# BIOEFFICACY OF *METARHIZIUM ANISOPLIAE* WITH NEEM DERIVATIVES AGAINST *HELICOVERPA ARMIGERA* INFESTING CHICKPEA

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#### **KEYWORDS**

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# **ABSTRACT**

The field experiments were carried out during the year 2013-2014 at the experimental field of JNKVV, Jabalpur to evaluate the efficacy of *M. anisopliae* and neem derivatives alone and in combination against *Helicoverpa armigera* infesting chickpea. All the neem derivatives and microbials proved their superiority over the control in reducing the pod damage and increasing the grain yield. On the basis of effectiveness of different treatments pod damage by pod borer and grain yield, revealed that *Metarhizium anisopliae* + Neem soap @ 1 x 10<sup>4</sup> spores/ml +0.5% (w/v), was the most effective treatment. On the basis of cost benefit ratio the same combination gave maximum cost benefit ratio followed by *Metarhizium anisopliae* @ 1 x 10<sup>8</sup> spores/ml (1:8.39 and 1:5.40, respectively).

# **INTRODUCTION**

Production of chickpea in our country is low and one of the major reasons is the losses caused by several pests and diseases, both in field and in storage. It is attacked by number of insect pests, among them, the gram pod borer *Helicoverpa armigera* Hubner (Lepidoptera: Noctuidae) is the most devastating insect pest inflicting serious damage to the crop (Sithanantham et al. 1984).

*H. armigera* is a polyphagous nature, the gram pod borer is also known as American cotton boll worm, corn ear worm, tomato fruit borer, tobacco bud worm, carnation worm, etc. It has been recorded feeding on 182 cultivated and uncultivated plant species belonging to 47 families and also seriously damages to 56 plant species (Pawar et al. 1986; Regupathy et al.1997). Hence, farmers are compelled to use chemical insecticides in order to cultivate lucratively, as traditional and cultural practices alone cannot not give satisfactory control over the pest menace (Vanlaldiki et al. 2013). This has necessitated the use of alternative eco-friendly insecticides to sustain the management of *Helicoverpa armigera*.

The growing awareness of pesticidal hazards on human being and environment is the real fact for the development of sustainable eco-friendly pest management program such as "Biological control". Biopesticides are inherently less harmful than conventional pesticides. Biopesticides are clearly and mostly target specific in contrast to broad spectrum conventional chemical pesticides that kill almost all living

organism. Biopesticides are effective in very small quantities and are often quickly biodegradable. (Bhushan et al. 2012) Biological control includes use of bioagents and microbial derived from fungi, bacteria and virus to control disease and insect pests. The most considerable fungal species are Metarhizium spp., Beauveria spp., Nomuraea rileyi, Verticillium lecanii and Hirsutella spp (Alves and Lopes 2008). Metarhizium causes a disease known as 'green muscardine' in insect hosts because of the green colour of its conidial cells. With the current thrust on Sustainable agriculture and organic farming the use of M. anisopliae and neem derivatives have been reported efficacious against pod borer however, information are scanty and combined effect of Metarhizium anisopliae with neem derivatives not known(Murray et al. 2000; Kavitha et al. 2009). Keeping the fact in background the present investigation are undertaken under field condition on bioefficacy of Metarhizium anisopliae with neem derivatives against Helicoverpa armigera infesting chickpea.

# MATERIALS AND METHODS

Field experiment was conducted at the experimental field of Department of Entomology, Adhartal, JNKVV, Jabalpur (M.P.) during *Rabi* 2013-14, under randomized block design. Chickpea variety JG-12 was sown on 12<sup>th</sup> December, 2013 in a plot size of 10 x 3.3 m<sup>2</sup> with a spacing of 30 x 10 cm. Other agronomic practices were followed as per local recommendation. There were eight treatments comprising of

Metarhizium anisopliae (1x108 spores/ml) and neem derivatives (Neem oil, NSKE, Neem soap) alone and combination of M. anisopliae with neem derivatives including control (Table 1). Neem seed kernel and crude neem oil were purchased from local market. Neem soap was purchased from Indian Institute of Horticulture Research, Hessarghatta Lake Post, Bengaluru. Preparation of NSKE was followed as per standard method (PURI et al., 1998) i.e. 5 kg of dried neem seeds were taken and cleaned it thoroughly. A day before schedule spraying, it was powdered by grinding and soaked overnight in 10 litres of distilled water and stirred with wooden plank till solution became milky white. It was then filtered through double layer of muslin cloth and made its volume to 100 litre by adding distilled water. To it 200 ml of sticker was added and the resultant solution was sprayed to cover upper as well as lower foliar portions of the crop. Treatment wise application of biochemical's were made at pod formation stage (90 DAS) of crop at ETL (i.e. e 2 larvae/mrl) level of H. armigera. Observation were recorded on H. armigera larvae on randomly selected 10 sites (1 meter row length/site) on 3,7,10 days after application (DAA). Larval count was also taken at 24 hrs before initiation of treatments. Pods of ten plants per plot were counted and per cent pod damage were calculated based on healthy and bored pods. Grain yield/plot was also assessed after harvesting and C:B ratio was calculated based on cost of application, market price of biopesticides and chickpea.

All the data were subjected to statistical analysis after appropriate transformation as suggested by Snedecor GW and Cochran WG, 1967.

# **RESULTS AND DISCUSSION**

# Effect of biopesticides on larval population of *H. armigera* Pre- treatment

Differences in the *Helicoverpa armigera* mean larval population per meter row length (mrl) among different plots were not significant, indicating more or less uniform distribution of the pest but it was above the ETL (2.40 L/mrl) in the experimental field.

# Three days after application

The mean *Helicoverpa armigera* larval population per mrl among different treatments ranged from 2.57 to 3.23 but without any significant difference indicating no immediate effect of treatments on larval population.

# Seven days after application

As revealed from the Table 1. at seven days after treatment, all the treatments significantly reduced the larval population as compared to control (2.80 larvae / mrl). Among the treatments, M. anisopliae + Neem soap @  $1 \times 10^4$  spores/ml + 0.5% (w/v) was found to be the most effective as it recorded lowest larval population (0.80 larvae/mrl) this was followed by Neem seed kernel extract @ 5% (w/v) (0.87 larvae/mrl), but they did not differ significantly from each other. The next effective treatment was M. anisopliae + Neem oil @  $1 \times 10^4$  spores/ml + 2.5% (v/v) (0.97 larvae/mrl), followed by M. anisopliae @  $1 \times 10^8$  spores/ml (1.07 larvae/mrl), M. anisopliae + NSKE @  $1 \times 10^4$  spores/ml + 2.5% (w/v) (1.10 larvae/mrl), Neem soap @ 1% (w/v)

Table 1: Bioefficacy of *Metarhizium anisoplia*e and neem derivatives alone and their combinations on *Helicoverpa armigera* and chickpea grain yield

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Treatments Dose Pre treatment Days after spraying Pre treatment Days after spraying Pre treatment Days after spraying    Netachizium anisopliae											
11 / ha (1 × 10 <sup>8</sup> spores/ml)min. 2.40(1.55) 2.50(1.58) 1.07(1.03) 0.94(0.97) 2.27(1.50) 2.33(1.53) 0.87(0.93) 0.60(0.77) 2.53(1.59) 2.63(1.59) 1.07(1.08) 1.10(1.05) 1.06(0.77) 2.53(1.59) 2.63(1.59) 2.63(1.59) 1.17(1.08) 1.10(1.05) 1.06(0.77) 1.06 spores /ml + 2.5% (w/v) 2.47(1.57) 2.53(1.59) 1.10(1.05) 0.97(0.98) 1.10(1.05) 0.97(0.98) 1.10(1.05) 0.97(0.98) 1.10(1.05) 0.97(0.98) 1.10(1.05) 0.97(0.98) 0.57(0.74) 2.23 (1.49) 2.27 (1.51) 0.80 (0.89) 0.57 (0.74) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.75 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.75 (1.60) 2.73 (1.60)		Treatments	Dose	-	Mean <i>H. armi</i> g Pre treatment	gera larval popu Days after spr	ulation / mrl* aying			Pod damage (%)**	Grain yield (kg/ha)
11 / ha (1 × 10 <sup>8</sup> spores/ml)min. 2.40(1.55) 2.50(1.58) 1.07(1.03) 0.94(0.97) 2.27(1.50) 2.33(1.53) 0.87(0.93) 0.60(0.77) 2.57(1.50) 2.33(1.53) 0.87(0.93) 0.60(0.77) 2.53(1.59) 2.63(1.62) 1.17(1.08) 1.10(1.05) 1.03(1.01) 1.1 × 10 <sup>4</sup> spores /ml + 2.5% (w/v) 2.47(1.57) 2.57(1.60) 1.13(1.06) 1.03(1.01) 1.1 × 10 <sup>4</sup> spores /ml + 2.5% (w/v) 2.37(1.54) 2.43(1.56) 0.97(0.98) 0.67(0.81) 1.10(1.05) 0.97(0.98) 0.57(0.74) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69)						3	7	10	Mean		
5KE) 5% (w/v) 5% (w/v) 5% (w/v) 5% (w/v) 1% (w/v) 1% (w/v) 1	-	Metarhizium anisopliae	11 / ha $(1 \times 10^8 \text{spores/m})$	l)min.	2.40(1.55)	2.50(1.58)	1.07(1.03)	0.94(0.97)	1.50(1.23)	12.00(20.23)	762.96
5% (v/v) 1% (w/v) 1% (w/v) 2.53(1.59) 2.53(1.62) 1% (w/v) 2.50(1.58) 2.57(1.60) 1.17(1.08) 1.10(1.05) 1.3(1.06) 1.3(1.06) 1.3(1.01) 1× 10 <sup>4</sup> spores /ml + 2.5% (w/v) 2.47(1.57) 1× 10 <sup>4</sup> spores /ml + 0.5 % (w/v) 2.37(1.54) 1× 10 <sup>4</sup> spores /ml + 0.5 % (w/v) 2.27 (1.60) 2.27 (1.51) 2.53 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.8F ± 0.02 0.03 0.03 0.05 0.16	7	Neem seed kernel extract (NSKE)	5% (w/v)		2.27(1.50)	2.33(1.53)	0.87(0.93)	0.60(0.77)	1.27(1.13)	7.67(16.03)	842.59
1% (w/v) 1% (w/v) 2.50(1.58) 2.57(1.60) 1.13(1.06) 1.03(1.01) 1× 10 <sup>4</sup> spores /ml + 2.5% (w/v) 2.47(1.57) 2.53(1.59) 1.10(1.05) 0.97(0.98) 1×10 <sup>4</sup> spores /ml + 2.5% (w/v) 2.37(1.54) 2.43(1.56) 0.97(0.98) 0.67(0.81) 1×10 <sup>4</sup> spores /ml + 0.5 % (w/v) 2.23 (1.49) 2.27 (1.51) 2.67 (1.60) 2.73 (1.65) 2.87 (1.69	3	Neem oil	5% (v/v)		2.53(1.59)	2.63(1.62)	1.17(1.08)	1.10(1.05)	1.63(1.28)	22.86(28.54)	556.48
1× 10 <sup>4</sup> spores /ml + 2.5% (w/v) 2.47(1.57) 2.53(1.59) 1.10(1.05) 0.97(0.98) 1× 10 <sup>4</sup> spores /ml + 2.5% (w/v) 2.37(1.54) 2.43(1.56) 0.97(0.98) 0.67(0.81) 1× 10 <sup>4</sup> spores /ml + 0.5% (w/v) 2.23 (1.49) 2.27 (1.51) 0.80 (0.89) 0.57 (0.74) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69)	4	Neem soap	1% (w/v)		2.50(1.58)	2.57(1.60)	1.13(1.06)	1.03(1.01)	1.58(1.26)	19.84(26.41)	568.52
1× 10 <sup>4</sup> spores /ml + 2.5% (v/v) 2.37(1.54) 2.43(1.56) 0.97(0.98) 0.67(0.81) 1×10 <sup>4</sup> spores /ml + 0.5 % (w/v) 2.23 (1.49) 2.27 (1.51) 0.80 (0.89) 0.57 (0.74) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.57 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.87	2	M. anisopliae + NSKE			2.47(1.57)	2.53(1.59)	1.10(1.05)	0.97(0.98)	1.53(1.24)	16.83(24.20)	703.70
1×10 <sup>4</sup> spores /ml + 0.5 % (w/v) 2.23 (1.49) 2.27 (1.51) 0.80 (0.89) 0.57 (0.74) 7.25 (1.60) 2.73 (1.65) 2.80 (1.67) 2.87 (1.69) 2.87 (1.6	9	M. anisopliae + Neem oil			2.37(1.54)	2.43(1.56)	0.97(0.98)	0.67(0.81)	1.36(1.16)	9.32(17.72)	812.04
SEm ± 0.02 0.03 0.03 0.05 0.05 CD at 5% NS NS 0.09 0.16 0.16 0.16 0.16 0.16 0.16 0.16 0.16	_	M. anisopliae + Neem soap			2.23 (1.49)	2.27 (1.51)	0.80 (0.89)	0.57 (0.74)	1.21 (1.10)	6.70 (14.96)	890.74
0.02 0.03 0.05 0.05 0.05 NS NS 0.09 0.16 0.0	ω	Control			2.57 (1.60)	2.73 (1.65)	2.80 (1.67)	2.87 (1.69)	2.80 (1.67)	25.95 (30.61)	523.15
NS NS 0.09 0.16			SEm ±	J	0.02	0.03	0.03	0.05	0.02	0.41	8.33
			CD at 5%	_	۲S	NS	60.0	0.16	0.05	1.25	25.28

= meter row length, \* Figures in parentheses are (v x + 0.5) square root transformed values, \*\* Figures in parentheses are arcsin transformed values; NS = Non significant

(1.13 larvae/mrl) and Neem oil @ 5% (v/v) (1.17 larvae/mrl) (T,) but they did not differ significantly from each other.

### Ten days after application

At ten days after treatment, All the treatments significantly reduced the larval population as compared to control (2.87 larvae / mrl). Among the treatments, M. anisopliae + Neem soap @  $1 \times 10^4$  spores/ml + 0.5% (w/v) was found to be most effective as it recorded lowest larval population (0.57 larvae/mrl), this was followed by Neem seed kernel extract @ 5% (w/v) (0.60 larvae/mrl) and M. anisopliae + Neem oil @  $1 \times 10^4$  spores/ml + 2.5% (v/v) (0.67 larvae/mrl), but all the three treatments were at par with each other. The next effective treatment was M. anisopliae @  $1 \times 10^8$  spores/ml (0.94 larvae/mrl), followed by M. anisopliae + NSKE @  $1 \times 10^4$  spores/ml + 2.5% (w/v) (0.97 larvae/mrl), Neem soap @ 1% (v/v) (1.03 larvae/mrl), Neem oil @ 5% (v/v) (1.10 larvae/mrl), but they did not differ significantly from each other .

#### Overall mean

On the basis of overall mean, the differences in the mean larval population among different treatments were significant. All the treatments significantly reduced the larval population as compared to control (2.80 larvae / mrl). Among the treatments, M. anisopliae + Neem soap @ 1×10<sup>4</sup> spores/ ml + 0.5% (w/v) was found to be most effective as it recorded lowest larval population (1.21 larvae/mrl). The next effective treatment was Neem seed kernel extract @ 5% (w/v) (1.27) larvae/mrl), followed by M. anisopliae + Neem oil @  $1 \times 10^4$ spores/ml + 2.5 % (v/v) (1.36 larvae/mrl) but they differ significantly from each other. The next effective treatment was M. anisopliae @ 1×108 spores/ml (1.50 larvae/mrl), followed by M. anisopliae + NSKE @  $1 \times 10^4$  spores/ml + 2.5 % (w/v) (1.53 larvae/mrl), but both of them were at par with each other. The least effective treatments were Neem soap @ 1 % (v/v) (1.58 larvae/mrl) and Neem oil @ 5% (v/v) (1.63 larvae/mrl) and were at par with each other.

# Effect of biopesticides on pod damage and grain yield of chickpea

# Chickpea Pod damage

Based on per cent infestation at harvest, chickpea pod damage caused by H. armigera was in the range of 6.70% to 22.86% in the biopesticides treatments as against 25.95% in untreated control. The lowest pod damage (6.70%) was due to combination treatment of M.  $anisopliae + Neem soap @ 1 \times 10^4 \text{ spores/ml} + 0.5\%$  (w/v) this was followed by Neem seed kernel extract @ 5% (w/v) recording (7.67%) pod damage but both were at par with each other. The treatment of M. anisopliae combined with Neem oil with their half doses also recorded lower (9.32%) pod damage. The higher pod damage in the range of 12 to 19.84% were recorded in M. anisopliae @  $1 \times 10^8$  spores/ml, M.  $anisopliae + NSKE @ <math>1 \times 10^4$  spores/ml + 2.5% (w/v), Neem soap @ 1% (w/v). Application of Neem oil @ 5% (v/v) was least effective with high pod damage of 22.86%.

#### Chickpea grain yield

The yield of net plot area of each treatment was recorded and converted into kg/ha. All the treatments registered significantly higher grain yields as compared to the control (523.15 kg/ha).

The highest grain yield was recorded in *M. anisopliae* + Neem soap @  $1 \times 10^4$  spores/ml+0.5% (w/v) treated plots (890.74 kg/ha) which was significantly superior than the other treatments. Subsequent higher grain yield was recorded with Neem seed kernel extract @ 5% (w/v) (842.59 kg/ha) followed by *M. anisopliae* + Neem oil @  $1 \times 10^4$  spores/ml+ 2.5% (v/v) (812.04 kg/ha), but they differed significantly from each other. The next effective treatment were *M. anisopliae* @  $1 \times 10^8$  spores/ml (762.96 kg/ha), followed by *M. anisopliae* + NSKE @  $1 \times 10^4$  spores/ml + 2.5% (w/v) (703.70 kg/ha), but they differed significantly from each other. The least effective treatments were Neem soap @ 1% (w/v) (568.52 kg/ha) and Neem oil @ 5% (v/v) (556.48 kg/ha) and they were at par with each other.

All the neem derivatives and *M. anisopliae* alone and their combination proved their superiority over control in reducing the pest population and pod damage and in increasing the grain yield. Several workers have also reported similar findings, that application of neem derivatives and *M. anisopliae* effectively reduced the damage due to pod borer with increased grain yield than control (Katole *et al.*, 2000; Kumar and Chowdhary 2004; Nahar *et al.* 2004; Kulkarni *et al.*, 2005; Singh and Yadav 2005; Gundannavar *et al.* 2007; Haque and Ghosh 2007; Ali *et al.*, 2008; Kale and Men 2008; Rijal *et al.*, 2008; 2008a; Wakil *et al.* 2008; Bhushan *et al.*, 2011; Moorthy *et al.*, 2011; Rao *et al.* 2011; Wakil and Ghaznafar 2011; Singh *et al.*, 2012; Ahmad *et al.*, 2014 and Chandel *et al.*, 2014).

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