- Subba Rao, N. S. 1999.** *Soil Microorganism and Plant Growth*, Oxford and IBH publishing Co, New Delhi. 252 p.
- Subbiah, B. V. and Asija, G. L. 1956.** A rapid procedure for the estimation of the available nitrogen in soils. *Current Science*. **25**: 259-60.
- Vincent, J. M. 1947.** Distortion of fungal hyphae in the presence of certain inhibitors. *Nature* **150**: 850.
- www.nhb.gov.in (2010).